

1 **Encapsulation of bioactive compounds for the formulation of functional animal feeds:**
2 **the biofortification of derivate foods**

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21 **Abstract**

22 Microencapsulation is a technology that has increasing application in animal nutrition. This
23 technology provides the possibility to protect sensitive compounds through the feed process
24 and the storage conditions. Furthermore, the application of this technique affords the
25 possibility to increase the bioavailability of compounds and to mask the unpalatable
26 compounds taste. Different are the compounds to consider for microencapsulation include
27 certain vitamins, amino acids, fatty acids, organic acids and live microorganisms.
28 Understanding the interdependence between bioactive compound, coating, technology and
29 environment is the key to successful application of microencapsulation also in animal
30 nutrition. In this review a focus on the use of microencapsulated bioactive compounds for the
31 feed supplementation and the related effects on the derivate animal product is given.

32 **Keywords:** animal-based products; biofortification; bioactive compounds; feed
33 supplementation; encapsulation.

34 **1. Introduction**

35 There is a need to overcome the pressing challenge of meeting the nutritional demands of the
36 growing population which is estimated to reach 9.7 billion in 2050 (United Nation, 2019). For
37 example, over two billion people worldwide are micronutrient deficient with poorest regions
38 been the most affected. Attractive approaches to tackle micronutrient deficiency include a
39 targeted integration of nutrients directly into food (fortification) and increasing the
40 concentration of the active compounds in edible crops (biofortification) (WHO, 2020).
41 Micronutrients could also be administrated as medical supplements. The main disadvantage of
42 this approach lies in the limited availability of particular compounds and shelf-life stability.
43 Also, it has been suggested that micronutrients function better when consumed as part of a

44 balanced diet, rather than as pills (Hunter, 2011). When using biofortification, the poor
45 bioavailability of certain nutrients of plant-based foods must be considered (Willer and
46 Aldridge, 2020) whereas the same nutrients in animal-based foods, like milk or meat, are
47 generally easily absorbed by the human body (O’Keeffe, 2020). On this ground, the
48 possibility to deliver micronutrients through animal products offers interesting additional
49 benefits. Different studies have shown that feed fortified with nutrients can be used as dietary
50 supplements in animal nutrition which contribute to improve the productivity of animals and
51 the quality of the derived products (Grilli et al., 2013; Hu et al., 2015; Kim et al., 2020;
52 Konkol and Wojnarowski, 2018; Natsir et al., 2019; Stamilla et al., 2020; Tao et al., 2020).
53 Taking a holistic approach, micronutrients could be considered as part of a network of
54 bioactive molecules within the food and the organism and as such, the same approach could
55 be applied to other bioactive compounds.

56 Bioactive compounds or ingredients are substances which in small amounts have the ability to
57 provide health benefits. These are usually natural occurring substances with specific
58 properties that may preserve or improve the animal health condition and, at the same time,
59 contribute to increasing their concentration in the products obtained from them (Korczynski et
60 al., 2017). However, some of these bioactive compounds such as polyunsaturated fatty acids,
61 essential oils, polyphenols and vitamins are prone to oxidation whereas plant extracts and
62 essential oils are often difficult to process in addition to their unpleasant taste. Encapsulation
63 has been proven as an efficient tool for improving functions and properties of bioactive
64 ingredients and allowing the incorporation of even sensitive ingredients into products. Indeed,
65 encapsulation allows a liquid, gaseous or solid substance to be protected into an envelope or
66 coat material and has several other benefits. These include control and target release, taste
67 masking, prevention of interaction with other incompatible ones in the product matrix as well
68 as protection of bioactive compounds against oxidation promoters, such as air, light and

69 metals (Piva et al., 2007; Tolve et al., 2016; Tolve et al., 2020). On the other hand,
70 consumers' demand for natural, high-quality animal products produced with no negative
71 effect or reduced environmental impact while protecting animal welfare reinforces the need
72 for the development of more efficient and sustainable production methods. This article aims
73 to review bioactive compounds used in animal feed formulations and different approaches for
74 encapsulating them alongside potential effect on the derived products and animal welfare. In
75 this review, the term “encapsulation” has been used if no size definition was given about the
76 particles whereas “microencapsulation” was used for specified size ranged from 1 μm to 1
77 mm.

78 **2. Encapsulation in animal feed and subsequent products biofortification**

79 Although encapsulation is also used to improve formulation and safety of animal feed, this
80 section focuses principally on its applications to improve product bioavailability, deliver
81 nutrients and additives to the digestive tract or specific sites for enhancing animal
82 performance and the quality of the resulting products. Animal farming industry is searching
83 for new ways to improve the overall quality of animal-based products such as meat, milk and
84 eggs. One of the recently developed approaches is the “biofortification” which utilizes animal
85 feed supplemented with encapsulated bioactive compounds (Konkol and Wojnarowski, 2018).
86 The choice of the encapsulation formulation and manufacturing processes depends upon the
87 type of animal to be fed, the chemistry of the bioactive compound and the purpose of its
88 encapsulation. As example, lipid based-shell material was proved effective to provide
89 sustained release of organic acids along the swine intestine (Piva et al., 2007) thereby
90 allowing to overcome any issue related to their rapid absorption and metabolism in the
91 duodenum. It has also been demonstrated that lipid base-shell protects nutrients from ruminal
92 degradation (Grilli et al., 2013), but may not be useful if the purpose of the encapsulation is to

93 mask unpleasant taste such as in the case of tannins applications (Adejoro et al., 2019a; Tolve
94 et al., 2021). In Figure 1 a schematic diagram showing the protecting effect and the site-
95 specific release site of the encapsulated bioactive compounds has been reported. Indeed,
96 different approaches have been employed for the encapsulation of bioactive compounds
97 intended for animal feed as highlighted in Tables 1-7. These Tables also show diverse
98 encapsulated commercial blends. Encapsulation methods used in animal feed have been
99 reviewed by Temiz and Öztürk (2018) and each method presents different advantages and
100 drawbacks. Coating materials include proteins (corn zein, wheat gluten, casein, gelatin, soy
101 and whey proteins), polysaccharides (cellulose and other plant-based polymers), synthetic
102 polymers (poly-2-vinylpyridine-co-styrene and polymethacrylate polymers) (Bribiesca et al.,
103 2017; Temiz and Öztürk, 2018) and composites materials whereas the particle size varies
104 from nano size to millimeter size. For example, particles diameter up to 6 mm has been
105 suggested as optimum size of an encapsulated product for cattle (Sýkora et al., 2007). Based
106 on the general values of rumen pH, 5.6 - 7.0, coating materials with ability to degrade a pH
107 below 5 could potentially survive rumen degradation and thus candidate for release in small
108 the intestine formulations if they survive microbial degradation. However, the mechanical
109 stability of the shell material should also be considered. Bioactive compounds encapsulated
110 for animal feed include aroma and fragrances, essential oils, omega fatty acids, plant extracts,
111 enzymes, prebiotic and micro-organisms, vitamins, amino acids and minerals.

112 *2.1 Encapsulation of minerals*

113 Micro- and macro elements deficits have been observed for many years in human concerns
114 about two billion people (Chojnacka et al., 2011). Generally, the animal feed supplementation
115 with minerals allows for increasing animal production however, for specific minerals, the
116 supplementation could also cause the increase of certain minerals in animal-based foods. This

117 is usually expressed in terms of the level of incorporation of an element into the final animal-
118 based product (Słupczyńska et al., 2014). In view of this, several authors (Bribiesca et al.,
119 2017; Grilli et al., 2013) have evaluated the possibility to enhance the nutritional quality of
120 animal-based product through the feed supplementation with encapsulated minerals (Table 1).
121 Selenium is a trace element crucial for homeostasis and oxidative status in animals and
122 essential component of about 25 selenoproteins in the body, including enzymes involved in
123 cellular antioxidant protection and it is essential for thyroid hormone metabolism. Suboptimal
124 Se status in humans has been associated with an increased risk of cancer, HIV/AIDS and
125 other viral diseases (measles, hepatitis, influenza) including endemic cardiomyopathy namely
126 Keshan Disease (Bribiesca et al., 2017). The recommended daily dosage (RDA) of Se per day
127 for normal adults is 60-75 µg in Europe. However, there is an estimated 30-100 million
128 people worldwide with Se deficiency which is mainly due to low Se concentrations in
129 frequently consumed foods (Thavarajah et al., 2011). On the other hand, selenium
130 supplements are partially transformed in insoluble form by ruminal microorganisms and this
131 process decreases its bioavailability in gastrointestinal system (Romero-Pérez et al., 2010).
132 Therefore, Romero-Pérez et al. (2010) investigated the possibility of protecting Se from
133 ruminal microbial complexation by encapsulation technology. The authors prepared
134 microencapsulated sodium selenite by emulsion-evaporation and by nanoprecipitation
135 methods and, by *in vitro* digestion study, found that the release of selenium from
136 nanoparticles was higher at acid pH (less than 4) when compared to that at pH 6. This result
137 indicates that a better bioavailability and absorption of Se would occur in the small intestine.
138 Eudragit® RL and RS were used as shell materials because of their insolubility in aqueous
139 environment with good permeability related to the presence of quaternary ammonium groups
140 in their structure (Bribiesca et al., 2017). Since Eudragit® RL has high permeability to
141 aqueous fluid than Eudragit® RS, the blend of the two polymers is usually used to tune the

142 release kinetics to match the desired release profile. The *in vivo* study conducted by Grilli et
143 al. (2013) confirmed the possibility to increase the bioavailability of Se via encapsulation by
144 demonstrating an increase in Se concentration in milk after supplementing cows feed with
145 microencapsulated Se in hydrogenated fat. Because the milk from yeast-supplemented feed
146 did not differ from that of the control, the study also indicated that Se microencapsulated in
147 lipid-based formulation of nanoparticles could be a promising route for protecting minerals
148 from rumen degradability or complexation. Similarly, Bribiesca et al. (2017) reported an
149 increase in both selenium and zinc content of meat from lambs fed with Se and Zn
150 nanoparticles with Eudragit RL as coating material when compared to feed supplemented
151 with nonencapsulated Se and Zn. The increased concentration of Se and Zn in meat was
152 attributed to a better bioavailability of Se and Zn when they were supplemented in the form of
153 nanoparticles as compared to sodium selenite and zinc carbonate. Moreover, the increase in
154 Se concentration in meat could enhance its antioxidant properties preventing lipid
155 peroxidation and consequently improving meat quality. The increase in Zn in the meat would
156 also contribute to tackle deficiency induced clinical symptoms in children, such as
157 undercurrent infections, body growth retardation, anorexia, dermatitis, impaired reproductive
158 performance, skeletal abnormalities and diarrhea, alopecia, scarring defects and behavior
159 disorders (Prasad, 2013).

160 *2.2 Encapsulation of vitamins*

161 The most common reason to encapsulate vitamins is their shelf-life extension, protection from
162 damage during processing (e.g., heat and steam in the pelleting or extrusion process), storage
163 and transport as well as delivery to the small intestine because of their rapid degradation in
164 the rumen by ruminal microbes (Table 2). Albright and Kowarski (1994) demonstrated an
165 improved stability of vitamin A encapsulated into a lipid bilayer containing lecithin,

166 cholesterol and functionalized stearyls. Their hypothesis was that the lipid bilayer would
167 physically separate vitamin A from the mineral components and reduce or inhibit hydrolytic
168 degradation through water capture in a form of liposome when it penetrates the bilayer,
169 thereby limiting water availability to interact with vitamin A and initiate its degradation.

170 Vitamins that are extensively metabolized in the rumen include choline and vitamin E
171 (Santschi et. al., 2005). Most research with encapsulated vitamins has focused on choline
172 however the use of rumen protected choline is generally related to the production of dairy
173 cows. For example, rumen protected choline has been reported to be useful in increasing milk
174 production during cow's first month of lactation and also to increase the milk choline
175 concentration (Pinotti et al., 2003). This effect was confirmed by by Arshad et al. (2020) who
176 through a meta-analysis showed an increased yield of milk, energy-corrected milk, fat,
177 protein, and a tendency of a reduction of the risk of retained placenta and mastitis for parous
178 dairy cows under rumen-protected choline supplementation. Similarly, Hu et al. (2015)
179 evaluated the possibility of using 20 mg/kg of a commercial vitamin E encapsulated with
180 sodium octenyl succinate in broiler chicken's diet for 42 days and found that vitamin E in
181 both free and encapsulated form in the feed improved breast meat quality in broilers with a
182 lower abdominal fat percentage. However, encapsulated-based feed resulted in better broiler
183 growth performance with higher body gain when compared free vitamin E containing feed
184 whereas higher pH and antioxidant enzyme activity was reported for boiler fed with vitamin
185 when compared to those on basal diets without vitamin E. Since vitamin E easily oxidized,
186 has poor water solubility and low bioavailability (Sagalowicz and Leser 2010), the
187 encapsulated form effectively increased its stability and overall bio-accessibility. Vitamin E is
188 an effective antioxidant that can protect cells from oxidative damage and numerous nutritional
189 and physiological studies have shown that its supplementation is beneficial for growth
190 performance in animals. In terms of biofortification, even more interesting are the results

191 obtained by Lee et al. (2007) who assessed the effect of feeding with a rumen protected α -
192 tocopheryl-acetate and sunflower oil, made using sodium caseinate on α -tocopherol
193 concentrations of muscle in lambs over the 10-week. Results showed that lambs fed with
194 encapsulated vitamin E and sunflower oil had higher α -tocopherol concentrations in their
195 muscle when compared to those fed with non-encapsulated vitamin E. It is worth mentioning
196 that no published work was found regarding the encapsulation of vitamin D for animal feed
197 even though research increasingly suggests its role in stimulating immunity and protecting
198 against viral and bacterial infection, its contribution to cardiovascular physiology, cells
199 proliferation and differentiation alongside its principal to modulate calcium and phosphorus
200 homeostasis. Fish meals and oils are good source of vitamin D₃ with 0.01 to 0.18 mg/kg
201 (meals) and 0.03 to 3.9 mg/kg (oils) depending on the species and the season (EFSA, 2017).
202 The increasing use of plant-based ingredients in aquaculture feeds has created more need for
203 vitamin D fish farm feed supplementation as these formulations is expected to contain only
204 25–33% of the vitamin D₃ content of the conventional marine-based formulations (EFSA,
205 2017). Since fish has no ability to endogenous synthesize vitamin D₃ like other vertebrates
206 such mammalian species and birds they have to be supplied through food (EFSA, 2017).
207 Estimated vitamin D₃ transfer rate from feed to fish to be in the range of 0.1–0.15, thus given
208 vitamin D₃ content in fish flesh ranging from 150 up to 220 μ g/kg for a feed containing
209 1.5 mg/kg of vitamin D₃. This supplementation could help addressing low dietary vitamin D
210 intake and consequently lowering the risk of vitamin D deficiency (Cashman, 2020). In this
211 field, encapsulation could be useful to increase the shelf-life of vitamin D in formulations and
212 for safety because vitamin D₃ is considered as irritant to skin and eyes as well as a dermal
213 sensitizer (EFSA, 2014). Nowadays, it is possible to find on the market only calcifediol
214 monohydrate containing nanoparticles used to protect the vitamin D analogue from oxidation
215 (EFSA, 2021).

216 2.3 Encapsulation of amino acids and urea

217 The main source of amino acids for animal is the diet, though ruminants also produce them
218 via their gut microflora during fermentative digestion. Amino acids are absorbed from the
219 small intestine as free amino acids or as di- and tripeptides and released into the blood mostly
220 as free amino acids but some peptides (Fuller, 2004). Protein and amino acid in the rumen are
221 also degraded by rumen microbes leading to dietary protein conversion into ammonia, and
222 excretion as urea. Sulphur containing amino acids are precursors for the others amino acids
223 for ruminants (Gaillard et al., 2018). In this contest, methionine and lysine are limiting for
224 milk and milk protein synthesis. In addition, histidine has been identified as first limiting
225 amino acid for milk protein production when cows are fed with grass silage and barley or oats
226 (Vanhatalo et al., 1999). Different approaches have been used to achieve delivery of amino
227 acids at the absorption sites (Table 3). The first paper about the supplementation of animal
228 feed with rumen protected amino acids was published in 1970 by Broderick et al. (1970), who
229 used an encapsulated DL-methionine with kaolin and tristearin. The authors fed lactating
230 cows with encapsulated methionine at 5, 15, and 45 g per day and observed no effect on yield
231 of the milk or milk components, but instead an increase in plasma methionine and
232 methionine/valine ratios in all cows at 15 g supplementation level. Although these results
233 were confirmed in the same years by Williams et al. (1970), it could be postulated that the
234 absence of improvement in milk or milk protein production using encapsulated methionine
235 could have been due, to some extent, to the use of only methionine for the supplementation.
236 Indeed, Rogers et al. (1987) demonstrated that rumen-protected methionine and lysine
237 increased production of milk protein, whereas Sevi et al. (1998) reported a modification of the
238 milk fatty acid composition in ewes with an increase in the ratio between long-chain and short
239 chain fatty acids and the ratio of unsaturated/saturated fatty acids which is desirable from
240 consumers' health perspective. Socha et al. (2005) obtained an enhanced milk quality when

241 cows were fed with the rumen protected methionine and lysine as more true protein (1306 vs.
242 1221 g/d), and fat (1632 vs. 1550 g/d) were obtained, thus a better outcome than that
243 previously reported by others researcher (Armentano et al., 1993). The authors supported the
244 hypothesis that the greatest responses to improved lysine and methionine concentration occur
245 during the earliest stages of lactation when the need for absorbed amino acids is the highest.
246 On the other hand, different polymers have been used in order to ensure rumen bypass.
247 Recently Carvalho Neto et al. (2019) demonstrated that a lipid matrix of carnauba wax with a
248 core:shell 1:4 in combination with melt-emulsification method is effective as encapsulating
249 system with high methionine protection through the rumen. It is well known that some lipids,
250 particularly non-esterified saturated fatty acids, theoretically provide viable protection against
251 ruminal digestion. In contrast, glycerol is converted to propionate without releasing carbon
252 dioxide and short acids (acetic, propionic, and butyric acids), whereas unsaturated fatty acids
253 undergo biohydrogenation by rumen microorganisms. Carnauba wax resistance to
254 microorganism's degradation has already been tested when used as microencapsulating agent
255 for urea with good outcomes (de Medeiros et al., 2019). A recent research published by
256 Albuquerque et al. (2020) corroborated with these results. The authors proposed the use of
257 lipid nanoparticles as novel rumen-bypass systems to carry lysine. These nanoparticles were
258 produced using different arachidic or stearic acids -based fatty acid and Tween 60 and resisted
259 for up to 24 h ruminal digestion. Tao et al. (2020) investigated the effects of coated
260 cysteamine on growth performance, carcass characteristics, meat quality, and lipid
261 metabolism in finishing pigs and found an increase in carcass lean percentage and *longissimus*
262 *muscle* area alongside higher total protein and free fatty acids but lower serum triacylglycerol
263 and total cholesterol level. These results were explained by an increase in enzymatic activity
264 related to lipid catabolism in adipose tissue and a decrease in activity of enzyme involved in
265 the process of fat synthesis, and thus reducing the fat deposition in subcutaneous adipose

266 tissue. Recently Zhang et al. (2019) demonstrated that the amino acid concentrations of
267 plasma and muscle increased in response to incremental amounts of supplementation with the
268 rumen protected amino acid. The same trend was observed for milk true protein yield. In
269 detail, milk true protein yield increased to 0.98 kg/d when feeding cows with 246 g/d of
270 rumen protected histidine.

271 Urea is the main source of nonprotein nitrogen used in the ruminant diet. It has a low cost per
272 unit of nutrients and is a valuable alternative for partially replace the true protein found in a
273 vegetable meal. However, when ingested urea is rapidly hydrolyzed in the rumen by
274 microbial enzymes producing ammonia (N-NH_3) and carbon dioxide. The accumulation of
275 ammonia in the bloodstream cause intoxication and could lead to animal death. Through the
276 encapsulation processes, the degradation rate of urea in the rumen can be reduced and a slow-
277 release could be obtained (Silva et al., 2013). This interest in slowly rumen released nitrogen
278 compounds primarily stems from their potential to slow ammonia release post-feeding,
279 thereby decreasing peak ammonia concentrations in the rumen. Slow-release urea compounds
280 used to feed ruminants include biuret, starea, urea phosphate, coatings based on oil,
281 formaldehyde-treated urea and polymer-coated urea. The development of products that slow
282 down the ruminal release of ammonia without limiting the extent of urea degradation in the
283 rumen has been challenging. To date, the coatings based on oil or polymers to control the
284 release rate of ammonia from urea seems to be the most promising product (Joysowal et al.,
285 2019). Several studies have been conducted to investigate the influence of feeding dairy cows
286 with slow-release urea with particular attention to the feed intake and nutrient digestibility. A
287 study from Puga et al. (2001) found that the ruminant feed with commercial controlled-release
288 urea significantly increased dry matter intake above the level of the control diet. The authors
289 ascribed the higher digestibility of the experimental diets to the better activity of fiber
290 fermentation in the rumen. It indicates that the commercial controlled-release urea improves

291 nutrient imbalance for rumen bacteria by increasing the availability of energy from simple
292 carbohydrates. These results were in agreement with the findings from Xin et al. (2010), who
293 found that polyurethane coated urea increased the ruminant dry matter intake and the nutrients
294 digestibility compared to the uncoated urea.

295 *2.4 Encapsulation of fatty acids*

296 Generally, animal-based products such as meat and milk are characterized by high levels of
297 saturated fatty acids and an unbalanced omega-6/omega-3 ratio. Therefore, increasing
298 unsaturated fat concentration in animal-based products is one of the research priorities to
299 respond to the concerns of the health-conscious consumers (Lee et al., 2004). High forage
300 quality is certainly crucial in producing meat and milk with a healthy fatty acids profile (Heck
301 et al., 2021). However, efforts to increase polyunsaturated fatty acids (PUFA) levels in
302 ruminant meat and milk had very limited success due to their biohydrogenation in rumen
303 (Francisco et al., 2015). Consequently, attempts to improve the proportion of beneficial fatty
304 acids have been made by protecting dietary PUFA from biohydrogenation through
305 encapsulation techniques (Table 4). In the early 70's Cook et al. (1970) produced PUFA
306 supplements protected with formaldehyde (a known carcinogen generally not allowed in
307 animal feed additives) in order to prevent ruminal biohydrogenation of unsaturated dietary fat
308 during digestion. Lee et al. (2004) encapsulated linoleic acid (LA) using casein treated with
309 acetaldehyde or diacetyl, as an alternative to formaldehyde and determined the concentration
310 of PUFA deposition in the body and muscle fats of lambs fed with the supplements. The
311 results showed that fat of animals fed with protected LA had higher concentrations of both
312 linoleic and linolenic acids (ALA), when compared with lambs fed with untreated
313 supplements after 18 days feeding. Similarly, feeding lambs with LA and ALA encapsulated
314 with formaldehyde-treated protein resulted in an increase in the concentration of LA and ALA

315 in lamb muscle of about 247% and 57%, respectively, compared to those fed with unprotected
316 lipids (Elmore et al., 2005). With the same aim, Alvarado-Gilis et al. (2015a) studied the
317 possibility to protect PUFA from ruminal biohydrogenation through three different methods.
318 In method 1 a blend of ground flaxseed, calcium oxide, and molasses was processed through a
319 dry extruder. For method 2 a blend of ground flaxseed, soybean meal, molasses, and baker's
320 yeast were moistened and pre-warmed and the mixture was subsequently processed through a
321 dry extruder whereas in method 3 ground flaxseed was encapsulated within a matrix of
322 dolomitic lime hydrate (L-Flaxseed) as a protective barrier against biohydrogenation. Steers
323 were fed for 12 days and jugular blood samples were taken for analysis of long-chain fatty
324 acids. Results showed that method 1 and 2 failed to improve resistance of PUFA to
325 biohydrogenation. In contrast, using microencapsulated ground flaxseed, the proportion of
326 ALA resistant to ruminal biohydrogenation was approximately 4-fold greater when compared
327 with ground flaxseed, suggesting that the dolomitic lime hydrate was effective as a protective
328 barrier against biohydrogenation. In a complementary study, the same authors evaluated not
329 only the plasma concentration of PUFA in steers fed with ground flaxseed and encapsulated
330 ground flaxseeds, but also the concentration of fatty acids in the animal tissues (Alvarado-
331 Gilis et al., 2015b). Supplementation with ground flaxseed significantly increased the
332 concentration of ALA in meat and, even more interesting the fact that the ALA content of
333 muscle tissue was 47% greater when flaxseed was encapsulated within the dolomitic lime
334 hydrate matrix. The same trend was obtained by Lee et al. (2007), when they assessed the
335 effect of feeding lambs with a rumen protected protein–oil supplement, made using sodium
336 caseinate and containing α -tocopheryl-acetate and sunflower oil, on fatty acid composition of
337 their muscles over the 10-week feeding trial. Lambs fed with encapsulated sunflower oil and
338 vitamin E were characterized by a higher LA level in both *longissimus dorsi* and *psoas major*
339 *muscles* lipids though a lower palmitic and oleic acid levels were observed when compared to

340 the control. Recently, Kim et al. (2020), fed cows with encapsulated linseed oil coated with
341 hydrogenated palm oil, in order to protect omega-3 in the rumen and found an increase in the
342 omega-3 fatty acids in the milk. The authors performed an *in vitro* and *in vivo* evaluation on
343 the use of a commercial product contained linseed oil, vitamin E, rosemary extract
344 encapsulated in a hydrogenated palm oil shell on rumen fermentation, physiological profile,
345 milk yield, and milk composition in dairy cows. Detailed information about the patented
346 spray-cooling encapsulation process is not available. However, hydrogenated palm oil mostly
347 consisted of molten saturated fatty acids was used as shell material and mixed with linseed oil
348 and other additives. The mixture was atomized into a chilling chamber and cut to a size of
349 1000 to 1500 microns. The obtained microcapsules made of linseed oil (35%), vitamin E
350 (0.5%), rosemary extract (0.3%), and hydrogenated palm oil (64.2%) were added at different
351 concentration 0, 1, 2, 3, 4, and 5% to the diets and the ruminal fluids were collected. The
352 results of the *in vitro* ruminal fermentation suggested that 2% of encapsulated linseed oil was
353 an optimal dosage in cows' diets, therefore this level was chosen for the feed supplementation
354 of 36 cows. Results of the *in vivo* experiment showed that the use of the commercial
355 encapsulated linseed oil significantly increased the yield of total omega-3 fatty acids
356 compared to the non-encapsulated linseed. Accordingly, the ratio of total omega-6 to omega-3
357 fatty acids significantly decreased when linseed oil was used in microencapsulated form.
358 Encapsulation technology also seemed to improve meat content in conjugated linoleic acids
359 (CLA). CLA are mixture of positional and geometric isomers of octadecadienoic acid
360 containing a pair of double bonds in a conjugated configuration are widely recognized as a
361 nutrient that exerts important physiological effects on human health. Numerous studies and
362 reviews have investigated the supposed health benefits including anti-cancer, anti-
363 atherogenic, anti-adipogenic, anti-diabetogenic, anti-inflammatory and effects on bone health
364 (Lehnen et al., 2015; McCrorie et al., 2011; McGuire and McGuire, 2000). It has been

365 reported that encapsulation of the CLA in a matrix of protein provided a protection of about
366 70% with a 30% hydrogenation of the CLA isomers (Gulati et al., 2000). Gulati et al. (2020)
367 also showed that lactating goats fed with 80 g/animal per day of freeze dryer protected CLA-
368 casein (1:1, w/w) led to an increase in the proportion of isomers 9-cis 11-trans and 10-trans
369 12-cis in milk fat. The total CLA levels were enhanced by about 10-fold above the control
370 levels present in milk fat of goats fed with unprotected CLA, with a transfer efficiency of to
371 the milk fat of 36-41% and 21-30% for the isomers 9-cis 11-trans and 10-trans 12-cis,
372 respectively. The values observed in this research were higher than those obtained by
373 Chouinard et al. (1999), with the CLA infusion directly into the abomasum of dairy cows.
374 Interesting is also the possibility to use CLA in order to reduce the milk fat yield, as
375 demonstrated by Peterson et al. (2002). The effectiveness of dietary supplements of CLA in
376 animal feed is related to the extent to which their metabolism by rumen bacteria is avoided.
377 On this background, Perfield et al. (2004) investigated the effects of rumen-protected CLA
378 using lipid-based coating material on milk fat synthesis. A dosage of 138 g/d over the 7-day
379 treatment period was able to reduce the milk fat yield of about the 22%. Encapsulation of oils
380 has also proved useful in improvement of the nutritional characteristics of eggs. The current
381 practice for producing omega-3 fatty acid-enriched eggs generally consists in feeding laying
382 hens with flaxseed containing the omega-3 fatty acid ALA. Since laying hens have the ability
383 to convert ALA to docosahexaenoic acid (DHA) as well as to eicosapentaenoic acid (EPA) to
384 some extent, eggs obtained in this way are richer in these important fatty acids (Gonzalez-
385 Esquerra and Leeson, 2001) The enrichment of eggs with EPA has been obtained through the
386 use of marine oils (fish oil, fish meal) (Cachaldora et al., 2006). Here encapsulation is also
387 used to mask the “fishy odors” and “fishy off-flavors” and to increase the feed consumption
388 by laying hens. Indeed Lawlor et al. (2010) evaluated the effect of a diet supplemented with
389 microencapsulated fish oil on DHA and EPA concentrations of table eggs, where 96 laying

390 hens were randomly divided into 4 groups and fed with 0, 20, 40, or 60 g/kg for 21 days with
391 a commercial product containing EPA and DHA from oily fish sources in a ratio of 18:12.
392 The total omega-3 fatty acids significantly increased as the level of microencapsulated fish
393 passed from 0 to 60 g/kg diet. Also, EPA and DHA were significantly higher with a 40-fold
394 increase for the EPA and a 2.6-fold increase for DHA, when the laying hens were fed with 60
395 g/kg diet. Hard boiled and scrambled eggs were also evaluated from a sensorial point of view
396 at the highest level of inclusion tested (60 g /kg diet) and only minor impact on the attributes
397 “off-flavor” and “sulphur” was noted for boiled eggs. Conversely, there were no significant
398 sensory differences between treatments for scrambled egg samples, demonstrating that the
399 off-flavor perception was dependent on the method of preparation.

400 *2.5 Encapsulation of organic acids*

401 The ban on the use of antibiotic growth promoters in animals’ diet resulted in an increasing
402 use of natural alternative substances that might be positive for both production performance
403 and animal health. In this regard, probiotics, prebiotics, symbiotics and organic acids or their
404 salts are promising alternatives. However, organic acids need to reach the intestine, without
405 being absorbed too rapidly upon leaving the stomach. In fact, different research papers
406 suggested that the effectiveness of unprotected organic acids may be limited, due to their
407 prompt absorption and metabolism in the duodenum (Partenen, 2001, Upadhaya et al. 2014),
408 which could cause bacterial resistance to acids and negative effects on gut mucosal membrane
409 (Bearson et al. 1997). Therefore, many studies have been performed to investigate the effect
410 of encapsulation as a tool to improve the delivery and efficacy of these compounds (Table 5).
411 Piva et al. (2007) demonstrated that the microencapsulation of vanillin and sorbic acid in a
412 hydrogenated vegetable lipid shell effectively allowed the slow-release of both active
413 ingredients and prevented the immediate disappearance of such compounds upon exiting the
414 stomach. Grilli et al. (2010) reported a significant antimicrobial effect of a commercial blend

415 of citric and sorbic acids microencapsulated with hydrogenated vegetable lipids, when feeding
416 pigs for 41 days with 3 kg/ton. They observed a reduction in volatile fatty acids and an
417 enhanced fermentation in the large bowel with increased acids and tyramine production
418 alongside an improved growth performance, with a higher average daily gain between 15 and
419 41 day and a higher body weight at 41 day for the treated group, when compared to control.
420 Also, there was higher pig's feed intake throughout the study. Some studies harnessed the
421 potential synergy between organic acids and plant-based essential oils (Gheisar et al., 2015;
422 Tugnoli et al., 2020). Indeed, supplementing diets with microencapsulated blends of organic
423 acids and essential oils (citric acid, 25%; sorbic acid, 16.7%; thymol, 1.7%; vanillin, 1.0%)
424 loaded inside starch as a carrier (55.6%) showed a positive effect on growth performance
425 broilers. The product used was a commercial microencapsulated organic acid added to the
426 basal diet from 0 to 0.075%. The study showed an improvement of the broiler's growth
427 performance and a decrease in drip loss percentage, without affecting other meat quality
428 parameters, such as pH, color, and the relative organs weights. Similar results were reported
429 by Cho et al. (2014), who demonstrated that the supplementation with the same
430 microencapsulated commercial blend added at 0.05% to a finisher pigs' diet, significantly
431 improved their growth performance. In addition, the supplementation was reflected in the
432 increase in nutrient digestibility. Possible mechanisms for these effects include a reduction in
433 the stomach pH and a direct action of the microencapsulated organic acids on the gastro-
434 intestinal tract microbiota. This is because, the supplementation with the microencapsulated
435 blend provokes changes in gut luminal metabolites, such as volatile fatty acids and
436 polyamines. Volatile fatty acids deriving from the bacterial are rapidly absorbed by the
437 epithelial cells of pigs, being metabolized to supply energy, and this could explain the
438 increased growth and nutrient digestibility observed. However, dietary supplementation with
439 the microencapsulated blend had no effect on meat color, sensory evaluation, cooking loss,

440 meat pH and water holding capacity. The same commercial blend was used also by Stamilla
441 et al. (2020), who fed chicken for the entire growing cycle at the level of 0.5%. Their dietary
442 strategy led to a significant reduction in intramuscular fat content and an overall improvement
443 in fatty acid profile. This dietary supplementation was also led to lipid oxidation reduction in
444 the meat, which is one of the main problems related to chicken meat shelf life. Recently also
445 Galli et al. (2020a) have dealt with this topic evaluating the effect encapsulated organic acid
446 blend based on formic, phosphoric, lactic, acetic, butyric, and propionic acid with glycerol
447 and silicon dioxide on the growth performance and meat quality of broilers. Results
448 demonstrated that the encapsulated organic acid blend improves meat quality, with increased
449 antioxidant levels and reduction of lipid peroxidation. Also, there was a decrease in total
450 saturated fatty acid and an enhancement in monounsaturated/polyunsaturated fatty acid levels
451 in broilers meat.

452 *2.6 Encapsulation of essential oils and plant extracts*

453 Phytogetic feed additives are commonly defined as plant secondary compounds and
454 metabolites with beneficial effects on animal health and production, including animal
455 products. Essential oils represent a major group of phytogetic feed additives. These
456 compounds are usually added, at recommended levels, into animal feed and can have a broad
457 range of bioactive properties, including feed palatability, digestive functions, animal intestinal
458 microbiome structure, as well as improved production performance and quality of animal
459 products in poultry, pigs, ruminant and aquaculture animals (Stevanović et al., 2018).
460 However, their high reactivity represents an obstacle for direct application and incorporation
461 into food and feed products. The main challenge in using these compounds is related to the
462 preservation of their biological activity, as well as minimizing their impact on organoleptic
463 properties of formulated feeds. The biological activity and the stability of essential oils could
464 be compromised by temperature, light, metals, and water and oxygen (Turek and Stintzing

465 2013). For example, when a temperature of 58 °C is applied, which is the processing
466 condition used for feed pelleting, the recovery of the essential oil indicator substances was
467 low (Maenner et al., 2008). Secondly, the interaction between the essential oils the feed
468 matrix could influence their activities (Śliwiński et al., 2002). Therefore, the encapsulation of
469 essential oils is a reasonable choice for the supplementation of animal feed and, often,
470 commercial blend containing different essential oils are used. The positive effect of
471 encapsulated essential oils on the quality of ruminant, poultry and lamb meat is evidenced by
472 several published works (Favaretto et al., 2020; Galli et al., 2020b; Spanghero et al., 2009),
473 some of which summarized in Table 6. Recently, Galli et al. (2020b) explored the possibility
474 of using a commercial phytogetic product with microencapsulated essential oil components
475 (based on thymol, cinnamaldehyde and carvacrol) in the diet of 225 broiler chickens, in order
476 to verify the effect on the meat composition with reference to the fatty acid profiles. There
477 were significantly lower total saturated fatty acid levels with reduction of lauric and palmitic
478 acid, which are undesirable as associated with the increase in LDL serum cholesterol.
479 Moreover, a significantly greater monounsaturated/polyunsaturated fatty acid levels in
480 broilers that consumed the commercial microencapsulated oil alone or in combination with
481 curcumin was observed. With this research, the authors demonstrated that the fatty acids
482 profile in broiler meat can be manipulated using various ingredients and/or additives.
483 Additionally, Gomathi et al. (2018) reported that the reduction of the meat cholesterol content
484 of broiler chicken supplemented with microencapsulated cinnamon oil and sodium butyrate
485 had no effect on pH, cooking loss, water-holding capacity of meat. The sensorial attributes
486 such as appearance, flavor, texture, mouth coating, juiciness and overall acceptability of meat
487 were not influenced by the supplementation. Recently, Favaretto et al. (2020) used a
488 commercial blend containing essential oils microencapsulated. According to the
489 manufacturer, the product contains carvacrol, thymol and cinnamaldehyde. The objective of

490 the study was to determine whether supplementation with these microencapsulated essential
491 oils added to feed of growing lamb would affect their growth performance or produce
492 antioxidants and immunological responses. Thirty female lambs were randomly distributed in
493 three groups, as follows: control (basal feed), T500 (basal feed supplemented with 500 mg of
494 microencapsulated essential oils /kg of concentrate) and T1000 (basal feed with 1000 mg of
495 microencapsulated essential oils /kg of concentrate). Weights were measured and blood was
496 collected on days 0, 15, 30 and 40 of the study. During the 40 days' trial, lambs belong to the
497 T1000 group showed greater weight gain. Moreover, T1000 lambs had lower numbers of
498 leukocytes and lymphocytes compared to control group. This supplementation changed the
499 antioxidant profile without altering the levels of free radicals and lipid peroxidation, which
500 could be a consequence of the high doses consumed by the animals. The activities of
501 glutathione peroxidase and glutathione reductase were lower in the T1000 animals than in the
502 control at day 30. In light of this, the authors concluded that supplementation with
503 encapsulated essential oil in lambs stimulated antioxidant and anti-inflammatory responses
504 and consequently increased weight gain in lambs. Similarly, a commercial blend of oregano,
505 cinnamon, thyme and orange-peel essential oils was used to assess the influence of increasing
506 concentration of the encapsulated mixture on milk yield and composition of primiparous dairy
507 cows (Spanghero et al., 2009). In detail, the ruminant feed was fortified to provide 0, 40, 80
508 or 120 g/d of the microencapsulated essential oils and the ruminants were assigned to one of
509 four dietary treatments. These authors reported a numerical trend for increased milk protein
510 and fat concentration. The dietary supplementation with microencapsulated essential oils had
511 no effect on dry matter intake, water consumption and milk production. Microencapsulated
512 essential oils were also used to assess the possibility of modifying milk characteristics where
513 Guasch et al. (2006) fed 40 cows for 56 days using 1.66 g/ kg DM with a combination of
514 microencapsulated essential oil compounds containing coriander oil, geranylacetate, and

515 eugenol. Their results showed that milk fat and milk protein were not affected by the
516 supplementation with the microencapsulated essential oil and only a higher milk production
517 between 25 and 56 day was observed. Different studies have disclosed that the use of
518 encapsulated plant extracts could be useful to improve the animal body weight. As example,
519 nanoencapsulation of aloe vera, dill, and nettle roots extracts using chitosan as shell material
520 significantly improved the broiler chickens body weight gain after a 42-day experiment.
521 Based on their results, Meimandipour et al. (2017) suggested that chitosan nanocapsules could
522 be used as a substitute for antibiotics in the diet of broiler chickens. Likewise, the
523 encapsulated form of garlic and *Phyllanthus niruri L.* mixture within an arabic gum shell
524 revealed promising effects on live weight gain of broilers (Natsir et al., 2013). In another
525 research, nanoencapsulated turmeric acid extract obtained through bottom-up approach with
526 ionic gelation method was able to reduce meat cholesterol of broiler chicken fed with
527 supplemented diet. According to the obtained results, the authors conclude that 0.4% of
528 nanocapsules was useful to improve the meat quality of broiler chicken (Sundari et al., 2014).
529 Promising results were also obtained on rainbow trout (*Oncorhynchus mykiss*) juveniles fed
530 with microencapsulated garlic extract especially in terms of growth performance. In addition,
531 fish fed with 0.5% microcapsules had higher protein and lower lipid content when compared
532 to the fish fed with the control diet (Adineh et al., 2020).

533 2.7 Encapsulation of phenolic compounds

534 The utilization of phenolic compounds and tannins has become very important in animal and
535 specifically in ruminant feeding as a result of increasing studies that evidence their biological
536 significance in animal nutrition. In general, the effects of tannins in ruminants' diet are related
537 to their ability to form complexes mainly with proteins, polysaccharides whilst modulating
538 ruminal fermentation which results in the improvement of the feed utilization and the final
539 product quality (Costa et al., 2018, Patra and Saxena, 2011). They have been proposed as

540 alternatives for antibiotic growth promoters in feed due to their antimicrobial, antioxidant,
541 anti-inflammatory and gut health promoting when used at the appropriate concentration (Choi
542 and Kim, 2020). Tannins can be broadly classified into hydrolysable and condensed tannins
543 and their effect strongly depends on several factors including their molecular structure. The
544 use of condensed tannin significantly reduces the enteric methane production (Costa et al.,
545 2018) due to the direct inhibition of growth of the target methane-producing archaeal
546 population (Animut et al., 2008). Condensed tannins also inhibit biohydrogenation (Vasta et
547 al., 2019) and this would contribute to improve the quality of animal-based products,
548 including the enrichment of conjugated linoleic acid (CLAs) in meat and milk (Patra and
549 Saxena, 2011). However, the use of tannins in animal feed has important limitations mostly
550 related to their astringency and bitter taste, which among other negative consequences, results
551 in the reduction of voluntary dry matter intake in the animals (Dschaak et al., 2011). The
552 administration of condensed tannin extracts could be thus facilitated by formulating them into
553 products that could mask their taste, retain their structural integrity until consumption,
554 increase their bioavailability, as well as release them precisely at the target site (Munin and
555 Edwards-Lévy 2011). A careful selection of the appropriate encapsulation technology would
556 pave the way for their integration into feeds. For example, the release of condensed tannins
557 from the microcapsules must take place in the rumen where the modulation of
558 biohydrogenation occurs (Tolve et al., 2021). Adejoro et al. (2019a) tested the ability to use
559 encapsulated condensed tannins extract in ruminant feeding, using a water-soluble extract
560 from the *Acacia mearnsii* tree as bioactive compound source and a combination of arabic gum
561 and maltodextrin or native starch as shell material. The effect of the shell material and the
562 core:shell ratio on the encapsulation efficiency, the loading and the *in vitro* kinetics of the
563 tannin extract from microcapsules. According to their results, although starch and
564 maltodextrin-arabic gum were successfully used in encapsulating the extract, the resulting

565 encapsulates exhibited burst release profile under *in vitro* release in the digestive conditions.
566 In order to overcome this problem, the same research group produced microencapsulated
567 condensed tannins using the solid-in-oil-in-water and melt dispersion method choosing lard
568 and palm oil as shell material. The results showed that both lard and palm oil could be used to
569 encapsulate the condensed tannin extract with high encapsulation efficiencies, and good
570 release of the tannin extract during the *in vitro* simulation of the digestive system. Moreover,
571 the condensed tannins microencapsulated using lard or palm oil as shell materials also
572 resulted in reduction in total gas and methane production evaluated through *in vitro* tests
573 (Adejoro et al., 2018). Adejoro et al. (2019b) evaluated the efficacy of encapsulated tannin
574 extract in a lipid matrix using Africa Mutton Merino wethers as a model animal and found a
575 higher nutrient digestibility (neutral detergent fiber digestibility) with better reduction in
576 methane production for encapsulated extract when compared to non-encapsulated
577 formulation. However, these researchers did not assess the effect of the encapsulated tannins
578 on the fatty acids biohydrogenation and the derivative animal product. A recent study
579 investigating the possibility of using microencapsulated anthocyanins as means to improve
580 the production of natural pigments and provide natural pigments that enhance growth and
581 pigmentation of fantail goldfish (Vanegas-Espinoza et al., 2019) is another compelling
582 example of the broad application of encapsulation in animal feed. In this case, the technology
583 allows the production of natural pigments and with enhanced stability which is known to be
584 deeply influenced by environmental conditions such as light, oxygen, pH and temperature. In
585 detail, Vanegas-Espinoza et al. (2019) microencapsulated Roselle calyx extract, (*Hibiscus*
586 *sabdariffa*) via spray drying, using maltodextrins and sucrose as shell materials. The
587 microcapsules were then used as supplement in Fantail goldfish (*Carassius auratus*) feed and
588 the skin pigmentation of the fish was used as an indicator of the effectiveness of
589 microencapsulation to protect anthocyanins and ensure their transport to the digestive system

590 of the fantail goldfish where pigments are absorbed. It was observed an enhanced fish skin
591 color with the increased supplementation level of microencapsulated anthocyanins from 150,
592 300, and 450 mg anthocyanin/kg of diet.

593 2.8 Encapsulation of probiotics and prebiotics

594 Probiotics are known as live microorganisms which when introduced into a host in adequate
595 amounts confer health benefit whereas prebiotics act as fermentable substrates for probiotics
596 and promote their growth. The positive effect of probiotics is now recognized with many
597 reviews and studies suggesting that their addition into the feed improves the growth
598 performance, population of microflora, intestinal microbiota, gut health, white blood cells and
599 regulates the immune system of animals (Chen and Yu 2020; Abd El-Ghany, 2020). For
600 example, lactating cows fed diets based on microbial ensiled straw resulted in increased milk
601 production when compared with diets based on untreated agricultural by-products (Guo et al.,
602 2002). *Lactobacillus* supplementation improved feed conversion rate, broiler chickens'
603 productive performances and metabolic functions (De Cesare et al., 2017) and enhanced the
604 fatty acid profile of meat even when broiler chickens were under heat stress (Jahromi et al.,
605 2016). This microbial effect is related to their metabolites (postbiotics) and enzymes
606 degrading fibrous compounds thereby increasing digestibility and nutrients utilization.
607 Encapsulation technology increases probiotics stability during storage and processes, as well
608 as their ability to pass unchanged through the mouth to the gizzard and intestines where they
609 are most effective (Natsir et al., 2019) and therefore could enhance the effect of probiotics
610 (Table 7). A study conducted on 192 hybrid ducks fed with non-encapsulated and
611 encapsulated mixtures of natural acidifier (lactic acid and citric acid), phytobiotic (*Allium*
612 *sativum* and *Phyllanthusiruri*) and probiotic (*Bacillus coagulans*) incorporated into the feed at
613 different concentrations (0, 0.5, 1.0 and 1.5 %), showed that encapsulated formulations

614 resulted in improved performance on production, intestinal morphometry and microflora with
615 better body weight gain, feed conversion, number of *Lactic acid bacteria* and reduced number
616 of *E. Coli* than non-encapsulated form of supplements (Natsir et al., 2019). The same authors
617 suggested an optimum concentration of 1.5% which could partially explain reason behind the
618 results obtained by Ardiansah et al. (2020) with no effect on dietary supplementation of either
619 powdered or encapsulated probiotic (*Lactobacillus fermentum*, *Lacobacillus acidophilus*,
620 *Bacillus spp*) on relative carcass, gibleet weight and intestinal morphometry of duck since the
621 concentration used was only up to 0.4%. Potential synergy between phytochemicals, probiotic
622 and organic acids used could have also contributed to the difference in the results from the
623 two studies. With a different approach, Park and Park (2011) reported that the use of
624 microencapsulated inulin, a prebiotic with beneficial effects on animal health due to its ability
625 to stimulate the selective growth of microorganism in the large intestine, in broiler chickens'
626 diet significantly increases the breast weight.

627 **3. Encapsulation of bioactive ingredients and animal welfare in a sustainable production** 628 **context**

629 Animal welfare is considered as a combination of both physical and mental well-being
630 (Phillips, 2016) and could be affected by several factors, including thermal stress, disease,
631 natural toxins and nutrient imbalances. This section discusses how the encapsulation of
632 bioactive compounds could be used to improve animals' welfare, while increasing
633 profitability for farmer and ensuring safety and quality through the food chains with reduced
634 environmental impact. The use of the encapsulation to improve animal welfare is viewed
635 through the five freedoms of animal welfare lenses according to the Farm Animal Welfare
636 Committee (FAWC) (2014) presented in Table 8. A compelling example where encapsulation
637 could have been used in this context is the study by Girard and Matte (2005), demonstrating

638 an increase in the packed cell volume, blood hemoglobin and serum vitamin B₁₂ of cows as
639 well as the quantity of milk produced and its concentration in vitamin B₁₂ after a weekly
640 intramuscular injection of 10 mg vitamin B₁₂ accompanied with folic acid and rumen-
641 protected methionine supplemented feeds. Although the injection intramuscular avoided first-
642 pass metabolism and achieved a precise plasma levels of the bioactive compound, it can cause
643 severe tissue trauma and represent local portal of infection, even when correctly administrated
644 (Abbate et al., 2018). Furthermore, the process is painful and may create a sense of fear, thus,
645 could have potentially negatively affected four freedoms of animal welfare: freedom from
646 discomfort, freedom from pain, injury and disease, freedom to express normal behavior, and
647 freedom from fear and distress (Table 8). The bioactive compound could have been
648 encapsulated in a suitable shell designed to allow good stability and targeted delivery (see
649 section with focus on vitamins) since oral administration is safer, non-invasive, simple and
650 convenient for the animals. For example, it has been demonstrated that encapsulated vitamin
651 E in feed resulted in the improvement of broilers growth and the quality of the meat whilst
652 enhancing hepatic mitochondrial total antioxidant capacity and glutathione peroxidase
653 enzyme activity (Hu et al., 2015), thus contributed to reinforcing the immune system of the
654 broilers and their resistance to free radical's attack. Similarly, acidification with various
655 organic acids to diets such as fumaric, propionic, lactic and sorbic acid have been reported to
656 decrease colonization of pathogen and production of toxic metabolites whilst improving
657 digestibility of protein Ca, P, Mg and Zn as well as serving as substrate for the intermediary
658 metabolism (Suiryanrayna and Ramana 2015; Swiatkiewicz and Arczewska-wlosek 2012).
659 Choi et al. (2020) reported that the supplementation of microencapsulated organic acid and
660 essential oil enhanced intestinal morphology and showed anti-diarrhea effects in weaned
661 piglets challenged with enterotoxigenic *E. Coli* F4, and thus contribute to ensuring freedom
662 from pain and disease. Zhang et al. (2015) showed an effective delivery of high concentration

663 of a model essential oil, carvacrol, to the jejunum and ileum of chickens using a combination
664 of food-grade biopolymers as shell material where alginate provided the gastric resistance
665 microparticles, while whey protein modulated the intestinal release. This high release was
666 sustained for more than 3 h after oral administration, though the *in vivo* release of carvacrol
667 alginate-whey microparticles was faster when compared to the release from *in vitro*
668 simulation. This work reinforced the point that *in vitro* simulation could only serve as a tool
669 for formulation optimization and the need for *in vivo* study. Purba et al. (2020), after a
670 systematic review and meta-analysis to predict and identify ways to increase conjugated
671 linoleic acid formation in ruminant-derived products, reached the same conclusion that *in vivo*
672 method was more suitable for the direct observation of fatty acid transformation than the *in*
673 *vitro* method. This is even more important when come to assessing the effect of formulations
674 of animal welfare as direct of observation of the behavior also come into play in addition to
675 biological markers of their well-being. Encapsulation also creates opportunities for an
676 integrated approach in the formulation and delivery of actives compounds for animal welfare
677 by harnessing synergy between different actives to allow maximum efficacy, reduction
678 potential risk of toxicity and to limit the production of greenhouse gas emissions. Indeed, it
679 has been demonstrated that symbiotic formulations had enhanced beneficial impact on
680 piglets' health especially during weaning period, when compared to commercial probiotics
681 used individually (Chlebicz-Wójcik and Śliżewska 2020). Here one could also imagine
682 formulations with symbiotic and other bioactive compounds such as polyphenols (Choi et al.,
683 2020), omega-3 fatty acids (Baéza et al., 2017) to avoid energy excesses, digestive disorders
684 and minimize inflammatory responses, thereby optimizing their immune system and
685 contributing to ensure the five freedoms of animal welfare. In the context of sustainable
686 production, where the use of renewable natural ingredients and available feed resources to
687 their full potential alongside waste reduction are becoming necessary, the encapsulation of

688 natural extracts containing mixed bioactive compounds from agricultural waste and by-
689 product from food processing to improve their stability, delivery and efficacy for animal
690 welfare and nutrition is important. In this regard, Mamvura et al. (2014) encapsulated nitrate
691 with sesame gum and demonstrated *in vitro* the sustained release of nitrate and the
692 microparticles potential for reduction methane production, thus enhancing sustainability of
693 ruminant production while reducing the risk of nitrite toxicity. A preliminary *in vitro* study by
694 Santos et al. (2020) showed calcium salts of long-chain fatty acids may accelerate propionate
695 production and reduce methane production by ruminants. Hence, calcium salts of long-chain
696 fatty acids could be a potential shell material for developing new multifunctional animal feeds
697 loaded with different bioactive compounds Finally, to ensure welfare of animal when using
698 encapsulated bioactive compounds, special attention should be paid to the toxicity aspect of
699 such approach, especially when considering nano size materials. Factors affecting the toxicity
700 include materials intrinsic toxicity, the size, shape and surface chemistry of the encapsulates.
701 Although nanoparticles have been proving to improve the bioavailability and absorption of
702 micronutrients and other bioactive compounds, it has been reported that epithelial cells of the
703 small intestine are capable of absorbing nanoparticles less than 200 nm in size, whereas some
704 literature suggested that smaller (<5 µm) are absorbed through the circulatory system
705 (Bribiesca et al., 2017). Hence, nanoparticles have the potential to reach the circulatory
706 system and be distributed anywhere in the body, principally in the liver, spleen and other
707 organs of the reticuloendothelial (Rabanel et al., 2012). The toxicity of nanoparticles depends
708 on their persistence in the organs, where they are deposited and the physiological response of
709 the host tissue in maintaining or eliminating waste of the shell materials used in their
710 manufacture.

711 **Conclusion and outlook**

712 This review has highlighted the potential of encapsulation technology to improve formulation
713 of bioactive compounds into the feed matrix and their delivery for better animal production
714 efficiency, animal welfare and quality of animal-based products. Different encapsulation
715 approaches with a wide range of bioactive compounds are reported in the literature showing
716 promising results for enriching animal-based product with feeds containing encapsulated live
717 micro-organism and their metabolites, probiotics, phytobiotic in a form of a single type
718 compound or blend of several compounds as growth promoter alternative to antibiotic for safe
719 production across the food chain. Products such as eggs, meat, fish and milk could be
720 enriched with selected micronutrients and other bioactive compounds through appropriate
721 feeding, but to our knowledge, no literature is present dealing the effect of the encapsulated
722 bioactive ingredients on dairy products. For example, it was observed an improvement in the
723 content and profile of amino acids, fatty acids with increase in omega -3 fatty acid and
724 conjugated linoleic acids while decreasing the content of cholesterol in some animals and
725 subsequent food products, which are desirable attributes from consumers' health perspective.
726 The review also showed how the encapsulated bioactive compound could contribute to
727 support the five freedoms of animal welfare: freedom from malnutrition, freedom from
728 discomfort, freedom from pain, injury and disease, freedom to express normal behavior, and
729 freedom from fear and distress. However, there was limited number of publications that
730 combined *in vitro* and *in vivo* studies of the effect of encapsulated bioactive compounds with
731 focus on the animal welfare. *In vivo* studies are very important when come to assessing the
732 effect of formulations of animal welfare as direct of observation of the behavior also come
733 into play in addition to biological markers of their well-being. Further investigations should
734 focus on addressing this apparent gap in this key knowledge. The development of simple,
735 cheap, effective and environmentally friendly smart encapsulation system for delivery of
736 bioactive compounds in animal feed deserves more research. Moreover, future research

737 should give more consideration to factors that affect the mechanical disintegration of
738 encapsulates during rumination, the effect of particles size on the amount lost from the rumen
739 alongside gas via the belching process as such information would be vital for optimizing the
740 encapsulation formulation and manufacturing process for better outcome in terms of
741 profitability for farmers, animal welfare and the derived food product quality as well as green
742 gas reduction during production.

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753

754 **Declaration of competing interest**

755 The authors declare no conflict of interest.

756

757 **References**

758 Abbate, A., Almasio, P.L., Mongitore, M., Di Vita, G., Patti, R., 2018. Necrotizing soft
759 tissue fasciitis after intramuscular injection. Case. Rep. Sug. 2018.
760 doi:[10.1155/2018/3945497](https://doi.org/10.1155/2018/3945497)

761 Abd El-Ghany, W.A., 2020. Paraprobiotics and postbiotics: Contemporary and promising
762 natural antibiotics alternatives and their applications in the poultry field. *Open Vet. J.* 10, 323-
763 330. doi:[10.4314/ovj.v10i3.11](https://doi.org/10.4314/ovj.v10i3.11).

764 Adejoro, F.A., Hassen, A., Thantsha, M.S., 2018. Preparation of acacia tannin loaded lipid
765 microparticles by solid-in-oil-in-water and melt dispersion methods, their characterization and
766 evaluation of their effect on ruminal gas production In Vitro. *PloS one* 13, e0206241.

767 Adejoro, F.A., Hassen, A., Thantsha, M.S., 2019a. Characterization of starch and gum
768 arabic-maltodextrin microparticles encapsulating acacia tannin extract and evaluation of their
769 potential use in ruminant nutrition. *Asian-australas. J. Anim. Sci.* 32(7), 977-987.
770 doi:[10.5713/ajas.18.0632](https://doi.org/10.5713/ajas.18.0632).

771 Adejoro, F.A., Hassen, A., Akanmu, A.M., 2019b. Effect of lipid-encapsulated acacia
772 tannin extract on feed intake, nutrient digestibility and methane emission in
773 sheep. *Animals* 9(11), 863-375. doi:[10.3390/ani9110863](https://doi.org/10.3390/ani9110863).

774 Adineh, H., Harsij, M., Jafaryan, H., Asadi, M., 2020. The effects of microencapsulated
775 garlic (*Allium sativum*) extract on growth performance, body composition, immune response
776 and antioxidant status of rainbow trout (*Oncorhynchus mykiss*) juveniles. *J. Appl. Anim. Res.*
777 48(1), 372-378. doi:[10.1080/09712119.2020.1808473](https://doi.org/10.1080/09712119.2020.1808473).

778 Albright, R.B., Kowarski, C.H., 1994. Improved stability of lipid coated vitamin A in
779 animal feed additives. *Drug Dev. Ind. Pharm.* 20(12), 2035-2039.
780 doi:[10.3109/03639049409049336](https://doi.org/10.3109/03639049409049336).

781 Albuquerque, J., Casal, S., de Jorge Páscoa, R.N.M., Van Dorpe, I., Fonseca, A.J.M.,
782 Cabrita, A.R.J., Neves, A.R., Reis, S., 2020. Applying nanotechnology to increase the rumen
783 protection of amino acids in dairy cows. *Sci. Rep.* 10(1), 1-12. doi:[10.1038/s41598-020-](https://doi.org/10.1038/s41598-020-63793-z)
784 [63793-z](https://doi.org/10.1038/s41598-020-63793-z).

785 Alvarado-Gilis, C.A., Aperce, C.C., Miller, K.A., Van Bibber-Krueger, C.L., Klamfoth,
786 D., Drouillard, J.S., 2015a. Protection of polyunsaturated fatty acids against ruminal
787 biohydrogenation: Pilot experiments for three approaches. *J. Anim. Sci.* 93(6), 3101-3109.
788 doi:[10.2527/jas.2014-8015](https://doi.org/10.2527/jas.2014-8015).

789 Alvarado-Gilis, C.A., Aperce, C.C., Miller, K.A., Van Bibber-Krueger, C. L., Klamfoth,
790 D., Drouillard, J.S., 2015b. Effects of flaxseed encapsulation on biohydrogenation of
791 polyunsaturated fatty acids by ruminal microorganisms: feedlot performance, carcass quality,
792 and tissue fatty acid composition. *J. Anim. Sci.* 93(9), 4368-4376. doi:[10.2527/jas.2015-9171](https://doi.org/10.2527/jas.2015-9171).

793 Animut, G., Puchala, R., Goetsch, A.L., Patra, A.K., Sahlu, T., Varel, V.H., Wells, J.,
794 2008. Methane emission by goats consuming diets with different levels of condensed tannins
795 from lespedeza. *Anim. Feed Sci. Technol.* 144(3-4), 212-227.
796 doi:[10.1016/j.anifeedsci.2007.10.014](https://doi.org/10.1016/j.anifeedsci.2007.10.014).

797 Ardiansah, I., Sholiha, K., Sjojfan, O., 2020. Dietary supplementation of powdered and
798 encapsulated probiotic: In vivo study on relative carcass, gibleet weight and intestinal
799 morphometry of local duck. *Acta Sci.* 42, e47140. doi:[10.4025/actascianimsci.v42i1.47140](https://doi.org/10.4025/actascianimsci.v42i1.47140).

800 Armentano, L.E., Swain, S.M., Ducharme, G.A., 1993. Lactation response to ruminally
801 protected methionine and lysine at two amounts of ruminally available nitrogen. *J. Dairy*
802 *Sci.* 76(10), 2963-2969. doi:[10.3168/jds.S0022-0302\(93\)77635-3](https://doi.org/10.3168/jds.S0022-0302(93)77635-3).

803 Arshad, U., Zenobi, M.G., Staples, C.R., Santos, J.E.P., 2020. Meta-analysis of the effects
804 of supplemental rumen-protected choline during the transition period on performance and
805 health of parous dairy cows. *J. Dairy Sci.* 103(1), 282-300. doi:[10.3168/jds.2019-16842](https://doi.org/10.3168/jds.2019-16842).

806 Baéza, E., Chartrin, P., Bordeau, T., Lessire, M., Thoby, J. M., Gigaud, V., Blanchet, M.,
807 Alinier, A., Leterrier, C., 2017. Omega-3 polyunsaturated fatty acids provided during
808 embryonic development improve the growth performance and welfare of Muscovy ducks
809 (*Cairina moschata*). *Poult. Sci.* 96(9), 3176-3187. doi: [10.3382/ps/pex147](https://doi.org/10.3382/ps/pex147).

810 Bearson, S., Bearson, B., Foster, J.W., 1997. Acid stress responses in
811 enterobacteria. *FEMS Microbiol. Lett.* 147(2), 173-180. doi:[10.1111/j.1574-](https://doi.org/10.1111/j.1574-6968.1997.tb10238.x)
812 [6968.1997.tb10238.x](https://doi.org/10.1111/j.1574-6968.1997.tb10238.x).

813 Bribiesca, J.E.R., Casas, R.L., Monterrosa, R.G.C., Pérez, A.R., 2017. Supplementing
814 selenium and zinc nanoparticles in ruminants for improving their bioavailability meat.
815 In *Nutrient Delivery* (pp. 713-747). Elsevier. ISBN 978-0-12-804304-2.

816 Broderick, G.A., Kowalczyk, T., Satter, L.D., 1970. Milk production response to
817 supplementation with encapsulated methionine per os or casein per abomasum. *J. Dairy*
818 *Sci.* 53(12), 1714-1721. doi:[10.3168/jds.S0022-0302\(70\)86468-2](https://doi.org/10.3168/jds.S0022-0302(70)86468-2).

819 Cachaldora, P., García-Rebollar, P., Alvarez, C., Blas, J. D., Méndez, J., 2006. Effect of
820 type and level of fish oil supplementation on yolk fat composition and n-3 fatty acids
821 retention efficiency in laying hens. *Br. Poult. Sci.* 47(1), 43-49.
822 doi:[10.1080/00071660500475541](https://doi.org/10.1080/00071660500475541).

823 Cashman, K.D., 2020. Vitamin D deficiency: defining, prevalence, causes, and strategies
824 of addressing. *Calcified Tissue International*, **106**, 14–29. doi:[10.1007/s00223-019-00559-4](https://doi.org/10.1007/s00223-019-00559-4).

825 Chen, Y. C., & Yu, Y. H. (2020). *Bacillus licheniformis*–fermented products improve
826 growth performance and the fecal microbiota community in broilers. *Poult. Sci.* 99(3), 1432-
827 1443. doi:[10.1016/j.psj.2019.10.061](https://doi.org/10.1016/j.psj.2019.10.061).

828 Chlebicz-Wójcik, A., Śliżewska, K., 2020. The Effect of Recently Developed Synbiotic
829 Preparations on Dominant Fecal Microbiota and Organic Acids Concentrations in Feces of
830 Piglets from Nursing to Fattening. *Animals* 10(11), 1999. doi:[10.3390/ani10111999](https://doi.org/10.3390/ani10111999).

831 Cho, J.H., Song, M.H., Kim, I.H., 2014. Effect of microencapsulated blends of organic
832 acids and essential oils supplementation on growth performance and nutrient digestibility in
833 finishing pigs. *Revista Colomb. Cienc. Pec.* 27(4), 264-272.

834 Choi, J., Kim, W.K., 2020. Dietary Application of Tannins as a Potential Mitigation
835 Strategy for Current Challenges in Poultry Production: A Review. *Animals* 10(12), 2389.
836 doi:[10.3390/ani10122389](https://doi.org/10.3390/ani10122389).

837 Choi, J., Wang, L., Liu, S., Lu, P., Zhao, X., Liu, H., Lahaye, L., Santin, E., Liu, S.,
838 Nyachoti, M., Yang, C., 2020. Effects of a microencapsulated formula of organic acids and
839 essential oils on nutrient absorption, immunity, gut barrier function, and abundance of
840 enterotoxigenic *Escherichia coli* F4 in weaned piglets challenged with *E. coli* F4. *J. Anim.*
841 *Sci.* 98(9), skaa259. doi:[10.1093/jas/skaa259](https://doi.org/10.1093/jas/skaa259).

842 Chojnacka, K., Mikulewicz, M., Cieplik, J., 2011. Biofortification of food with
843 microelements. *Am. J. Agric. Biol. Sci.* 6, 544-548.

844 Chouinard, P.Y., Corneau, L., Barbano, D.M., Metzger, L.E., Bauman, D.E., 1999.
845 Conjugated linoleic acids alter milk fatty acid composition and inhibit milk fat secretion in
846 dairy cows. *J. Nutr.* 129(8), 1579-1584. doi:[10.1093/jn/129.8.1579](https://doi.org/10.1093/jn/129.8.1579).

847 Cook, L.J., Scott, T.W., Ferguson, K.A., McDonald, I.W., 1970. Production of poly-
848 unsaturated ruminant body fats. *Nature* 228(5267), 178-179. doi:[10.1038/228178a0](https://doi.org/10.1038/228178a0).

849 Costa, M., Alves, S.P., Cappucci, A., Cook, S.R., Duarte, A., Caldeira, R.M., McAllister,
850 T. A., Bessa, R.J., 2018. Effects of condensed and hydrolyzable tannins on rumen metabolism
851 with emphasis on the biohydrogenation of unsaturated fatty acids. *Journal of Agricultural and*
852 *Food Chem.* 66(13), 3367-3377. doi:[10.1021/acs.jafc.7b04770](https://doi.org/10.1021/acs.jafc.7b04770).

853 de Carvalho Neto, J.P., Bezerra, L.R., da Silva, A.L., de Moura, J.F.P., Pereira Filho, J.M.,
854 da Silva Filho, E.C., Guedes, A.F., Araújo, M.J., Edvan, R.L., Oliveira, R.L., 2019.
855 Methionine microencapsulated with a carnauba (*Copernicia prunifera*) wax matrix for
856 protection from degradation in the rumen. *Livest. Sci.* 228, 53-60.
857 doi:[10.1016/j.livsci.2019.07.024](https://doi.org/10.1016/j.livsci.2019.07.024).

858 De Cesare, A., Sirri, F., Manfreda, G., Moniaci, P., Giardini, A., Zampiga, M., Meluzzi,
859 A., 2017. Effect of dietary supplementation with *Lactobacillus acidophilus* D2/CSL (CECT
860 4529) on caecum microbioma and productive performance in broiler chickens. *PLoS*
861 *one*, 12(5), e0176309. doi:[10.1371/journal.pone.0176309](https://doi.org/10.1371/journal.pone.0176309).

862 de Medeiros, T.T.B., de Azevedo Silva, A.M., da Silva, A.L., Bezerra, L.R., da Silva
863 Agostini, D.L., de Oliveira, D.L.V., Mazzetto, S.E., Kotzebue, L.R.V., Oliveira, R.L., 2019.
864 Carnauba wax as a wall material for urea microencapsulation. *J. Sci. Food Agric.* 99(3), 1078-
865 1087. doi:[10.1002/jsfa.9275](https://doi.org/10.1002/jsfa.9275).

866 Dschaak, C.M., Williams, C.M., Holt, M.S., Eun, J.S., Young, A.J., Min, B.R., 2011.
867 Effects of supplementing condensed tannin extract on intake, digestion, ruminal fermentation,
868 and milk production of lactating dairy cows. *J. Dairy Sci.* 94(5), 2508-2519.
869 doi:[10.3168/jds.2010-3818](https://doi.org/10.3168/jds.2010-3818).

870 EFSA Panel on Additives and Products or Substances used in Animal Feed (FEEDAP),
871 2014. Scientific Opinion on the safety and efficacy of vitamin D3 (cholecalciferol) as a feed
872 additive for all animal species or categories based on a dossier submitted by Lohmann Animal
873 Health GmbH. *EFSA J.* 12(2), 3568. doi:[10.2903/j.efsa.2014.3568](https://doi.org/10.2903/j.efsa.2014.3568).

874 EFSA Panel on Additives and Products or Substances used in Animal Feed (FEEDAP),
875 2017. Safety of vitamin D3 addition to feedingstuffs for fish. *EFSA J.* 15(3),
876 e04713. doi:[10.2903/j.efsa.2017.4713](https://doi.org/10.2903/j.efsa.2017.4713).

877 EFSA Panel on Nutrition, Novel Foods and Food Allergens (NDA), Turck, D.,
878 Castenmiller, J., De Henauw, S., Hirsch-Ernst, K. I., Kearney, J., ... & Knutsen, H. K.
879 (2021). Safety of calcidiol monohydrate produced by chemical synthesis as a novel food
880 pursuant to Regulation (EU) 2015/2283. *EFSA J.* 19(7), e06660.
881 doi:[10.2903/j.efsa.2021.6660](https://doi.org/10.2903/j.efsa.2021.6660).

882 Elmore, J.S., Cooper, S.L., Enser, M., Mottram, D.S., Sinclair, L.A., Wilkinson, R.G.,
883 Wood, J.D., 2005. Dietary manipulation of fatty acid composition in lamb meat and its effect
884 on the volatile aroma compounds of grilled lamb. *Meat Sci.* 69(2), 233-
885 242. doi:[10.1016/j.meatsci.2004.07.002](https://doi.org/10.1016/j.meatsci.2004.07.002).

886 Farm Animal Welfare Council (FAWC), 2014. Report on evidence and farm animal
887 welfare, part one, the evidence base. Farm Animal Welfare Council, London.

888 Favaretto, J.A., Alba, D.F., Marchiori, M.S., Marcon, H.J., Souza, C.F., Baldissera, M.D.,
889 ... Da Silva, A.S., 2020. Supplementation with a blend based on micro-encapsulated
890 carvacrol, thymol, and cinnamaldehyde in lambs feed inhibits immune cells and improves
891 growth performance. *Livest. Sci.* 240, 104144. doi:[10.1016/j.livsci.2020.104144](https://doi.org/10.1016/j.livsci.2020.104144).

892 Francisco, A., Dentinho, M.T., Alves, S.P., Portugal, P.V., Fernandes, F., Sengo, S., ...
893 Santos-Silva, J., 2015. Growth performance, carcass and meat quality of lambs supplemented
894 with increasing levels of a tanniferous bush (*Cistus ladanifer* L.) and vegetable oils. *Meat Sci.*
895 100, 275-282. doi:[10.1016/j.meatsci.2014.10.014](https://doi.org/10.1016/j.meatsci.2014.10.014).

896 Fuller, M.F., 2004. *The Encyclopedia of Farm Animal Nutrition*; Ed.; CABI Pub:
897 Wallingford, Oxon; Cambridge, MA, 2004; ISBN 978-0-85199-369-0.

898 Gaillard, C., Bhatti, H. S., Novoa-Garrido, M., Lind, V., Roleda, M.Y., Weisbjerg, M.R.,
899 2018. Amino acid profiles of nine seaweed species and their in situ degradability in dairy
900 cows. *Anim. Feed Sci. Technol.*, 241, 210-222. doi:[10.1016/j.anifeedsci.2018.05.003](https://doi.org/10.1016/j.anifeedsci.2018.05.003).

901 Galli, G.M., Aniecevski, E., Petrolli, T. G., da Rosa, G., Boiago, M.M., Simões, C.A., ...Da
902 Silva, A.S., 2020a. Growth performance and meat quality of broilers fed with
903 microencapsulated organic acids. *Anim. Feed Sci. Technol.* 271, 114706.
904 doi:[10.1016/j.anifeedsci.2020.114706](https://doi.org/10.1016/j.anifeedsci.2020.114706).

905 Galli, G.M., Gerbet, R.R., Griss, L.G., Fortuoso, B.F., Petrolli, T.G., Boiago, M.M., ... Da
906 Silva, A.S., 2020. Combination of herbal components (curcumin, carvacrol, thymol,

907 cinnamaldehyde) in broiler chicken feed: Impacts on response parameters, performance, fatty
908 acid profiles, meat quality and control of coccidia and bacteria. *Microb. Pathog.* 139, 103916.
909 doi:[10.1016/j.micpath.2019.103916](https://doi.org/10.1016/j.micpath.2019.103916).

910 Gheisar, M.M., Hosseindoust, A., Kim, I.H., 2015. Evaluating the effect of
911 microencapsulated blends of organic acids and essential oils in broiler chicken's diet. *J. Appl.*
912 *Poult. Res* 24(4), 511-519. doi:[10.3382/japr/pfv063](https://doi.org/10.3382/japr/pfv063).

913 Girard, C.L., Matte, J.J., 2005. Effects of intramuscular injections of vitamin B12 on
914 lactation performance of dairy cows fed dietary supplements of folic acid and rumen-
915 protected methionine. *J. Dairy Sci.* 88(2), 671-676. doi:[10.3168/jds.S0022-0302\(05\)72731-4](https://doi.org/10.3168/jds.S0022-0302(05)72731-4).

916 Gomathi, G., Senthilkumar, S., Natarajan, A., Amutha, R., Purushothaman, M.R., 2018.
917 Effect of dietary supplementation of cinnamon oil and sodium butyrate on carcass
918 characteristics and meat quality of broiler chicken. *Vet. World* 11(7), 959-964.
919 doi:[10.14202/vetworld.2018.959-964](https://doi.org/10.14202/vetworld.2018.959-964).

920 Gonzalez-Esquerria, R., Leeson, S., 2001. Alternatives for enrichment of eggs and chicken
921 meat with omega-3 fatty acids. *Canadian J. Anim. Sci.* 81(3), 295-305. doi:[10.4141/A00-092](https://doi.org/10.4141/A00-092).

922 Grilli, E., Gallo, A., Fustini, M., Fantinati, P., Piva, A., 2013. Microencapsulated sodium
923 selenite supplementation in dairy cows: effects on selenium status. *Animal* 7(12), 1944-1949.
924 doi:[10.1017/S1751731113001547](https://doi.org/10.1017/S1751731113001547).

925 Grilli, E., Messina, M. R., Tedeschi, M., Piva, A., 2010. Feeding a microencapsulated
926 blend of organic acids and nature identical compounds to weaning pigs improved growth
927 performance and intestinal metabolism. *Livest. Sci.* 133(1-3), 173-175.
928 doi:[10.1016/j.livsci.2010.06.056](https://doi.org/10.1016/j.livsci.2010.06.056).

929 Guasch, I., Elcoso, G., Zweifel, B., Bach, A., 2016. Effects of a blend of essential oils on
930 milk yield and feed efficiency of lactating cows. *J. Anim. Sci.* 94 (12), 718-719.
931 doi:[10.2527/jam2016-1480](https://doi.org/10.2527/jam2016-1480).

932 Gulati, S.K., Kitessa, S.M., Ashes, J.R., Fleck, E., Byers, E.B., Byers, Y.G., Scott, T.W.,
933 2000. Protection of conjugated linoleic acids from ruminal hydrogenation and their
934 incorporation into milk fat. *Anim. Feed Sci. Technol.* 86(3-4), 139-148. doi:[10.1016/S0377-](https://doi.org/10.1016/S0377-8401(00)00170-X)
935 [8401\(00\)00170-X](https://doi.org/10.1016/S0377-8401(00)00170-X).

936 Guo, T., Sanchez, M. D., Guo, P. (Eds.), 2002. Animal production based on crop residues:
937 Chinese experiences. Food and Agriculture Organization of the United Nations, Eds.; FAO
938 animal production and health paper; Food and Agriculture Organization of the United
939 Nations: Rome, ISBN 978-92-5-104639-5.

940 Heck, R.T., Lorenzo, J.M., Dos Santos, B.A., Cichoski, A.J., de Menezes, C.R.,
941 Campagnol, P.C.B., 2020. Microencapsulation of healthier oils: an efficient strategy to
942 improve the lipid profile of meat products. *Curr. Opin. Food Sci.* 40, 6–12.
943 doi:[10.1016/j.cofs.2020.04.010](https://doi.org/10.1016/j.cofs.2020.04.010).

944 Hu, Z.P., Wang, T., Ahmad, H., Zhang, J. F., Zhang, L. L., Zhong, X., 2015. Effects of
945 different formulations of α -tocopherol acetate (vitamin E) on growth performance, meat
946 quality and antioxidant capacity in broiler chickens. *Br. Poul. Sci.* 56(6), 687-695.
947 doi:[10.1080/00071668.2015.1080814](https://doi.org/10.1080/00071668.2015.1080814).

948 Hunter, P., 2011. Nutrition: more than the sum of its parts: The modern craze for dietary
949 supplements is under increasing scrutiny, while biofortified crops look promising in the quest
950 to deliver nutrition in developing countries. *EMBO Reports* 12(4), 307-310.
951 doi:[10.1038/embor.2011.42](https://doi.org/10.1038/embor.2011.42).

952 Jahromi, M.F., Altaher, Y.W., Shokryazdan, P., Ebrahimi, R., Ebrahimi, M., Idrus, Z.,
953 Tufarelli, V., Liang, J. B. 2016. Dietary supplementation of a mixture of *Lactobacillus* strains
954 enhances performance of broiler chickens raised under heat stress conditions. *Int. J.*
955 *Biometeorol.* 60(7), 1099-1110. doi:[10.1007/s00484-015-1103-x](https://doi.org/10.1007/s00484-015-1103-x).

956 Joysowal, M., Tyagi, A. K., Tyagi, N., Kumar, S., Keshri, A. 2019. Use of slow release
957 ammonia products in ruminant diet: A review. *J. Entomol. Zool. Stud.*, 7, 882-888.

958 Kim, T.B., Lee, J.S., Cho, S.Y., Lee, H.G., 2020. In vitro and in vivo studies of rumen-
959 protected microencapsulated supplement comprising linseed oil, vitamin E, Rosemary extract,
960 and hydrogenated palm oil on rumen fermentation, physiological profile, milk yield, and milk
961 composition in dairy cows. *Animals* 10(9), 1631. doi:[10.3390/ani10091631](https://doi.org/10.3390/ani10091631).

962 Konkol, D., Wojnarowski, K., 2018. The use of nanominerals in animal nutrition as a way
963 to improve the composition and quality of animal products. *J. Chem.* 2018.
964 doi:[10.1155/2018/5927058](https://doi.org/10.1155/2018/5927058).

965 Korczynski, M., Kupczynski, R., Swiniarska, M., Konkol, D., Opalinski. S., 2017.
966 Fortification of animal's foodstuff in Food Biofortification Technologies, A. Saeid, Ed., pp.
967 273–312, CRC Press, Boca Raton, FL, USA.

968 Lawlor, J.B., Gaudette, N., Dickson, T., House, J.D., 2010. Fatty acid profile and sensory
969 characteristics of table eggs from laying hens fed diets containing microencapsulated fish
970 oil. *Anim. Feed Sci. Technol.* 156(3-4), 97-103. doi:[10.1016/j.anifeedsci.2010.01.003](https://doi.org/10.1016/j.anifeedsci.2010.01.003).

971 Lee, J.H., Waller, J.C., Melton, S.L., Saxton, A.M., Pordesimo, L.O., 2004. Feeding
972 encapsulated ground full-fat soybeans to increase polyunsaturated fat concentrations and
973 effects on flavor volatiles in fresh lamb. *J. Anim. Sci.* 82(9), 2734-2741.
974 doi:[10.2527/2004.8292734x](https://doi.org/10.2527/2004.8292734x).

975 Lee, J.H., Waller, J.C., Yilmaz, Y., Melton, S.L., 2007. Effect of feeding rumen-protected
976 dietary protein–oil supplements on fatty acid composition and α -tocopherol content of blood
977 serum and muscle lipids of lambs. *Small Ruminant Res.* 72(2-3), 101-110.
978 doi:[10.1016/j.smallrumres.2006.08.012](https://doi.org/10.1016/j.smallrumres.2006.08.012).

979 Lehnen, T.E., da Silva, M.R., Camacho, A., Marcadenti, A., Lehnen, A M., 2015. A review
980 on effects of conjugated linoleic fatty acid (CLA) upon body composition and energetic
981 metabolism. *J. Int. Soc. Sports Nutr.* 12(1), 1-11. doi:[10.1186/s12970-015-0097-4](https://doi.org/10.1186/s12970-015-0097-4).

982 Rabanel, M., Aoun, J., Elkin, V., Mokhtar I., Hildgen, P., 2012. Drug-loaded nanocarriers:
983 passive targeting and crossing of biological barriers. *Curr. Med. Chem.* 19(19), 3070-3102.
984 doi:[10.2174/092986712800784702](https://doi.org/10.2174/092986712800784702).

985 Maenner, K., Vahjen, W., Simon, O., 2011. Studies on the effects of essential-oil-based
986 feed additives on performance, ileal nutrient digestibility, and selected bacterial groups in the
987 gastrointestinal tract of piglets. *J. Anim. Sci.* 89(7), 2106-2112. doi:[10.2527/jas.2010-2950](https://doi.org/10.2527/jas.2010-2950).

988 Mamvura, C.I., Cho, S., Mbiriri, D.T., Lee, H.G., Choi, N.J., 2014. Effect of encapsulating
989 nitrate in sesame gum on in vitro rumen fermentation parameters. *Asian-Australasian J.*
990 *Anim. Sci.* 27(11), 1577. doi:[10.5713/ajas.2014.14280](https://doi.org/10.5713/ajas.2014.14280).

991 McCrorie, T.A., Keaveney, E.M., Wallace, J.M., Binns, N., Livingstone, M.B.E., 2011.
992 Human health effects of conjugated linoleic acid from milk and supplements. *Nutr. Res. Rev.*
993 24(2), 206-227. doi:[10.1017/S0954422411000114](https://doi.org/10.1017/S0954422411000114).

994 Mcguire, M.A., Mcguire, M.K., 2000. Conjugated linoleic acid (CLA): a ruminant fatty
995 acid with beneficial effects on human health. *J. Anim. Sci.* 77, 1-8.
996 doi:[10.2527/jas2000.00218812007700ES0033x](https://doi.org/10.2527/jas2000.00218812007700ES0033x).

997 Meimandipour, A., Nouri Emamzadeh, A., Soleimani, A., 2017. Effects of
998 nanoencapsulated aloe vera, dill and nettle root extract as feed antibiotic substitutes in broiler
999 chickens. *Arch. Anim. Breed.* 60(1), 1-7. doi:[10.5194/aab-60-1-2017](https://doi.org/10.5194/aab-60-1-2017).

1000 Munin, A., Edwards-Lévy, F., 2011. Encapsulation of natural polyphenolic compounds; a
1001 review. *Pharmaceutics* 3(4), 793-829. doi:[10.3390/pharmaceutics3040793](https://doi.org/10.3390/pharmaceutics3040793).

1002 Natsir, M.H., Hartutik, O.S., Widodo, E., 2013. Effect of either powder or encapsulated
1003 form of garlic and *Phyllanthus niruri* L. mixture on broiler performances, intestinal

1004 characteristics and intestinal microflora. *International J. Poult. Sci.* 12(11), 676-
1005 680. doi:[10.3923/ijps.2013.676.680](https://doi.org/10.3923/ijps.2013.676.680).

1006 Natsir, M.H., Sjofjan, O., Widodo, E., Ardiansah, I., Widyastuti, E.S., 2019. Effect of
1007 either non-encapsulated or encapsulated acidifier-phytobiotic-probiotic on performance,
1008 intestinal characteristics and intestinal microflora of local hybrid ducks. *Livestock Res. Rural*
1009 *Dev.*31(1), 1-5.

1010 O’Keeffe, P., 2020. Nutrition for plant-based diets: managing nutrient intake and
1011 bioavailability, KHNI (The Kerry Health and Nutrition Institute).

1012 Park, S., Park, B., 2011. Effect of dietary microencapsulated-inulin on carcass
1013 characteristics and growth performance in broiler chickens. *J. Anim. Vet. Adv.* 10(10), 1342-
1014 1349. doi:[10.3923/javaa.2011.1342.1349](https://doi.org/10.3923/javaa.2011.1342.1349).

1015 Partenen, K., 2001. Organic acids - Their efficacy and modes of action in pigs. in *Gut*
1016 *environment of pigs*. Piva, A., K. E. Bach Knudsen, and J. E. Lindberg, eds. Nottingham
1017 Univ. Press, Nottingham, UK.

1018 Patra, A. K., Saxena, J., 2011. Exploitation of dietary tannins to improve rumen
1019 metabolism and ruminant nutrition. *J. Sci. Food Agric.* 91(1), 24-37. doi:[10.1002/jsfa.4152](https://doi.org/10.1002/jsfa.4152).

1020 Perfield II, J.W., Lock, A.L., Pfeiffer, A.M., Bauman, D.E., 2004. Effects of amide-
1021 protected and lipid-encapsulated conjugated linoleic acid (CLA) supplements on milk fat
1022 synthesis. *J. Dairy Sci.* 87(9), 3010-3016. doi:[10.3168/jds.S0022-0302\(04\)73432-3](https://doi.org/10.3168/jds.S0022-0302(04)73432-3).

1023 Peterson, D.G., Baumgard, L.H., Bauman, D.E., 2002. Milk fat response to low doses of
1024 trans-10, cis-12 conjugated linoleic acid (CLA). *J. Dairy Sci.* 85(7), 1764-1766.
1025 doi:[10.3168/jds.S0022-0302\(02\)74250-1](https://doi.org/10.3168/jds.S0022-0302(02)74250-1).

1026 Phillips, C.J.C., 2016. Introduction to welfare and nutrition. In *Nutrition and the Welfare of*
1027 *Farm Animals* (pp. 1-9). Springer International Publishing: Cham, ISBN 978-3-319-27354-9.

1028 Pinotti, L., Baldi, A., Politis, I., Rebucci, R., Sangalli, L., Dell'Orto, V., 2003. Rumen-
1029 protected choline administration to transition cows: effects on milk production and vitamin E
1030 status. *J. Vet. Med.* 50(1), 18-21. doi:[10.1046/j.1439-0442.2003.00502.x](https://doi.org/10.1046/j.1439-0442.2003.00502.x).

1031 Piva, A., Pizzamiglio, V., Morlacchini, M., Tedeschi, M., Piva, G., 2007. Lipid
1032 microencapsulation allows slow release of organic acids and natural identical flavors along
1033 the swine intestine. *J. Anim. Sci.* 85(2), 486-493. doi:[10.2527/jas.2006-323](https://doi.org/10.2527/jas.2006-323).

1034 Prasad, A. S., 2013. Discovery of human zinc deficiency: its impact on human health and
1035 disease. *Adv. Nutr.* 4(2), 176-190. doi:[10.3945/an.112.003210](https://doi.org/10.3945/an.112.003210).

1036 Puga, D. C., Galina, H. M., Pérez-Gil, R. F., Sanginés, G. L., Aguilera, B. A., Haenlein, G.
1037 F. W. (2001). Effect of a controlled-release urea supplement on rumen fermentation in sheep
1038 fed a diet of sugar cane tops (*Saccharum officinarum*), corn stubble (*Zea mays*) and King
1039 grass (*Pennisetum purpureum*). *Small Rum. Res.* 39(3), 269-276. doi:[10.1016/S0921-](https://doi.org/10.1016/S0921-4488(00)00196-6)
1040 [4488\(00\)00196-6](https://doi.org/10.1016/S0921-4488(00)00196-6)

1041 Purba, R.A.P., Paengkoum, P., Paengkoum, S., 2020. The links between supplementary
1042 tannin levels and conjugated linoleic acid (CLA) formation in ruminants: A systematic review
1043 and meta-analysis. *PloS one*, 15(3), e0216187. doi:[10.1371/journal.pone.0216187](https://doi.org/10.1371/journal.pone.0216187).

1044 Rogers, J.A., Krishnamoorthy, U., Sniffen, C.J., 1987. Plasma amino acids and milk
1045 protein production by cows fed rumen-protected methionine and lysine. *J.Dairy Sci.*70(4),
1046 789-798. doi:[10.3168/jds.S0022-0302\(87\)80075-9](https://doi.org/10.3168/jds.S0022-0302(87)80075-9).

1047 Romero-Pérez, A., García-García, E., Zavaleta-Mancera, A., Ramírez-Bribiesca, J.E.,
1048 Revilla-Vázquez, A., Hernández-Calva, L.M., López-Arellano, R., Cruz-Monterrosa, R.G.,
1049 2010. Designing and evaluation of sodium selenite nanoparticles in vitro to improve selenium
1050 absorption in ruminants. *Vet. Res. Commun.* 34(1), 71-79. doi:[10.1007/s11259-009-9335-z](https://doi.org/10.1007/s11259-009-9335-z).

1051 Sacadura, F.C., Robinson, P.H., Evans, E., Lordelo, M., 2008. Effects of a ruminally
1052 protected B-vitamin supplement on milk yield and composition of lactating dairy cows. *Anim.*
1053 *Feed Sci. Technol.* 144(1-2), 111-124. doi:[10.1016/j.anifeedsci.2007.10.005](https://doi.org/10.1016/j.anifeedsci.2007.10.005).

1054 Sagalowicz, L., Leser, M.E., 2010. Delivery systems for liquid food products. *Curr. Opin.*
1055 *Colloid In.* 15(1-2), 61-72. doi:[10.1016/j.cocis.2009.12.003](https://doi.org/10.1016/j.cocis.2009.12.003).

1056 Santos, A.R.M.D., Barros, L.V.D., Abreu, M.L.C., Pedreira, B.C., 2020. In vitro ruminal
1057 fermentation parameters and methane production of Marandu palisadegrass (*Urochloa*
1058 *brizantha*) in a silvopastoral system associated with levels of protein supplementation. *Grass*
1059 *Forage Sci.*75(3), 339-350. doi:[10.1111/gfs.12476](https://doi.org/10.1111/gfs.12476).

1060 Santschi, D.E., Berthiaume, R., Matte, J.J., Mustafa, A.F., Girard, C.L., 2005. Fate of
1061 supplementary B-vitamins in the gastrointestinal tract of dairy cows. *J. Dairy Sci.* 88(6),
1062 2043-2054. doi:[10.3168/jds.S0022-0302\(05\)72881-2](https://doi.org/10.3168/jds.S0022-0302(05)72881-2).

1063 Sevi, A., Rotunno, T., Di Caterina, R., Muscio, A., 1998. Rumen-protected methionine or
1064 lysine supplementation of Comisana ewes' diets: effects on milk fatty acid composition. *J.*
1065 *Dairy Res.* 65(3), 413-422. doi:[10.1017/S0022029998002945](https://doi.org/10.1017/S0022029998002945).

1066 Śliwiński, B.J., Soliva, C.R., Machmüller, A., Kreuzer, M., 2002. Efficacy of plant extracts
1067 rich in secondary constituents to modify rumen fermentation. *Anim. Feed Sci.*
1068 *Technol.* 101(1-4), 101-114. doi:[10.1016/S0377-8401\(02\)00139-6](https://doi.org/10.1016/S0377-8401(02)00139-6).

1069 Słupczyńska, M., Jamroz, D., Orda, J., Wiliczekiewicz, A., 2014. Effect of various sources
1070 and levels of iodine, as well as the kind of diet, on the performance of young laying hens,
1071 iodine accumulation in eggs, egg characteristics, and morphotic and biochemical indices in
1072 blood. *Poult. Sci.* 93(10), 2536-2547. doi:[10.3382/ps.2014-03959](https://doi.org/10.3382/ps.2014-03959).

1073 Silva, F. C., Lima, L. C., Viseras, C., Osajima, J. A., da Silva Júnior, J. M., Oliveira, R. L.,
1074 ... Silva-Filho, E. C. (2019). Understanding Urea Encapsulation in Different Clay Minerals as

1075 a Possible System for Ruminant Nutrition. *Molecules* 24(19), 3525. doi:
1076 10.3390/molecules24193525.

1077 Socha, M.T., Putnam, D.E., Garthwaite, B.D., Whitehouse, N.L., Kierstead, N.A., Schwab,
1078 C.G., Ducharme, G.A., Robert, J.C., 2005. Improving intestinal amino acid supply of pre-and
1079 postpartum dairy cows with rumen-protected methionine and lysine. *J. Dairy Sci.*88(3), 1113-
1080 1126. doi:[10.3168/jds.S0022-0302\(05\)72778-8](https://doi.org/10.3168/jds.S0022-0302(05)72778-8).

1081 Spanghero, M., Robinson, P.H., Zanfi, C., Fabbro, E., 2009. Effect of increasing doses of a
1082 microencapsulated blend of essential oils on performance of lactating primiparous dairy
1083 cows. *Anim. Feed Sci. Technol.* 153(1-2), 153-157. doi:[10.1016/j.anifeedsci.2009.06.004](https://doi.org/10.1016/j.anifeedsci.2009.06.004).

1084 Stamilla, A., Russo, N., Messina, A., Spadaro, C., Natalello, A., Caggia, C., Randazzo, C.
1085 L., Lanza, M., 2020. Effects of Microencapsulated Blend of Organic Acids and Essential Oils
1086 as a Feed Additive on Quality of Chicken Breast Meat. *Animals* 10(4),1-17.
1087 doi:[10.3390/ani10040640](https://doi.org/10.3390/ani10040640).

1088 Stevanović, Z. D., Bošnjak-Neumüller, J., Pajić-Lijaković, I., Raj, J., Vasiljević, M., 2018.
1089 Essential oils as feed additives—future perspectives. *Molecules* 23(7), 1717.
1090 doi:[10.3390/molecules23071717](https://doi.org/10.3390/molecules23071717).

1091 Suiryanrayna, M.V., Ramana, J.V., 2015. A review of the effects of dietary organic acids
1092 fed to swine. *J. Anim. Sci. and Biotechnology*, 6(1), 1-11. doi:[10.1186/s40104-015-0042-z](https://doi.org/10.1186/s40104-015-0042-z).

1093 Sundari, Z., Yuwanta, T., Martien, R., 2014. Effect of nanocapsule level on broiler
1094 performance and fat deposition. *Int. J. Poult. Sci.* 13 (1), 31–35. doi:[10.3923/ijps.2014.31.35](https://doi.org/10.3923/ijps.2014.31.35).

1095 Swiatkiewicz, S., Arczewska-Wlosek, A., 2012. Prebiotic fructans and organic acids as
1096 feed additives improving mineral availability. *Poult. Sci. J.* 68(2), 269-279.
1097 doi:[10.1017/S0043933912000323](https://doi.org/10.1017/S0043933912000323).

1098 Sýkora, T., Rabišková, M., Třináctý, J., Vetchý, D., Häring, A., Dvořák, P., 2007.
1099 Postprandial delivery system for amino acids and proteins in cattle. *Acta Vet. Brno* 76(4),
1100 547-552. doi:[10.2754/avb200776040547](https://doi.org/10.2754/avb200776040547).

1101 Tao, W.J., Liu, L.J., Li, H., Pei, X., Wang, G., Xiao, Z.P., Xiao Z.P., Yu, R., Li, Z.F.,
1102 Wang, M.Q., 2020. Effects of coated cysteamine on growth performance, carcass
1103 characteristics, meat quality and lipid metabolism in finishing pigs. *Anim. Feed Sci.*
1104 *Technol.* 263, 1-5. doi:[10.1016/j.anifeedsci.2020.114480](https://doi.org/10.1016/j.anifeedsci.2020.114480).

1105 Temiz, U., Öztürk, E., 2018. Encapsulation methods and use in animal nutrition. *Selcuk*
1106 *Journal of Agriculture and Food Sciences*, 32(3), 624-631. doi:[10.15316/SJAIFS.2018.145](https://doi.org/10.15316/SJAIFS.2018.145).

1107 Thavarajah, P., Wejesuriya, A., Rutzke, M., Glahn, R.P., Combs, G.F., Vandenberg, A.,
1108 2011. The potential of lentil (*Lens culinaris* L.) as a whole food for increased selenium, iron,
1109 and zinc intake: preliminary results from a 3-year study. *Euphytica* 180(1), 123-128.
1110 doi:[10.1007/s10681-011-0365-6](https://doi.org/10.1007/s10681-011-0365-6).

1111 Tolve, R., Galgano, F., Caruso, M. C., Tchienbou-Magaia, F. L., Condelli, N., Favati, F.,
1112 Zhang, Z., 2016. Encapsulation of health-promoting ingredients: applications in
1113 foodstuffs. *International Int. J. Food Sci. Nutr.* 67(8), 888-918.
1114 doi:[10.1080/09637486.2016.1205552](https://doi.org/10.1080/09637486.2016.1205552).

1115 Tolve, R., Cela, N., Condelli, N., Di Cairano, M., Caruso, M. C., Galgano, F., 2020.
1116 Microencapsulation as a tool for the formulation of functional foods: The phytosterols' case
1117 study. *Foods* 470, 1-19. doi:[10.3390/foods9040470](https://doi.org/10.3390/foods9040470).

1118 Tolve, R., Galgano, F., Condelli, N., Cela, N., Lucini, L., Caruso, M.C., 2021.
1119 Optimization model of phenolics encapsulation conditions for biofortification in fatty acids of
1120 animal food products. *Foods* 2021. doi.org/10.3390/foods10040881.

1121 Tugnoli, B., Giovagnoni, G., Piva, A., Grilli, E., 2020. From acidifiers to intestinal health
1122 enhancers: How organic acids can improve growth efficiency of pigs. *Animals* 10(1), 134.
1123 doi:[10.3390/ani10010134](https://doi.org/10.3390/ani10010134).

1124 Turek, C., Stintzing, F.C., 2013. Stability of essential oils: a review. *Compr. Rev. Food*
1125 *Sci. F.* 12(1), 40-53. doi:[10.1111/1541-4337.12006](https://doi.org/10.1111/1541-4337.12006).

1126 United Nations, 2019. Department of Economic and Social Affairs; Population Division.
1127 World Population Prospects 2019: Highlights; UN: New York, NY, USA, 2019; ISBN 978-
1128 92-1-148316-1.

1129 Upadhaya, S.D., Lee, K.Y., Kim, I.H., 2014. Protected organic acid blends as an
1130 alternative to antibiotics in finishing pigs. *Asian-Australasian J. Anim. Sci.*, 27(11), 1600-
1131 1607. doi:[10.5713/ajas.2014.14356](https://doi.org/10.5713/ajas.2014.14356).

1132 Vanegas-Espinoza, P.E., Pérez-Escalante, V., Aguirre-Guzman, G., Hoyos-Leyva, J.D.,
1133 Del Villar-Martínez, A.A., 2019. Microencapsulation of anthocyanins from roselle (*Hibiscus*
1134 *sabdariffa*) and its application on a pigment supplied diet to fantail goldfish (*Carassius*
1135 *auratus*). *Aquac. Int.* 27(6), 1801-1811. doi:[10.1007/s10499-019-00430-1](https://doi.org/10.1007/s10499-019-00430-1).

1136 Vanhatalo, A., Huhtanen, P., Toivonen, V., Varvikko, T. 1999. Response of dairy cows fed
1137 grass silage diets to abomasal infusions of histidine alone or in combinations with methionine
1138 and lysine. *J. Dairy Sci.*, 82(12), 2674-2685. doi: 10.3168/jds.S0022-0302(99)75524-4.

1139 Vasta, V., Daghighi, M., Cappucci, A., Buccioni, A., Serra, A., Viti, C., Mele, M., 2019.
1140 Invited review: Plant polyphenols and rumen microbiota responsible for fatty acid
1141 biohydrogenation, fiber digestion, and methane emission: Experimental evidence and
1142 methodological approaches. *J. Dairy Sci.* 102(5), 3781-3804. doi:[10.3168/jds.2018-14985](https://doi.org/10.3168/jds.2018-14985).

1143 Willer, D.F., Aldridge, D.C., 2020. Vitamin bullets. microencapsulated feeds to fortify
1144 shellfish and tackle human nutrient deficiencies. *Front. Nutr.* 7, 102.
1145 doi:[10.3389/fnut.2020.00102](https://doi.org/10.3389/fnut.2020.00102).

1146 Williams, L.R., Martz, F.A., Hilderbrand, E.S., 1970. Feeding encapsulated methionine
1147 supplement to lactating cows. *J. Dairy Sci.* 53(12), 1709-1713. doi:[10.3168/jds.S0022-](https://doi.org/10.3168/jds.S0022-0302(70)86467-0)
1148 [0302\(70\)86467-0](https://doi.org/10.3168/jds.S0022-0302(70)86467-0).

1149 World Health Organization (WHO), 2020. Micronutrient deficiencies. Available:
1150 <https://www.who.int/nutrition/topics/vad/en/>.

1151 Xin, H.S., Schaefer, D.M., Liu, Q.P., Axe, D.E., Meng Q.X. 2010. Effects of polyurethane
1152 coated urea supplement on in vitro ruminal fermentation, ammonia release dynamics and
1153 lactating performance of Holstein dairy cows fed a steam-flaked corn-based diet. *Asian-Aust.*
1154 *J. Anim. Sci.*, 23: 491-500.

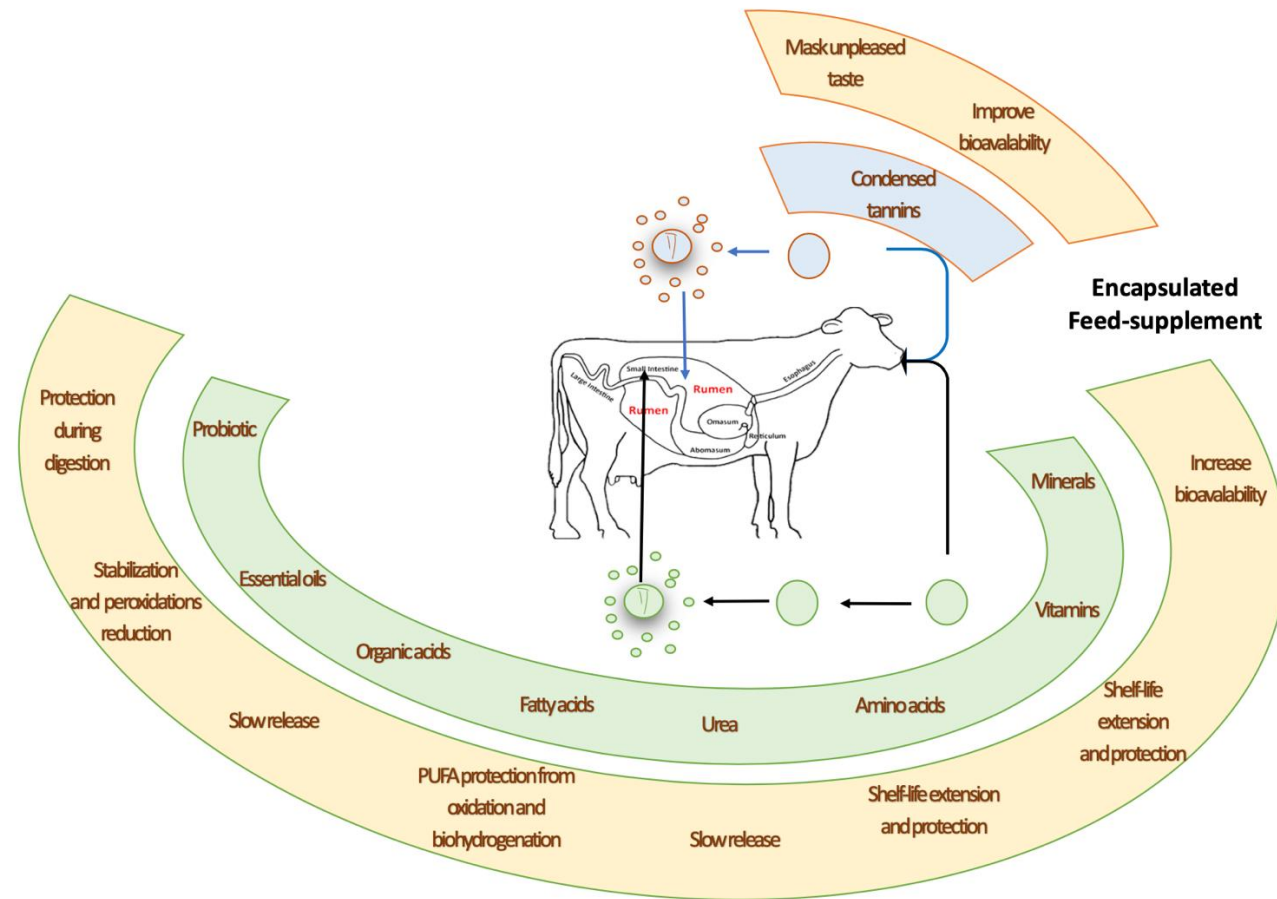
1155 Zhang, Y., Gong, J., Yu, H., Guo, Q., Defelice, C., Hernandez, M., Yin, Y., Wang, Q.
1156 2014. Alginate-whey protein dry powder optimized for target delivery of essential oils to the
1157 intestine of chickens. *Poult. Sci.* 93(10), 2514-2525. doi:[10.3382/ps.2013-03843](https://doi.org/10.3382/ps.2013-03843).

1158 Zang, Y., Silva, L. H. P., Ghelichkhan, M., Miura, M., Whitehouse, N. L., Chizzotti, M. L.,
1159 Brito, A. F. 2019. Incremental amounts of rumen-protected histidine increase plasma and
1160 muscle histidine concentrations and milk protein yield in dairy cows fed a metabolizable
1161 protein-deficient diet. *J. Dairy. Sci.* 102(5), 4138-4154. doi: [10.3168/jds.2018-15780](https://doi.org/10.3168/jds.2018-15780).

1162 Zimbelman, R.B., Collier, R.J., Bilby, T.R., 2013. Effects of utilizing rumen protected
1163 niacin on core body temperature as well as milk production and composition in lactating dairy
1164 cows during heat stress. *Anim. Feed Sci. Technol.* 180(1-4), 26-
1165 33. doi:[10.1016/j.anifeedsci.2013.01.005](https://doi.org/10.1016/j.anifeedsci.2013.01.005).

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Figure 1. Schematic diagram showing the protecting effect and the site-specific release of the encapsulated bioactive compounds. Generally, the feed supplements that need to be protected from the environment and the microbial degradation that could occur in the rumen are protected by coating materials with the ability to degrade a pH below 5, thus are a candidate for release in small the intestine. The bioactive compounds characterized by unpleasant taste generally are encapsulated in coating materials that can mask the taste.

Table 1. Encapsulated minerals with potential in animal feed

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Encapsulation method	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
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Sodium selenite	Increase of the mineral content of milk	Protect minerals from the transformation in insoluble compounds in rumen	Spray-chilling	Hydrogenated fat	From 500 to 2000 μm	Microcapsules in order to provide 0.3 and 0.5 mg/kg DM of total Se	Cows fed with microencapsulated Se at 0.3 mg/kg had 38% higher milk Se content than control animals	Grilli et al. (2013)
Sodium selenite and zinc carbonate	Increase of the mineral content of meat	Protect minerals from the transformation in insoluble compounds in rumen	Emulsion-evaporation	Eudragit RL	From 500 to 2000 μm	0.6 $\mu\text{g g}^{-1}$ Se + 30.0 $\mu\text{g g}^{-1}$ Zn nanoparticles	Selenium and Zn nanoparticles increased the concentration of these minerals in lam's meat as compared with supplementation of raw Se and Zn	Bribiesca et al. (2017)
Sodium selenite	Increase of the mineral content of milk/meat	Protect minerals from the transformation in insoluble compounds in rumen	Emulsion-evaporation and nano-precipitation	Eudragit® RL and RS	From 36.64 to 213.86 nm	No <i>in vivo</i> study	The release of selenium from nanoparticles was higher at acid pH (less than 4); this condition may represent a better availability of the mineral in the small intestine	Romero-Pérez. et al. (2010)

Table 2. Encapsulated vitamins with potential in animal feed

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Encapsulation method	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
Vitamin E	Improvement of growth and performance	Increase of the overall bio-accessibility of vitamin E	NR	Sodium octenyl succinate	NR	20 mg/kg	Microencapsulated vitamin E was effective in increasing its bio-accessibility with a positive impact on stabilizing breast broiler and yielded better broiler growth performance (weight gain) when compared to non-encapsulated vitamin E fed groups	Hu et al. (2015)
Vitamin A	Beneficial effect on the immune, the central nervous system, reproductive performance, and normal bone growth	Vitamin protection	NR	Lecithin, cholesterol and functionalized stearyls	NR	NR	Retards of the vitamin A degradation	Albright and Kowarski (1994)
α -tocopheryl-acetate	Increase vitamin E concentration in meat	Vitamin protection	NR	Sodium caseinate	NR	250 IU	Higher α -tocopherol concentrations in psoas major muscle than lambs fed with nonencapsulated vitamin E	Lee et al. (2007)
Vitamin B (biotin, folic acid, pantothenic acid and pyridoxine)	Impact on health and milk production	Vitamin protection	NR	NR	NR	3 g/d	Supplying cows with a ruminally protected B vitamin blend increased milk and milk component yields, especially milk protein	Sacadura et al. (2008)

NR= commercial product/blend; information not reported

(continued)

Table 2. Continued

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Encapsulation method	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
Vitamin B (niacin)	Increase of resistance to thermal stress	Increase vitamin absorption	NR	Lipidic shell	NR	12 g/d	Supplementation of lactating cows with encapsulated niacin reduced cow's body temperature	Zimbelma et al. (2013)

NR= commercial product/blend; information not reported

Table 3. Continued

Table 3. Encapsulated amino acids and urea with potential in animal feed

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Technique of encapsulation	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
Cysteamine	Beneficial effect on animal's growth performance	Cysteamine protection from the oxidation	NR	Palm stearin (vegetal)	NR	27, 54, or 108 mg/kg cysteamine	Microencapsulated cysteamine significantly increased carcass lean percentage and <i>longissimus muscle</i> area	Tao et al. (2020)
Methionine and Lysine	Improvement in milk or milk protein production	Delivery of amino acids in duodenal sites	NR	NR	NR	10.2 g/d of methionine and 16.0 g/d of lysine	Cows produced milk with more true protein (1306 vs. 1221 g/d), and fat (1632 vs. 1550 g/d)	Socha et al. (2005)
Methionine	Improvement in milk or milk protein production	Protection of methionine from the ruminal environment	Melt-emulsification technique	Carnauba wax	NR	No <i>in vivo</i> study	Efficient ruminal protection	Carvalho Neto et al. (2019)
Lysine	Improvement in milk or milk protein production	Protection of lysine from the ruminal environment	Organic solvent, free emulsification, sonication method	Stearic acid and SPAN 80	From 200 to 500 nm	No <i>in vivo</i> study	Solid lipid nanoparticles composed of arachidic or stearic acids and Tween 60 resisted ruminal digestion for up to 24 h	Albuquerque et al. (2020)

NR= commercial product/blend; information not reported

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Encapsulati-on method	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
Histidine	Improvement in milk or milk protein production	Protection of histidine from the ruminal environment	NR	NR	NR	164 g/d	Supplementation of lactating cows with encapsulated histidine significantly increased the amino acid concentration in plasma and muscle. The milk true protein increased as well.	Zhang et al. (2019)
Urea	Urea is the main source of nonprotein nitrogen used in the ruminant diet	Slow release	NR	Polyuret hane	NR	1.7%	Coated urea increased the ruminant dry matter intake and the nutrients digestibility compared to the uncoated urea.	Xin et al. (2010)

0 NR= commercial product/blend; information not reported

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Table 4. Encapsulated fatty acids with potential in animal feed

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Technique of encapsulation	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
Sunflower oil	Reduce SFA and increase PUFA concentration in animal products	PUFA protection from biohydrogenation	In situ polymerization	Sodium caseinate	<625 mm	2 kg/day	Lambs feed with encapsulated sunflower oil and vitamin E were characterized by a higher LA levels in both <i>longissimus dorsi</i> and <i>psoas major muscles</i>	Lee et al. (2007)
Ground whole soybeans	Reduce SFA and increase PUFA concentration in animal products	PUFA protection from biohydrogenation	In situ polymerization	Casein treated with acetaldehyde or diacetyl	<625 mm	10% of DM intake	Fat of lambs supplemented with encapsulated linoleic acid had higher concentrations of linoleic (LA) and linolenic acids (ALA)	Lee et al. (2004)
Oil high in linoleic and alfa-linolenic acid	Reduce SFA and increase PUFA concentration in animal products	PUFA protection from biohydrogenation	Spray-dryer	Formaldehyde-treated protein	NR	60 g of oil containin g more than the 60% of PUFA	LA and ALA in lamb muscle of about 247% and 57%, respectively compared to those fed unprotect-ed lipids	Elmore et al. (2005)
Ground flaxseed	Reduce SFA and increase PUFA concentration in animal products	PUFA protection from biohydrogenation	High-speed turbulizer + dryer	Dolomitic lime hydrate	NR	2-4 or 6% of DM	ALA content of muscle tissue was 47% greater when flaxseed was encapsulated within the dolomitic lime hydrate matrix	Alvarado-Gilis et al. (2015b)

3 NR= commercial product/blend; information not reported

(continued)

Table 4. Continued

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Technique of encapsulation	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
Commercial pro-duct contained linseed oil, vitamin E, rosemary extract	Reduce SFA and increase PUFA concentration in ani-mal products	PUFA protection from biohydroge-nation	Spray chilling	Hydrogenated palm oil	From 1000 to 1500 µm	2% of DM	The use of the commercial microencapsulated lin-seed oils significantly in-creased the yield of total omega-3 fatty acids in milk com-pared with non-encapsulated linseed	Kim et al. (2020)
Conjugated linoleic acid (CLA)	Increase co-njugated linoleic acid con-centration in milk	CLA protection from biohydroge-nation	Freeze dryer	Formaldehyde-treated casein	NR	80 g/day	The total CLA levels were enhanced by about 10-fold above the control levels present in milk fat of goats fed with unprotected CLA	Gulati et al. (2000)
CLA	Reduces milk fat yield	CLA protection from biohydroge-nation	NR	Lipidic coating	NR	138 g/day	Reduction of milk fat yield of about the 22%	Perfield et al. (2004)
Fish oil rich in eicosapentaenoic acid and docosahe-xaenoic acid	Increase unsaturated fatty acids	eggs fatty protection from the oxidation and fishy odors and flavors mas-king	NR	NR	NR	60 g/kg/day	Increase of 40-fold for the eicosapentaenoic acid and 2.6-fold for docosahexa-enoic acid feeding the la-ying hens with microen-capsulated fish oil	Lawlor et al. (2010)

4 NR= commercial product/blend; information not reported

Table 5. Encapsulated organic acids with potential in animal feed

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Technique of encapsulation	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
Citric acid, sorbic acid, thymol, vanillin	Increase the growth performance	Allows a slow release	NR	NR	NR	0.5 % of DM	The addition of microencapsulated organic acid and essential oils to the chicken diet bring to a significant reduction in intramuscular fat content and an overall improvement in fatty acid profile. In addition the reduction of the lipid oxidation in meat was observed	Stamilla et al. (2020)
Citric and sorbic acids	Increase the growth performance	Allows a slow release	NR	Hydrogenated vegetable lipids	NR	3 kg/ton/day	As well as exert an antimicrobial effect, microencapsulated organic acids significantly affect the growth performance due to a higher pig's feed intake throughout the study (+ 4.6%)	Grilli et al. (2010)
Citric acid, sorbic acid, thymol, vanillin	Increase the growth performance	Allows a slow release	NR	NR	NR	0.075 % of DM	The addition of microencapsulated organic acid and essential oils to the diet of broiler chickens improved their growth performance and decreased drip loss percentage	Gheisar et al. (2015)
Citric acid, sorbic acid, thymol, vanillin	Increase the growth performance	Allows a slow release	NR	NR	NR	0.05 % of DM	The addition of microencapsulated organic acid and essential oils to the diet of pigs significantly affect the growth performance	Cho et al. (2014)

6 NR= commercial product/blend; information not reported

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(continued)

Table 5. Continued

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Technique of encapsulation	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
Formic, phosphoric, lactic, acetic, butyric, and propionic acid	Increase the growth performance	Reduces the lipid peroxidation	NR	NR	NR	1-3 kg/ton/day	The addition of microencapsulated organic acid to the broilers significantly increased antioxidant levels and bring to a reduction of lipid peroxidation	Galli et al. (2020a)

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9 NR= commercial product/blend; information not reported

Table 6. Encapsulated essential oils and plant extract with potential in animal feed

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Technique of encapsulation	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
Carvacrol, thymol and cinnamaldehyde	Increase the growth performance	Reduces the lipid peroxidation	NR	NR	NR	1000 mg/kg/day	The supplementation with microencapsulated essential oil in lamb stimulated antioxidant and anti-inflammatory responses and consequently increased weight gain in lamb	Favaretto et al. (2020)
Thymol, cinnamaldehyde and carvacrol	Increase the growth performance	Stabilizes essential oils characterized by high reactivity	NR	NR	NR	100 mg/g/day	The addition of microencapsulated essential oils to broiler chicken diet significantly reduces total saturated fatty acid levels	Galli et al. (2020b)
Oregano, cinnamon, thyme and orange-peel essential oils	Increase the growth performance	Reduces the lipid peroxidation	NR	NR	NR	120 g/day	The addition of microencapsulated essential oil on dairy cows increased milk protein and fat concentration	Spanghero et al. (2009)
Cinnamon oil and sodium butyrate	Increase the growth performance	Reduces the lipid peroxidation	NR	Vegetable fatty acids	NR	250 mg/kg/day	Cinnamon microencapsulated significantly reduced the meat cholesterol of broiler chickens	Gomathi et al. (2018)

11 NR= commercial product/blend; information not reported
12 (*continued*)

Table 6. Continued

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Technique of encapsulation	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
Coriander oil, geranylacetate, and eugenol	Modify milk characteristics	Reduce the lipid peroxidation	NR	NR	NR	1.66 g/ kg DM	The addition of microencapsulated essential oil had no effect on milk fat and milk protein	Guasch et al. (2006)
Aloe vera, dill and nettle roots extracts	Improve animal body weight	Overcome the extract limitations related to the hydrophobic, highly active, reactive and volatile properties of bioactive compounds in the extracts	Ionic gelation	Chitosan	NR	0.02, 0.025 and 0.05 %	The addition of microencapsulated plant extract significantly improved the boiler chickens body weight gain	Meimandipour et al. (2017)
Garlic and <i>Phyllanthus niruri</i> L. mixture	Improve animal body weight	Overcome the extract limitations related to the hydrophobic, highly active, reactive and volatile properties of bioactive compounds in the extracts	NR	Arabic gum	NR	0.4%	The addition of microencapsulated plant extract had promising effects on live weight gain of broiler	Natsir et al. (2013)
Garlic extract	Improve meat quality	Overcome the extract limitations related to the hydrophobic, highly active, reactive and volatile properties of bioactive compounds in the extracts	Freeze dryer	Maltodextrine and arabic gum	NR	0.5%	Rainbow trout fed with microencapsulated garlic extract had higher protein and lower lipid content compared to the fish fed with a control diet	Adineh et al. (2020)

NR= commercial product/blend; information not reported

(continued)

Table 6. Continued

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Technique of encapsulation	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
<i>Acacia mearnsii</i> extract	Increase the PUFA in milk	Masks polyphenols taste, retaining their structural integrity until consumption, increase their bioavailability, and release them precisely at the target site	Freeze dryer	Maltodextrine and arabic gum or native starch	From 25 to 40 µm	No <i>in vivo</i> study	Starch and maltodextrin-arabic gum were successfully used in tannin extract encapsulation. Microparticles were smaller and homogenous	Adejoro et al. (2019a)
Roselle calyx extract (<i>Hibiscus sabdariffa</i>)	Improve skin pigmentation in fish	Masks polyphenols taste	Spray dryer	Maltodextrine and sucrose	From 1.0 to 16.0 µm	150, 300, and 450 mg anthocyanin/kg of DM	It was observed that fish skin color increased, while the supplementation of microencapsulated anthocyanins increased	Vanegas-Espinoza et al. (2019)
Quebracho extract	Increase the PUFA in milk	Masks polyphenols taste, retaining their structural integrity until consumption, increase their bioavailability, and release them precisely at the target site	Spray dryer	Maltodextrine and arabic gum	< 100 µm.	No <i>in vivo</i> study	Maltodextrin-arabic gum were successfully used in tannin extract encapsulation. Microparticles were smaller and a greater release occurred at the ruminal level	Tolve et al. (2021)

Table 7. Encapsulated probiotics with potential in animal feed

Bioactive compound	Putative beneficial biological effects	Potential advantage of encapsulation	Technique of encapsulation	Shell material	Particle size	Dose	Key Findings as Reported by Authors	Reference
Lactic acid and citric acid, <i>Allium sativum</i> and <i>Phyllanthus niruri</i> and <i>Bacillus coagulans</i>	Improve the performances, population of microflora, white blood cells and cholesterol content	Improves stability during processing and transport and storage. Protection of pro-biotics during digestion	Microwave oven drying	Arabic gum and whey protein	NR	1.5%	The microencapsulated form of the mixture provided better results, in terms of body weight gain, than a non-encapsulated form	Natsir et al. (2019)
<i>Lactobacillus fermentum</i> , <i>Lactobacillus acidophilus</i> , <i>Bacillus spp</i>	Improve the performances, population of microflora, white blood cells and cholesterol content	Improve stability during processing and storage. Protect probiotics during digestion	NR	Arabic gum and whey protein	NR	0.2-0.4%	No significant effect of encapsulated probiotic on relative carcass weight, giblet weight, and intestinal morphometry of duck	Ardiansah et al. (2020)
Inulin	Positive effect on the productivity Antibacterial effect	Inulin is vulnerable to the air and has a low transit rate in caecum	Freeze dryer	Sureteric®	NR	200-250 g/ton	Significantly higher breast weights and thigh muscles and lower abdominal fat in broiler chickens fed with microencapsulated inulin compared to the control	Park and Park (2011)

Table 8. Five freedoms of animal welfare and their relation to nutrition alongside potential benefits of encapsulated ingredients-based feed with specific example (adapted from Phillips (2016) and FAWC, 2014).

Freedom	Relation to nutrition and environmental stress	Example of potential effect of feeds containing encapsulated bioactive ingredients
1. Freedom from thirst, hunger and malnutrition	Short-term malnutrition can result in the emotions of hunger in animals but long-term consequences of malnutrition are uncertain	Encapsulation results in increased nutrients bioavailability, thus reducing the risk of malnutrition at equivalent nutrients in the feed Fat coated cysteamine to mask its strong smell which may affect feed palatability and intake
2. Freedom from discomfort and exposure	Many consequences of unbalanced diet are likely to lead to discomfort such as bloating; acidosis, ketosis, deficiencies and toxicities. Some diseases causing discomfort have many predisposing factors including malnutrition	Encapsulation with slow-release of bioactive compounds to prevent their immediate absorption and reduce potential risk of toxicity. Increase of resistance to thermal stress with encapsulated vitamin B in the feed
3. Freedom from pain, injury and disease	Unbalanced diet often ends in a diseased state usually accompanied by pain	Encapsulated symbiotic and natural antioxidants such as polyphenols
4. Freedom to express normal behavior (both appetitive and consummatory)	Diets are often too concentrated in energy and other essential nutrients to ease distribution to the animals and accelerate production or reproduction rates. Because of the inverse relationship between energy and fiber concentration in forages, concentrated feeds are often deficient in fiber. Livestock that evolved to consume fibrous herbage or forage for a substantial portion of the day display abnormal oral behaviors in response to highly concentrated diets that can be rapidly eaten and digested	Sustained release of nutrients, modulate microbial flora with probiotic and prebiotic to increase fiber digestibility and conversion Use encapsulated tannins and polyphenol to increase mask taste, increase appetite and animal well-being
5. Freedom from fear and distress	Responses to repeated nutritional stress or environmental stress (heat) may invoke fear responses. Farmers should avoid or limit treatment which could cause mental suffering	Sustained and targeted delivery at desirable site using feed containing encapsulated of bioactive compounds to instead of frequent injections