

Parenteral Provision of Micronutrients to Pediatric Patients: An International Expert Consensus Paper

Gil Hardy, PhD¹ ; Theodoric Wong, BMed Sc, MBBS² ;
 Hana Morrissey, BPharm, PhD³ ; Collin Anderson, PharmD, PhD⁴ ;
 Sissel J. Moltu, MD, PhD⁵ ; Brenda Poindexter, MD, MS⁶ ;
 Alexandre Lapillonne, MD, PhD⁷ ; and Patrick A. Ball, PhD, MSc(Paeds)³ 

Journal of Parenteral and Enteral Nutrition
 Volume 44 Supplement 2
 September 2020 S5–S23
 © 2020 The Authors. *Journal of Parenteral and Enteral Nutrition* published by Wiley Periodicals LLC on behalf of American Society for Parenteral and Enteral Nutrition
 DOI: 10.1002/jpen.1990
 wileyonlinelibrary.com
WILEY

Abstract

Introduction: Micronutrients (vitamins and trace elements) are essential to all nutrition. For children and neonates who are dependent upon nutrition support therapies for growth and development, the prescribed regimen must supply all essential components. This paper aims to facilitate interpretation of existing clinical guidelines into practical approaches for the provision of micronutrients in pediatric parenteral nutrition. **Methods:** An international, interdisciplinary expert panel was convened to review recent evidence-based guidelines and published literature to develop consensus-based recommendations on practical micronutrient provision in pediatric parenteral nutrition. **Results:** The guidelines and evidence have been interpreted as answers to 10 commonly asked questions around the practical principles for provision and monitoring of micronutrients in pediatric patients. **Conclusion:** Micronutrients are an essential part of all parenteral nutrition and should be included in the pediatric nutrition therapy care plan. (*JPEN J Parenter Enteral Nutr.* 2020;44(suppl S2):S5–S23)

Keywords

minerals/trace elements; neonates; parenteral nutrition; pediatrics; public policy; vitamins

Introduction

Micronutrients refers to all vitamins and trace elements (TEs) known to be essential constituents of the diet that are required to maintain fundamental metabolic functions. A lack of any of these essential components results in nutrient-specific deficiencies that can be symptomatic

and interfere with growth and development. Routine provision of micronutrients from commencement of pediatric nutrition therapy is widely recommended but is far from universal practice. The underlying condition of the patient may result in specific requirements for individual micronutrients.^{1,2}

From the ¹Ipanema Research Trust, Auckland, New Zealand; ²Consultant Pediatric Gastroenterologist, Women's and Children's Hospital, Birmingham, UK; ³School of Pharmacy and Pharmaceutical Science, Faculty of Science and Engineering, University of Wolverhampton, Wolverhampton, UK; ⁴Pharmacy, Intermountain Healthcare Primary Children's Hospital, Salt Lake City, Utah, USA; ⁵Department of Neonatology, Oslo University Hospital, Ullevål, Oslo, Norway; ⁶Department of Pediatrics, Cincinnati Children's Hospital and Medical Centre, Cincinnati, Ohio, USA; and the ⁷Hôpital Universitaire Necker Enfants Malades, Paris, France.

Financial disclosure: None declared.

Conflicts of interest: G. Hardy has received financial reimbursement from Baxter Healthcare for consultancy work, advisory boards, and speaking engagements. T. Wong declares no known conflict of interest. He has been funded by Shire and Mead Johnson for attending conferences with no direct financial transactions. H. Morrissey declares no known conflict of interest. C. Anderson declares no known conflicts of interest and no funding sources. S. J. Moltu reports receipt of research support (nutritional supplement) from DSM and lecture fees from Baxter. B. Poindexter declares no known conflicts of interest. A. Lapillonne received lecture fees and/or nonfinancial support from Baxter, Fresenius, Nestlé, and Mead Johnson Nutrition. P. A. Ball has received travel and accommodation for lecturing from Fresenius Kabi and Pall Corporation.

Received for publication April 2, 2020; accepted for publication August 4, 2020.

This article originally appeared online on August 7, 2020.

Corresponding Author:

Prof Patrick A. Ball, PhD, MSc(Paeds), School of Pharmacy and Pharmaceutical Science, Faculty of Science and Engineering, University of Wolverhampton, Wulfruna Street, Wolverhampton, WV1 1LY, UK.
 Email: patrick.ball@wlv.ac.uk

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.

Providing micronutrients to patients in a parenteral nutrition (PN) regimen requires all nutrients to be in a form suitable for administration via the parenteral route.^{3,4} Mixing everything into a single PN admixture simplifies administration at the bedside but presents the challenge of physical and chemical compatibility and stability.

Incorporating >50 different chemical species, some in an emulsified form, in the same container means that the majority of admixtures have limited shelf-life, usually not more than 1–7 days' stability.^{3–5} PN products marketed by different manufacturers may differ markedly in their composition.^{6,7} Mixing products from different manufacturers and suppliers without supporting stability data is not advisable.⁸

Monitoring micronutrient status is complex; whereas an absence of water-soluble vitamins may produce visible symptoms such as Wernicke's encephalopathy within 2–3 days, for many other micronutrients the deficiency picture is more complex and slower to develop. For many patients receiving PN, there will be concurrent enteral nutrient intake, moderating the need for parenteral micronutrient supplementation, although the extent of absorption may vary. For TEs, monitoring is complex but nevertheless important to avoid potential toxicity, notably in, for example, patients with liver disease and patients receiving long-term parenteral support.^{2,7} Furthermore, it is known that significant but irregular amounts of TEs are present as contaminants of the other PN products that could influence daily dosage.⁹

National and international nutrition societies have produced guidelines to assist those responsible for prescribing PN to children and neonates and reduce variations in practice.^{10–13} However, these guidelines are large and complex documents, often covering oral, enteral, and parenteral requirements. The availability of products and the skill mix and experience of local staff and the facilities at their disposal vary widely, often making it difficult to precisely follow the official guidelines. The aim of this paper is to bridge the gap between evidence-based guidelines and practical application of PN at the bedside by providing recommendations based upon our expert consensus interpretation of published guidelines together with links back to the evidence. It aims to provide advice on what a PN regimen should provide and assist users to recognize when to seek region-specific advice from national and regional centers of excellence. The provision of micronutrients in adults was similarly interpreted recently.¹⁴

The micronutrients addressed are fat-soluble vitamins (A, D, E, K), water-soluble vitamins (B and C), and TEs copper (Cu), iodine (I), iron (Fe), selenium (Se), zinc (Zn), chromium (Cr), manganese (Mn), and molybdenum (Mo).

Methodology

An interdisciplinary panel was convened representing clinical, pharmaceutical, laboratory, and academic input with a focus on practical experience in provision of nutrition therapy in neonatal and pediatric patients. Working from existing published guidelines and evidence-based publications, supplemented by searching databases such as Medline and Science Direct and personal resources, the group has attempted to answer 10 common questions to guide clinical practice in a range of settings. The sections were allocated to individual members of the team to prepare a working draft to be circulated for discussion among the group.

Terminology and Metrology

The term "supplementation" is used for the delivery of micronutrients to cover basal needs when PN administration aims at restoring deficiencies and ongoing losses, or when the aim is to achieve supranormal levels, including pharmaconutrition. Dietary recommended intakes (DRIs), will be used to indicate recommended dosages. With no international agreement on micronutrient Units, posology will be expressed in both microgram and micromol.¹⁵ Ten common questions for practical consideration regarding the use of micronutrients in pediatric PN are:

1. Which are the essential micronutrients for neonatal and pediatric patients?
2. When and under what conditions are parenteral micronutrients indicated?
3. Which micronutrients are important and when are they required for neonatal and pediatric critically ill patients?
4. Which micronutrients are important and when are they required for neonatal and pediatric burns patients?
5. Which micronutrients are important and when are they required for neonatal and pediatric surgical patients?
6. Which micronutrients are important and when are they required for pediatric home PN (HPN) patients?
7. What are the practical considerations when providing micronutrients parenterally?
8. How and when should micronutrient status in neonatal and pediatric patients be assessed/monitored?
9. What are the clinical risks of providing micronutrients to neonatal and pediatric patients?
10. What are the recommendations for providing micronutrients to neonatal and pediatric patients when suitable products are unavailable?

Table 1. Recommended Doses for Parenteral Supply of Fat-Soluble and Water-Soluble Vitamins for Preterm Infants, Infants, and Children.

Vitamins	Preterm infant <37 weeks' gestation	Infant >12 months old	Children/adolescents 1–16 years old
Vitamin A	700–1500 IU/kg/d	500–1000 IU/kg/d	500 IU/d
Thiamin B ₁	0.35–0.5 mg/kg/d	0.35–0.5 mg/kg/d	1.2 mg/d
Riboflavin	0.15–0.2 mg/kg/d	0.15–0.2 mg/kg/d	1.4 mg/d
Pyridoxine	0.15–0.2 mg/kg/d	0.15–0.2 mg/kg/d	1.0 mg/kg/d
Niacin	4–6.8 mg/kg/d	4–6.8 mg/kg/d	17 mg/d
Vitamin B ₁₂	0.3 µg/kg/d	0.3 µg/kg/d	1 µg/d
Pantothenic acid	2.5 mg/kg/d	2.5 mg/kg/d	5 mg/d
Biotin	5–8 µg/kg/d	5–8 µg/kg/d	20 µg/d
Folic acid	56 µg/kg/d	56 µg/kg/d	140 µg/d
Vitamin C	15–25 mg/kg/d	15–25 mg/kg/d	80 mg/d
Vitamin D	200–1000 IU/d or 80–400 IU/kg/d	400 IU/d or 40–150 IU/kg/d	400–600 IU/d
Vitamin E	2.8–3.5 mg/kg/d	2.8–3.5 mg/kg/d	11 mg/d
Vitamin K	10 µg/kg/d	10 µg/kg/d	200 µg/d

Modified with permission from ESPGHAN 2018 recommendations by Bronsky et al.¹

Table 2. Suggested Parenteral Trace Element Recommendations.

Trace element	Maintenance daily PN dose, µg/kg (µmol/kg)		
	Preterm infants	Infants and children	Maximum per day
Chromium	–	0.2 (0.004)	0.5 (0.1)
Copper	40 (0.6)	20 (0.3)	500 (10)
Iodine	1–10 (0.01–0.1)	1.0 (0.01)	
Iron	200–250 (3.6–4.5)	50–100 (0.9–1.8)	5000 (89)
Manganese	1.0 (0.02)	1.0 (0.02)	50 (1.0)
Molybdenum	1.0 (0.012)	0.25 (0.003)	5.0 (0.06)
Selenium	7.0 (0.09)	2–3 (0.03–0.04)	100 (1.3)
Zinc	400–500 (6.2–7.8)	50–250 (0.78–3.8)	5000 (78)

Modified with permission from ESPGHAN 2018 recommendations by Domellöf.²

Summary of Recommendations

Q1. Which Are the Essential Micronutrients for Neonatal and Pediatric Patients?

1. As micronutrients are essential for optimal human growth and development, daily provision should be an integral part of any PN therapy.
2. The appropriate route of micronutrient administration should be determined in an initial comprehensive patient assessment and included within an interdisciplinary nutrition care plan.
3. Recommended daily vitamin and TE requirements for PN are shown in Tables 1 and 2.
4. The underlying condition of the patient may result in specific requirements for individual micronutrients.

Q2. When and Under What Conditions Are Parenteral Micronutrients Indicated?

1. Micronutrients are indicated in all pediatric PN regimens and should be administered as early as possible and certainly not withheld for more than a few days.

Q3. Which Micronutrients Are Important and When Are They Required for Neonatal and Pediatric Critically Ill Patients?

1. Micronutrients should be an integral part of nutrition therapy in the critically ill pediatric patient to reduce oxidative stress and support immune functions, wound healing, and organ recovery.

2. If clinically indicated, micronutrient status during an ongoing inflammatory state should be assessed and interpreted based on physical examination, dietary history, and biomarkers unaffected by the acute inflammatory response.
3. Clinicians should prescribe according to the individual patient requirements to address deficiencies while avoiding toxicity.
4. The pediatric critically ill cardiac patient may require higher micronutrient supplementation, especially during long-term therapy with diuretics.
5. Significant micronutrient losses due to prolonged continuous renal replacement therapy (CRRT) in the critically ill should be replaced daily.
6. Special consideration regarding micronutrient supplementation should be given to both obese and undernourished critically ill children, particularly taking account of dietary history, ideal body weight, inflammation, and possible organ dysfunction.

Q4. Which Micronutrients Are Important and When Are They Required for Neonatal and Pediatric Burn Patients?

1. Burned neonatal and pediatric patients must be assessed as early as possible for their micronutrient needs.
2. If PN is deemed necessary, then the full range of micronutrient supplementation should be part of their ongoing nutrition management, both during hospitalization and for up to 24 months after the burn incident.
3. Vitamins A, C, and D and TEs Fe, Cu, Se, and Zn may be significantly depleted in patients with burns, necessitating supplementation with these specific micronutrients at doses greater than those provided in standard products.

Q5. Which Micronutrients Are Important and When Are They Required for Neonatal and Pediatric Surgical Patients?

1. Clinicians must be familiar with the extent of tissue resection and the remaining anatomy to help predict and manage postoperative micronutrient deficiencies.
2. Assessment of the patient's surgical history, including the length of the remaining bowel, is essential to determine whether fat-soluble vitamins are being absorbed enterally.
3. Zn, Fe, Cu, Se, and Mn status must be assessed in postoperative intestinal failure (IF).
4. The implementation of a standard nutrition therapy protocol, including micronutrients, can improve

outcomes of necrotizing enterocolitis (NEC) in very low-birth-weight (VLBW) infants.

Q6. Which Micronutrients Are Important and When Are They Required for Pediatric HPN Patients?

1. Micronutrients should be provided routinely to all pediatric patients receiving HPN.
2. Daily micronutrient requirements for HPN are listed under Q1 and Tables 1 and 2, but higher doses may be necessary when there are abnormal intestinal losses, such as post surgery.
3. Patients receiving long-term HPN need their micronutrient status monitored periodically to avoid deficiencies and/or toxicities.
4. HPN represents an extremely diverse group of patients, for whom advice from national or regional specialist centers will often be helpful, particularly with practical funding and supply issues in specific countries and regions.

Q7. What Are the Practical Considerations When Providing Micronutrients Parenterally?

1. Clinicians should be aware of the potential stability issues with nonroutine pediatric PN regimens containing higher concentrations of specific nutrients/micronutrients.
2. Fe may lead to destabilization of intravenous lipid emulsion (ILE), so all-in-one (AIO) admixtures containing Fe should be avoided unless stability information is available for the specific PN formulation.
3. Addition of multivitamins to the PN bag on the day of administration reduces the risk of degradation.
4. Protection of the PN bag from oxygen and light minimizes light-catalyzed oxidation of micronutrients and has been shown to reduce mortality in preterm infants.
5. Reliance on variable amounts of micronutrient contaminants in some PN components to provide the daily PN requirement would require monitoring to ensure adequate provision.

Q8. How and When Should Micronutrient Status Be Assessed/Monitored?

1. Assessment of blood measurements of micronutrients should be best performed in the absence of systemic inflammation and should be interpreted in the context of the clinical condition and history. Water-soluble vitamins should be assessed more frequently than fat-soluble vitamins.

2. Patients suspected to have a previous micronutrient deficiency should be initially monitored at least monthly.
3. Patients receiving stable micronutrient supplementations can have a reduced 3-monthly frequency of monitoring.
4. Measuring serum/plasma levels alone might not reflect true micronutrient status.

Q9. What Are the Clinical Risks of Providing Micronutrients to Neonatal and Pediatric Patients?

1. To avoid the risk of deficiencies, micronutrients should be included routinely in all neonatal and pediatric PN regimens
2. The risk of refeeding syndrome (RFS) can be minimized with a nutrition care plan incorporating a protocol for administering an immediate dose of thiamin, restricting energy provision, and closely monitoring electrolytes, especially serum phosphate.
3. Fe, Cu, and Mn levels should be regularly monitored, especially in patients with liver disease who are receiving long-term (or home) PN, to avoid potential toxicity.
4. In centers using PN products and other pharmaceuticals packaged in glass, aluminum levels in blood should be checked monthly.
5. Potential hypersensitivity reactions, largely due to excipients in certain vitamin preparations, should be heeded.
6. In some situations, such as in premature neonates, individual micronutrients may need to be prescribed separately, as there is a risk that standard commercial micronutrient products may provide too much or too little of the other micronutrients.

Q10. What Are the Recommendations for Providing Micronutrients When Suitable Pediatric Products Are Unavailable

1. Maintain regular access to national society and/or regulatory agency websites for updates on the supply/availability situation.
2. Evaluate the use of adult multivitamin/multi-TE products at reduced doses for pediatric PN regimens.
3. If adult multivitamins are used in neonates, products containing polysorbate 80 or 20 or propylene glycol should be avoided.
4. Administer individual micronutrient parenteral additives, especially the key vitamins—thiamin, folic acid, and pyridoxine—that are required daily.
5. Consider using oral/enteral micronutrient alternative products when clinically possible.

6. Increase monitoring and awareness of micronutrient deficiencies.
7. Document all adverse reactions related to shortages or unavailability of pediatric products.

Q1. Why Are Micronutrients Important for PN and When Should They Be Provided?

Answer 1: Micronutrients are essential for optimal human growth, health, and development, necessitating daily supplementation in PN regimens. Depletion can affect immune status, lead to organ dysfunction and muscle weakness, or impair wound healing.

Recommendations

1. As micronutrients are essential for optimal human growth and development, daily provision should be an integral part of any PN therapy.
2. The appropriate route of micronutrient administration should be determined in an initial comprehensive patient assessment and included within an interdisciplinary nutrition care plan.
3. Recommended daily vitamin and trace element requirements for PN are shown in Tables 1 and 2.
4. The underlying condition of the patient may result in specific requirements for individual micronutrients.

Rationale

Micronutrients are essential to life. They function as important coenzymes and cofactors for the metabolism of macronutrients and are usually obtained through the diet.¹⁴ Provision must be appropriate to the life stage and clinical requirement of the patient and should be part of any nutrition intervention from commencement of therapy. Patients with insufficient gastrointestinal (GI) function, who are unable to maintain adequate nutrition by GI absorption, will also have micronutrient depletion. In cases of insufficient dietary intake, signs and clinical symptoms of micronutrient deficiency may manifest as functional or structural abnormalities that may be reversed by supplementation of the micronutrient.^{16,17}

Many patients will have higher demands caused by excessive losses, redistribution from circulation to tissues, abnormalities in metabolism, or inadequate GI absorption. In children, malabsorption conditions include short-bowel syndrome, autoimmune enteropathies, and congenital diarrhea, among others. Micronutrient deficiencies can deleteriously affect enzyme functions and other biochemical processes, leading to organ dysfunction, muscle weakness, poor wound healing, and altered immune status. Deficiency or excess of a single micronutrient may impact the availability and function of another.¹⁸ Since it is known that reserves of most water-soluble vitamins are minimal¹ and

little is known about tissue reserves of TEs,^{2,19} early supplementation of micronutrients seems reasonable to support their essential roles in metabolic processes. The appropriate route of administration should be determined in the initial assessment and if there is a change in the clinical state of the patient.

Micronutrient deficiency, when unrecognized, may lead to developmental delay or organ damage,²⁰ but there is insufficient research to clarify which micronutrients are critical for regular monitoring. Mild to moderate Zn deficiency, but only severe vitamin A and Fe deficiencies, appears to affect growth,²¹ whereas properly planned and delivered nutrition therapy may improve growth and weight gain.^{22,23} Screening for vitamin D deficiency is currently recommended only for individuals who present with risk factors for hypovitaminosis.²⁴

Micronutrient-enriched enteral nutrition (EN) or oral nutritional supplements are the preferred first option for the provision of micronutrients in hospital. However, when the enteral route is unavailable or inefficient, micronutrients must be administered in PN. DRIs have been developed in healthy populations, and as such, their application to acutely or chronically ill patients requiring parenteral supplementation are only estimates and should always be considered together with a comprehensive initial assessment aimed at identifying deficiencies so that preexisting malnutrition or specific requirements can be addressed.²⁵ Whatever the administration route, individual micronutrients undergo the same metabolic and elimination pathways. Nevertheless, when provided orally, they are regulated by normal physiological mechanisms, whereas the parenteral route may lead to deposition of nonphysiological levels and chemical forms in tissues.²⁶

The pan-European guidelines endorsed by the European Society for Paediatric Gastroenterology Hepatology and Nutrition (ESPGHAN)/European Society for Clinical Nutrition and Metabolism (ESPEN)/European Society for Pediatric Research (ESPR)/Chinese Society for Parenteral and Enteral Nutrition (CSPEN)²⁷ have published evidence-based recommendations concerning provision of micronutrients in PN for children. These emphasize the importance of an interdisciplinary team developing an overall “nutrition care plan” when administering PN. This should include defined goals and an estimate of the expected duration of PN, based upon assessment of vital signs, physical state, anthropometry, laboratory indices, and dietary intake.^{11,12,19} A thorough initial assessment is required to determine special needs. The consensus view is that micronutrients should be an integral part of PN from the first day of therapy.

Q2. When and Under What Conditions are Parenteral Micronutrients Indicated?

Answer 2: Regular administration of micronutrients is essential for all hospitalized pediatric patients, and when the

oral/enteral route is not available or is insufficient, then PN is indicated.

Recommendation

1. Micronutrients are indicated in all pediatric PN regimens and should be administered as early as possible and certainly not withheld for more than a few days.

Rationale

Micronutrients are generally provided by a balanced diet in neonates and infants by the normal progression from breastfeeding to mixed feeding to “normal” diet; but in patients requiring EN support and/or PN, they must be prescribed and provided by these alternative routes.^{23,28} All PN regimens should include micronutrients. Regular parenteral administration of vitamins and TEs ensures the provision of essential substrates and cofactors involved in many metabolic processes.

The assessment and prescription of micronutrients, as part of a PN regimen, should ideally be performed by an experienced interdisciplinary nutrition support team (NST),¹² including a pediatrician or neonatologist, specialist pharmacist, dietitian, nutrition nurse, and a medical laboratory scientist. The NST should also be responsible for children with IF in need of long-term PN or HPN. In some countries, care of long-term patients is further supported by one or more national “center(s) of excellence.”

In pediatrics, and especially neonatology, the nutrition needs for growth and development require a higher nutrient density in the PN admixture. Ideally, regimens should include the “conditionally essential” amino acids: cysteine and taurine²⁹ contain sufficient calcium and phosphate for skeletal development and the full range of micronutrients.²⁷ Pediatric patients require differing amounts of the TEs Cr, Cu, I, Mn, Mo, Se, and Zn, according to age and weight (see Table 2, Q1), and may be particularly susceptible to Fe deficiency, especially neonates requiring multiple frequent blood sampling or patients requiring long-term PN with minimal enteral intake. When Fe is required, it should be administered separately as an intermittent infusion or as an oral supplement.²

Q3. Which Micronutrients Are Important and When Are They Required for Pediatric Critically Ill Patients?

Answer 3: Micronutrients play important roles as free radical scavengers and in supporting immune functions and tissue repair during critical illness.

Recommendation

1. Micronutrients should be an integral part of nutrition therapy in the critically ill pediatric patient to reduce oxidative stress and support immune functions, wound healing, and organ recovery.
2. If clinically indicated, micronutrient status during an ongoing inflammatory state should be assessed and interpreted based on physical examination, dietary history, and biomarkers unaffected by the acute inflammatory response.
3. Clinicians should prescribe according to the individual patient requirements to address deficiencies while avoiding toxicity.
4. The pediatric critically ill cardiac patient may require higher micronutrient supplementation, especially during long-term therapy with diuretics.
5. Significant micronutrient losses due to prolonged CRRT in the critically ill should be replaced daily.
6. Special consideration regarding micronutrient supplementation should be given to both obese and undernourished critically ill children, particularly taking account of dietary history, ideal body weight, inflammation, and possible organ dysfunction.

Rationale

During critical illness, micronutrients play important roles as free radical scavengers and in supporting immune functions and tissue repair.³⁰ Deficiencies may increase the critically ill child's susceptibility to sepsis and ventilator support.³¹

The redistribution of micronutrients between tissues and body fluids and the reduced synthesis of carrier proteins induced by the inflammatory response syndrome result in a significant depletion of many plasma micronutrient concentrations regardless of actual body stores (Se, Zn, vitamin A, vitamin B₆, vitamin D, and vitamin C).^{32,33} This has important implications for the interpretation of individual plasma concentrations. For instance, low vitamin D concentration is associated with worse clinical outcomes in pediatric intensive care patients, but it is difficult to determine whether a low concentration reflects a true deficiency or if it is an epiphenomenon.³⁰ Thiamin and folic acid have also been reported to be reduced in congestive heart failure, sepsis, and inflammation and during oxidative stress, whereas the concentrations of the positive acute phase reactants serum ferritin and ceruloplasmin increase during critical illness, likely underestimating Fe and Cu deficiencies.³³ The interpretation of micronutrients status may further be complicated by anti-inflammatory treatment counteracting the effects of the inflammatory response syndrome. Postnatal dexamethasone administration has been shown to increase vitamin A concentrations (retinol and retinol-binding pro-

tein) in preterm infants.³³⁻³⁵ Long-term use of the diuretic furosemide may also cause acute depletion of thiamin, leading to exacerbation of congestive heart failure.³⁷

Sepsis, trauma, or multiple organ failure may induce acute renal failure (ARF).⁴⁰ The principal aim of nutrition therapy in ARF is to enhance immunocompetence and improve wound healing and organ dysfunction. Individual ARF patients' nutrition requirements can vary considerably,⁴¹ but those with underlying malnutrition will be at increased risk of micronutrient deficiencies due to decreased intake, malabsorption, increased utilization, and greater losses from CRRT because of the high fluid turnover. Prolonged CRRT can contribute to Cu deficiency, and plasma levels of Se, Zn, and most vitamins, except vitamin K, are also decreased, such that requirements will generally exceed healthy recommended dietary allowance (not daily) (RDA). Thiamin removal during CRRT may potentiate the deleterious effects of decreased thiamin levels,⁴² and a loading dose has been advocated upon intensive care unit (ICU) admission,⁴³ followed by regular intermittent infusions during CRRT while monitoring whole-blood levels. Recent evidence suggests pyridoxine and folate losses with CRRT may be greater than earlier reported, requiring higher daily supplementation and routine monitoring of serum levels in accordance with ESPEN guidelines.⁴⁴ Supplementation of vitamin C has also been proposed,⁴¹ but caution is advised because of the potential for excess ascorbate to be converted to the toxic oxalate salt.

Childhood obesity can increase the risk of mortality in the critically ill child,⁴⁵ necessitating special considerations for nutrition therapy in the pediatric ICU (PICU). Obesity is an inflammatory syndrome that results in increased blood volume, increased cardiac output, and decreased renal and/or hepatic function, which can all affect the metabolism of parenteral micronutrients. Previous dietary intake of foods with low nutrient density and bariatric surgery increase the risk of micronutrient deficiencies in the PICU, particularly vitamin D, thiamin, folate, B₁₂, and Fe.⁴⁶ PN dosing should be based on ideal body weight, but because of the complex factors that affect safe and effective administration and dosing of nutrients and other medications, consultation with a pediatric pharmacist is important.⁴⁷

Clinicians and NSTs should address micronutrient deficiencies according to the individual ICU patients' requirements and prescribe accordingly. Of note, Dao reported that supplementation of micronutrients during times of severe illness has not demonstrated clear benefit in either survival advantage or reduction of adverse outcomes.³⁰ Conversely, Berger asserted that there is evidence that a combined PN supplement of Cu, Zn, and Se can decrease the risk of nosocomial infections in the ICU.⁴⁸ Any supplementation of TEs at greater than RDA posology must be accompanied by serial monitoring of renal function and blood levels.

Table 3. Vitamin and Trace Element Requirements in Children With Burns.

Age, y	Vit A, IU	Vit D, IU	Vit E, IU	Vit C, IU	Vit K, µg	Folate, µg	Cu, mg	Fe, mg	Se, µg	Zn, mg
0–13										
Nonburned	1300–2000	600	6–16	15–50	2–60	65–300	0.2–0.7	0.3–8	15–40	2–8
Burned	2500–5000			250–500		1000 ^a	0.8–2.8		60–140	12.5–25
≥13										
Nonburned	200–3000	600	23	75–90	75–120	300–400	0.9	8–18	40–60	8–11
Burned	10,000			1000		1000 ^a	4		300–500	25–40

Vit, vitamin.

Reproduced under creative commons license (<http://creativecommons.org/licenses/by/4.0/>) from Clark et al.⁵⁰

^aAdministered 3 times weekly.

However, standard commercial fixed-formulation products have limitations.⁴⁹

Q4. Which Micronutrients Are Important and When Are They Required for Pediatric Burns Patients?

Answer 4: Micronutrients are essential to improve immune status and wound healing after burns.

Recommendation

1. Burned neonatal and pediatric patients must be assessed as early as possible for their micronutrient needs.
2. If PN is deemed necessary, then the full range of micronutrient supplementation should be part of ongoing nutrition management, both during hospitalization and for up to 24 months after the burn incident.
3. Vitamins A, C, and D and TEs Fe, Cu, Se, and Zn may be significantly depleted in burns, and nutrition management could benefit from supplementation with these specific micronutrients at doses greater than those provided in standard products.

Rationale

The metabolic response of the human body to severe burn (>40% of total body surface area [TBSA]) increases >2-fold, leading to a hypermetabolic and hyperdynamic state. In children, a significant increase in resting energy expenditure has been found to persist for up to 24 months.^{50,51} Protein loss, with concurrent loss of micronutrients and insulin resistance and an increase in liver size by up to 200%, is also reported.⁵¹

Children with burns have unique clinical and nutrition challenges—fluid and electrolytes, energy requirements, differing body proportions, TBSA to body mass ratio, rate of fluid loss, risk of hypothermia, nonshivering thermogenesis (increased metabolic rate, oxygen consumption, and lactate production), and thin skin—resulting in difficulties

assessing the depth of the burn.⁵² If nutrition therapy is commenced early, these challenges may be addressed, preventing impairment of wound healing, weight loss, and immune compromise.⁵¹ Fluid retention in children may mask loss of body mass.

Vitamins and TEs are essential from the initiation of therapy because of their importance for the immune system and wound healing process.⁵⁰ The greatly increased inflammatory response and consequent oxidative stress may exceptionally deplete a number of micronutrients, including vitamins A, C, and D (See Table 3). Vitamin A is known to improve epithelial growth, and vitamin C is known to enhance collagen production and cross-linking.⁵⁰ Burned skin is not able to manufacture vitamin D, and both calcium and vitamin D homeostasis are altered because of an increase in osteoblast apoptosis, bone resorption, and urinary calcium loss. Additionally, the TEs Fe, Cu, Se, and Zn are lost in burn wound cellular exudates.⁵⁰

Q5. Which Micronutrients Are Important and When Are They Required for Neonatal and Pediatric Surgical Patients?

Answer 5: Important micronutrient deficiencies arising from surgery may need specific supplementation.

Recommendation

1. Clinicians must be familiar with the extent of tissue resection and the remaining anatomy to help predict and manage postoperative micronutrient deficiencies.
2. Assessment of the patient's surgical history, including the length of the remaining bowel, is essential to determine whether fat-soluble vitamins are being absorbed enterally.
3. Zn, Fe, Cu, Se, and Mn status must be assessed in postoperative IF.
4. A standard nutrition therapy protocol including micronutrients can improve NEC outcome in VLBW infants.

Table 4. Association of Micronutrient Deficiencies With Intestinal Surgery.

Zone of GIT resection	Potential micronutrient deficiency
Gastric	Vitamin D ^{53,56} Vitamin K ⁵⁶ Iron ^{19,53} Vitamin B ₁₂ ^{19,53}
Gastric bypass	Vitamin K deficiency ⁵⁶ Copper ²³
Cholecystectomy	Vitamins A, D, E, and K ⁵³
Jejuno-ileal bypass	Vitamins A, D, E, and K ^{56,57} Calcium ⁵⁷
Pancreatico-duodenectomy	Vitamins A, D, E, K, and B ₁₂ and iron ⁵⁸
Proximal jejunum	Zinc ⁵⁹ and copper ⁶⁰
Terminal ileum	Vitamin B ₁₂ ^{19,53,56,57}
Extensive short bowel	Vitamins B ₂ , ⁵⁶ A, E, and K and, if colon resected, folic acid, chromium, ⁵⁶ zinc, and iron ^{19,23}

GIT, gastrointestinal tract.

Rationale

Pediatric surgical patients often respond differently than adults to the stresses of surgery. Although many of the basic principles of nutrition therapy still apply, the nature of the surgical procedures and any specific diagnoses warrant individual consideration. Postoperative complications can also impact micronutrient loss, and previously asymptomatic deficiency states may become symptomatic following surgery. Thus, as documented in adult patients, Zn and Se depletion has been associated with enterocutaneous fistulae,^{23,53} and Se deficiency with leakage of chyle.⁵⁴

Following GI surgery, an accurate clinical assessment of the patient's surgical history, including the length of the remaining bowel, is essential. The presence or absence of the ileum will determine whether long-chain fatty acids and fat-soluble vitamins A, D, E, K, and B₁₂ are being absorbed enterally. Table 4 associates the zone of GI tract resected with potential micronutrient deficiencies. Vitamin K deficiency may not be well recognized, as the international normalized ratio for prothrombin time lacks sensitivity and treatment with broad-spectrum antibiotics changes the intestinal flora, a major source of vitamin K. Irrespective of laboratory tests, the prudent clinician should consider additional parenteral vitamin K either by intramuscular injection or by addition to the PN, especially prior to elective surgery.⁵⁵ Small and highly variable amounts of vitamin K may be available from the ILE, and some of the standard multivitamin preparations provide insufficient vitamin K.

TE deficiencies are relatively common, so assessment should include status of Zn, Fe, Cu, and Se.⁶⁰ Symptoms

that may be observed are shown in Table 5. All outputs from stoma, stool, and urine must be assessed in IF, as large stool losses are associated with acidosis and micronutrient deficiencies. Ileostomy effluent particularly contains high levels of Zn.⁶¹ Serum Zn levels therefore need to be monitored and appropriate supplementation prescribed. Since evidence shows that Cu and Mn can accumulate in the liver, the PN dose of standard TE supplements is commonly halved in the presence of IF-associated liver disease, and regular monitoring is instituted to avoid excessive Cu/Mn toxicity while aiming to avoid deficiencies of the other elements.

Nutrition management of pediatric IF in multidisciplinary centers has been instrumental in improving patient outcomes, whereas treatment of NEC remains controversial, with many different practices being employed. Using a standard plan for feeding VLBW (<1500 g) infants with a regimen that included micronutrients contributed to improvements in NEC rates and infant mortality but the authors did not separately investigate any contribution from micronutrients.⁶³ Some children with NEC or gastroschisis can tolerate EN, but anorexia, feeding intolerance, and perioperative ileus can limit the effectiveness of EN and present unique nutrition challenges. Small amounts of trophic EN can be hepatoprotective for PN patients. Oral intake should therefore be encouraged as early as possible, and whenever feasible, the EN should contain contributory micronutrients for ambulatory surgical PN patients, but parenteral supply should continue until enteral feeding is well established and PN weaned. Cyclic PN can prevent hyperinsulinemia and may decrease the risk of hepatic steatosis and cholestasis in older children but is not recommended for neonates and young children.⁶⁴

Q6. Which Micronutrients Are Important and When Are They Required for Pediatric HPN Patients?

Answer 6: Micronutrient deficiencies can be high in long-term HPN. Pediatric HPN patients need daily supplementation and their status monitored regularly.

Recommendation

1. Micronutrients should be provided routinely to all pediatric patients receiving HPN.
2. Daily micronutrient requirements for HPN are listed under Q1 and Tables 1 and 2, but higher doses may be necessary when there are abnormal intestinal losses, such as post surgery.
3. Patients receiving long-term HPN need their micronutrient status monitored periodically to avoid deficiencies and/or toxicities.

4. HPN represents an extremely diverse group of patients, for whom advice from national or regional specialist centers will often be helpful, particularly with practical funding and supply issues in specific countries and regions.

Rationale

Physiological reasons for the need for micronutrients include maintenance of GI epithelial integrity and development, but micronutrients are also important for intestinal adaptation. Individual requirements may depend upon gestational age, presence of a high-output stoma, hepatic or renal dysfunction, and any enteral absorption. Abnormal losses such as from fistulae and chronic inflammation should also be considered in the individual patient requirements.

Without supplementation, micronutrient deficiencies are common among pediatric patients receiving HPN. Vitamin D, Fe, vitamin A, Zn, and Cu are among the most common deficiencies^{60,65,66} and can be as high as 90% of HPN patients.⁶⁵ Rat models showed vitamin A deficiency is associated with reduced mucosal protein and DNA content as well as alkaline phosphatase activity.⁶⁰ A nonexhaustive list of symptoms of deficiency include cardiomyopathy (Se), growth retardation and poor wound healing (Zn), leukopenia (Cu), contribution to type 2 diabetes development (Cr), anemia (Fe), goiter (I),²⁰ loss of vision (vitamin A), weak muscles (vitamin B₁), bleeding gums (vitamin C), osteopenia (vitamin D), and macrocytic anemia (vitamin B₁₂).⁴⁹

The reason for the extent of deficiencies seen can be due to a lack of appropriate provision with standard micronutrient parenteral supplements, limitations of solubility in PN, and premature weaning from micronutrient-containing PN. Commercially available standard supplements might not deliver, for example, sufficient Zn for premature infants.² Equally, when there is cholestasis or renal impairment, micronutrients such as Cu should be reduced or removed.⁶⁷

Q7. What Are the Practical Considerations When Administering Micronutrients Parenterally?

Answer 7: Practitioners should be cognizant of micronutrient contamination and consider stability/compatibility issues when devising a PN formulation containing micronutrients.

Recommendations

1. Clinicians should be aware of the potential stability issues with nonroutine pediatric PN regimens containing higher concentrations of specific nutrients/micronutrients.

2. Fe may lead to destabilization of ILEs, so AIO admixtures containing Fe should be avoided unless stability information is available for the particular PN formulation.
3. Addition of multivitamins to the PN bag on the day of administration reduces the risk of degradation.
4. Protection of the PN bag from oxygen and light minimizes light-catalyzed oxidation of micronutrients and has been shown to reduce mortality in preterm infants.
5. Reliance on variable amounts of micronutrient contaminants in some PN components to provide the daily PN requirement would require monitoring to ensure adequate provision.

Rationale

Regular daily administration of vitamins and TEs allows for the provision of essential substrates and cofactors involved in many metabolic processes. Micronutrients are usually administered as part of a PN regimen or separately, if PN is not required. Patient/caregiver convenience is usually increased when micronutrients are incorporated in the PN, thereby decreasing the number of infusions that must be managed on a daily basis. Nevertheless, it is vital that parents or caregivers are trained in aseptic techniques if tasked with making additions of multivitamins to the PN bag.

Stability issues related to vitamins and TEs guide how micronutrients should be administered. When “off the shelf” standard multitrace and/or multivitamin products are used within approved stability matrices, PN admixtures are generally safe. Institutions should employ processes to ensure the integrity of the additives during PN compounding and/or administration.

Ascorbic acid (vitamin C) exhibits significant chemical lability through oxidation within PN.⁶⁸ Light and PN packaging composition influences the rate of oxidation.⁶⁹ Various B vitamins are susceptible to breakdown, such as the chemical reduction of thiamin and photodegradation of vitamin B₂ or riboflavin (RF). RF is extremely light sensitive and an efficient photosensitizer inducing oxidative damage to light-exposed tissues, foods, and nutrients.⁷⁰ It absorbs both visible and UV light, is an efficient oxygen radical sensitizer, and is a strong oxidant in its triplet state. Consequently, UV therapy in the neonatal ICU for hyperbilirubinemia can degrade the vitamin and lead to RF deficiency.⁷¹ RF photosensitization is also responsible for oxidative degradation of protein, polyunsaturated lipids, and other vitamins such as folate, thiamin, and ascorbate. For example, *in vitro* studies have demonstrated that free radical-mediated reactions contributed to the rapid photodegradation of up to 69.0% of methionine.⁷² The S-amino acid methionine protects against hepatotoxic

agents by providing intracellular cysteine for biosynthesis of the hepatoprotective antioxidant glutathione (GSH). Thus, by degrading this important GSH substrate, RF photo-oxidation can have an inhibitory effect on amino acid uptake and may contribute to the cytotoxicity of hepatocytes. Partial protection against RF photosensitization is offered by specific nutrients such as chromanols, like vitamin E, but the most simple and effective practice is to exclude oxygen and light from the PN system. Ribeiro et al have reported that oxidation of an unprotected pediatric PN admixture decreases RF content by around 40% within 6 hours and 63% in 24 hours. With photoprotection of the PN container, residual RF remained at 99% after 72 hours at 4 °C and 95% at 25 °C.⁷³

Thus, destruction time of vitamins (and other nutrients) appears dependent on the PN formula, storage, transportation, and administration conditions such as light exposure. To maximize the integrity of the micronutrients, one approach utilizes adding vitamins to PN admixtures on the day of administration, rather than at the time of compounding, when the admixture is going to be stored at home or within an institution, for multiple days before use. The suggestion that administering micronutrients in ILE or AIO PN¹ protects the light-sensitive vitamins appears to miss the point that such practice leads inevitably to light-induced peroxidation of the lipid component. Simple light protection of the PN system is the obvious solution and is now a regulatory requirement in some countries.⁷⁷

Administration of Fe in PN has been particularly problematic because of the trivalent cation's destabilizing effect on ILE. Of all PN components, iron dextran disrupted ILE stability to the greatest extent,⁷⁸ but the more recent reformulated dextran products and alternative compounds, such as iron sucrose/saccharate, may be more stable. However, even in AIO admixtures protected by light, Fe supplementation in the presence of vitamin C causes reduction of ferric iron to ferrous iron, leading to an increase in lipid peroxidation, and is not recommended.⁷⁹ The stability of Fe in lipid-free 2-in-1 PN varies by formulation, with low-protein admixtures being least stable.⁷⁹ Based on rather old data, the American Society for Parenteral and Enteral Nutrition (ASPEN) suggests options for supplementation of adult PN with iron dextran or iron sucrose, but the guideline emphasizes the lack of stability evidence for supplementing pediatric PN, pointing out that many hospitalized children also receive blood transfusions, which may provide adequate Fe. Compounding iron salts in AIO PN should therefore be avoided. Administering orally or as an intermittent Fe infusion, combined with regular monitoring of Fe status, is preferred by the ESPGHAN guidelines.²

The necessity of additional provision of certain TEs, such as Cr and Mn, has been questioned because of the ubiquitous contamination of these elements within other PN components.⁸⁰ Reliance on unknown and variable

amounts of contaminants to provide these micronutrients is still being recommended but is not a defensible policy.² Since neonates may be particularly vulnerable to excessive doses of these contaminants, more research is needed to further inform the maximum allowable levels of these contaminants and their relative safety.

Q8. How and When Should Micronutrient Status Be Assessed/Monitored?

Answer 8: Preexisting deficiencies/toxicities will influence the frequency and method of monitoring.

Recommendations

1. Assessment of blood measurements of micronutrients should be best performed in the absence of systemic inflammation and should be interpreted in the context of the clinical condition and history. Water-soluble vitamins should be assessed initially, more frequently than fat-soluble vitamins.
2. Patients suspected to have a previous micronutrient deficiency should be initially monitored at least monthly.
3. Patients receiving stable micronutrient supplementations can have a reduced 3-monthly frequency of monitoring.
4. Measuring serum/plasma levels alone might not reflect true micronutrient status.

Rationale

Assessment of preexisting micronutrient issues. Assessment and interpretation of micronutrient status in critically ill children is difficult and should ideally be delayed until the inflammatory state is resolved and the patient's condition has stabilized.³³

An ESPGHAN position paper on the assessment and interpretation of vitamin and TE status in sick children was published in 2020.⁸¹ It particularly emphasized that the use of a multimodal approach, including clinical examination, dietary assessment, and biomarkers, is the optimal way to ascertain the vitamin and TE status of individual patients. It is recommended that blood measurements of vitamins and TEs should be best performed in the absence of an acute inflammatory response and should be interpreted in the context of the clinical condition and history. As a consequence, it is suggested that C-reactive protein and serum albumin level should be measured alongside plasma vitamin and TE concentrations, particularly where the disease state may result in a systemic inflammatory response.

The likelihood of pre-existing micronutrient deficiency needs to be ascertained by a detailed history (to include dietary, medical, and surgical). Seeing patients from geographical areas where certain micronutrients are scarce,

from socially disadvantaged backgrounds, or with certain medical conditions should alert prescribers to potential deficiencies. For example, Se deficiency in Heilongjiang province in China, folate deficiency in patients with neural tube defects, mixed or generalized low micronutrient accretion in premature infants and refugees, deficiency in fat-soluble vitamins in patients with chronic liver disease, and changes in growth pattern might also warrant investigation.³³

Below is an indicative list of scenarios where screening infants and children for vitamins and trace elements may be required, based upon Gerasimidis et al (2020).⁸¹ This is not an exhaustive list but considered typical of examples of patients at risk.

- Clinical symptoms of malabsorption or protracted vomiting
- Established malnutrition/growth failure
- Presence of multiple food allergies
- Long-term exclusion of major food groups; inherited disorders of metabolism, exclusion diets
- Presence of >15% unintentional weight loss
- Medications interacting with vitamins/trace elements, eg, folate antagonists

- Use of artificial nutrition lacking vitamins and trace elements for >2 weeks
- Pancreatic insufficiency, eg, cystic fibrosis with poor compliance on replacement therapy
- Long-term use of postpyloric feeding
- Presence of refeeding syndrome
- Major burns
- Major resection of small intestine or high-output stoma
- Severe insensible losses such as severe skin disease, eg, epidermolysis bullosa
- Severe liver disease or cholestasis

The decision to perform vitamin and trace element assessments remains at the discretion of the health professionals within the context of the clinical case.

Frequency of monitoring. Water-soluble vitamins rapidly deplete when intake is insufficient, as they are not stored in the body in significant amounts (apart from vitamin B₁₂). Deficiency can arise within 2–3 weeks of a micronutrient-deficient diet. If a deficiency is suspected, then biochemical monitoring should initially be no less than monthly. Appropriate investigations are listed in Table 6.

Table 5. Typical Symptoms Observed in Deficiency States of Micronutrients.

Fat-soluble vitamins	Symptoms
Vitamin A	Ocular manifestations: night blindness, dry eyes, poor growth, papillary hyperkeratosis, and impaired resistance to infections
Vitamin D	Rickets (enlargement of costochondral junctions, cranial bossing, persistently open anterior fontanelle, bowed legs, and epiphyseal enlargement)
Vitamin E	Hemolytic anemia in the newborn, hyporeflexia, and spinocerebellar and retinal degeneration
Vitamin K	Prolonged bleeding and hemorrhagic manifestations
Water-soluble vitamins	
Thiamin (vitamin B ₁) ^a	Peripheral neuropathy, cardiac failure, lactic acidosis
Riboflavin (vitamin B ₂)	Cheilosis, glossitis, corneal vascularization, and photophobia
Niacin	Pellagra: diarrhea, dermatitis, dementia
Pyridoxine (vitamin B ₆)	Microcytic anemia, seizures
Vitamin B ₁₂	Megaloblastic anemia, neurological changes
Folate	Megaloblastic anemia
Vitamin C ^a	Scurvy, petechial hemorrhages, bleeding gums
Trace elements	
Iron	Microcytic anemia, irritability
Zinc ^a	Hypogonadism, growth failure, diarrhea, decreased taste acuity, hair loss, and skin rash
Chromium	Glucose intolerance
Copper ^a	Neutropenia, anemia, neurological manifestations
Selenium ^a	Myalgia, cardiomyopathy

Adapted from Wong and Hardy.¹⁸

^aDeficiencies that may be more commonly apparent in the intensive care unit are shown in bold.

Table 6. Tests for Assessing/Monitoring Micronutrients in Pediatric PN Patients.^{106,107}

Micronutrient ^a	Test ^b	Comment	Reference
Vitamin A (retinol)	RBP and serum retinol levels	Plasma RBP and the serum retinol response to parenteral vitamin A is a better assessment of functional vitamin A status compared with random serum vitamin A levels alone, as its level only decreases when liver vitamin A storage is severely depleted	78, 80, 86, 94
Vitamin E (tocopherol)	Serum tocopherol level (common) Tocopherol: total lipid ratio (preferred)	Although tissue vitamin E level is the most informative for vitamin E status, serum or plasma tocopherol level is more commonly used. Because vitamin E level depends on plasma lipid concentrations the vitamin E, total lipid ratio is preferred	85
Vitamin D	25-OHD	25-OHD is considered the best biomarker in blood	85
Vitamin K (phytomenadione)	APTT and PTT	Adequacy of vitamin K-dependent clotting factors is normally used to determine status using APTT and PTT	85
Vitamin B ₁ (thiamin)	Whole-blood concentration of thiamin (excess), erythrocyte transketolase assay (deficiency), urine thiamin	As thiamin is integral to carbohydrate metabolism, those infants with lactic acidosis receiving high quantities of glucose would be suspected of having thiamin deficiency	85
Vitamin B ₂ (riboflavin)	Vitamin B ₂ activation coefficient	Erythrocyte glutathione reductase activity with flavin adenine dinucleotide treatment before and after is the method of choice in assessing riboflavin deficiency. Activation coefficient > 1.2 is suggestive of deficiency	85
Vitamin B ₆ (pyridoxine)	No single agreed best test	Methods include using microbiological assays, plasma pyridoxal-5-phosphate, erythrocyte aspartate, and alanine aminotransferase activity as well as a tryptophan load test	85
Vitamin B ₁₂	Serum vitamin B ₁₂	Serum vitamin B ₁₂ is most commonly used, although for functional studies, measurement of methylmalonic acid excretion is used	85
Vitamin C	Plasma and leukocyte vitamin C	Urinary measurement of niacin metabolites (<i>N</i> -methylnicotinamide and <i>N</i> -methyl-6-pyridone-3-carboxamide) is considered the best measure of niacin deficiency	85
Niacin	Urine <i>N</i> -methylnicotinamide and <i>N</i> -methyl-6-pyridone-3-carboxamide		85
Folate	Red blood cells and serum folate level	Red blood cells and serum folate level are used to assess long-term intake	85
Zinc	Serum alkaline phosphatase level, serum zinc level	There are no sensitive markers of zinc status, but serum alkaline phosphatase (marker for zinc stores) is commonly used. Results can be affected by infection and stress as well as by exercise	85
Copper	Serum copper and ceruloplasmin levels	Marginal deficiency might be normal when using serum copper and ceruloplasmin levels. These may also be raised in inflammation	77, 85

(continued)

Table 6. (continued)

Micronutrient ^a	Test ^b	Comment	Reference
Selenium	Serum selenium (common), glutathione peroxidase levels (preferred)	Serum selenium indicates more recent selenium intakes whereas erythrocyte concentration is a marker for long-term (120-day) intake. For functional status, glutathione peroxidase is now preferred	85
Iodine	Urine iodine (common), serum thyroxine level, or TSH levels (surrogate)	Iodine excretion in urine is the best method for iodine status determination, but this might not be available in most centers. Surrogate measurements of serum thyroxine level or TSH levels can be used	85
Chromium	Serum levels	Chromium is present in small quantities in the body and it is extremely difficult to measure. Serum levels may not reflect body stores. Chromium can be also measured in the hair	77, 85, 87
Manganese	Serum levels (for deficiencies), whole blood, or urine (for suspected toxicity)	Serum levels should be monitored on long-term PN, but contamination of samples is problematic and levels might not reflect nutrition status. If Mn toxicity is suspected, then an MRI scan is recommended	2, 85
Iron	As part of iron study test	Ferritin and hemoglobin status should be monitored regularly in long-term PN	2, 85

25-OHD, 25-hydroxyvitamin D; APTT, activated partial thromboplastin time; MRI, magnetic resonance imaging; PN, parenteral nutrition; PTT, prothrombin time; RBP, retinol-binding protein; TSH, thyroid stimulating hormone.

^aRed flags for possible deficiencies: malnutrition/malabsorption, complex drug therapy, complex comorbidities, high physiological demands due to infection, surgical procedures, blood loss, or severe burns.

^bFrequencies of testing: normal pediatric patient if deficiency or toxicity symptoms clinically manifested; pediatric patient with nutrition support—every clinical nutrition review; sick or very sick pediatric patient if deficiency or toxicity symptoms clinically manifested.

Fat-soluble vitamins, on the other hand, can be measured on a 6- to 12-weekly basis^{80,83} in patients receiving long-term PN.

All patients receiving long-term PN should have their TE status monitored regularly.^{1,84}

In centers using PN products packaged in glass, aluminum blood levels of pediatric PN patients should be checked monthly (see Q9 and Answer 9)

Limitations of biochemical assessment. Clinicians need to be aware that plasma/serum measurements might not be appropriate for all micronutrients (see below). Note also that inflammation might change micronutrient levels, and pairing with C-reactive protein is recommended. Not all micronutrients are routinely measured, and only those that are commonly measured are listed.

Q9. What Are the Clinical Risks of Providing Micronutrients to Neonatal and Pediatric Patients

Answer 9: The greatest risk is not providing micronutrients in a PN regimen.

Recommendations

1. To avoid the risk of deficiencies, micronutrients should be included routinely in all neonatal and pediatric PN regimens.
2. The risk of RFS can be minimized with a nutrition care plan incorporating a protocol for administering an immediate dose of thiamin, restricting energy provision, and closely monitoring electrolytes, especially serum phosphate.
3. Fe, Cu, and Mn levels should be regularly monitored, especially in patients with liver disease receiving long-term PN or HPN, to avoid potential toxicity.
4. In centers using PN products and other pharmaceuticals packaged in glass, aluminum levels in blood should be checked monthly and maintained at <5 µg/kg/d.
5. Potential hypersensitivity reactions, largely due to excipients in certain vitamin preparations, should be heeded.
6. In some situations, such as premature neonates, individual micronutrients may need to be prescribed

separately, as there is a risk that standard commercial micronutrient products may provide too much or too little of the other micronutrients.

Rationale

Deficiency of water-soluble vitamins can occur within days of commencing nonsupplemented PN, and stores of other micronutrients in pediatric patients are negligible. The greatest risk in PN therefore appears to be not providing micronutrients routinely.

RFS, caused by overenthusiastic use of nutrition support after a period of starvation, is potentially life-threatening, especially in the critically ill, but unfortunately there is no standard definition for assessment. Nevertheless, routine laboratory markers for hypophosphatemia, hypokalemia, and hypomagnesemia, plus clinical symptoms, indicate a risk of cardiac, pulmonary, or GI complications and/or organ failure. RFS and its management was reviewed by Boateng,⁸⁸ and a pediatric focus was provided by Dunn and Fuentebella.^{89,90}

A full assessment as part of a multidisciplinary nutrition care plan prior to commencement of nutrition support, with a protocol for regular monitoring of patients who develop RFS, can potentially decrease complications and overall mortality. Therapeutically, an immediate dose of 100 mg thiamin^{13,91} and thereafter maintenance doses, together with close monitoring of calcium, magnesium, potassium, and especially serum phosphate, minimizes the risk. However, recent trials among critically ill patients suggest supplementation of electrolytes and vitamins alone is insufficient. Energy restriction for several days and subsequent gradual increase of energy intake is now recommended.⁹²

Fe is an essential micronutrient but is not currently included in most pediatric TE products. However, the symptoms of “anemia of inflammation” such as lethargy and tachycardia, due to the inflammatory response and/or Fe deficiency, are relatively common in the critically ill child, leading to loss of appetite and susceptibility to infection.⁹³ Consequently, monitoring of Fe status and evaluating the need for separate oral or parenteral Fe supplements need to be included in the nutrition therapy care plan. The destabilizing effects of Fe on lipid stability limit the amount that can be safely incorporated into AIO admixtures. Nevertheless, supplementation at 50–100 $\mu\text{g}/\text{kg}/\text{d}$ (0.9–1.8 $\mu\text{mol}/\text{kg}/\text{d}$), to a maximum of 5 mg/d (88 mmol/d), is recommended.² Most allergic reactions have been associated with earlier iron dextran products. More recently, reformulated product and the use of iron sucrose/saccharate appears to minimize this risk. Nevertheless, the products are not recommended for administration to children younger than 4 months, and there may still be stability issues when either product is combined with AIO admixtures.⁸⁰

The most recent ESPGHAN/ESPEN/ESPR/CSPEN guidelines provide a summary of the special

requirements and toxicity risks of excessive micronutrient supplementation.^{1,2} Cu and Mn require additional monitoring in patients with hepatic failure and/or cholestasis, as they are known to be excreted in bile, and levels higher than normal are potentially toxic.⁶⁷ Previous recommendations to remove Cu from PN in cholestasis, however, have been rescinded.^{94–97} Current guidelines recommend 20–40 $\mu\text{g}/\text{kg}/\text{d}$ (0.3–0.6 $\mu\text{mol}/\text{kg}/\text{d}$) with a maximum of 500 $\mu\text{g}/\text{d}$ (7.5 $\mu\text{mol}/\text{d}$).² In long-term PN, there have been reports of Mn accumulation and toxicity, from deposition in the basal ganglia of the brain⁹⁸ and neuropsychiatric symptoms.⁹⁹ These occurrences are reported to be reversible over time, upon removing Mn from the PN, but this is difficult when using a fixed-formula multi-TE product. With only very few published case reports of PN-related Mn deficiency, the need to routinely supplement PN has been questioned.¹⁰⁰ Nevertheless, the lower Mn content in some of the recently reformulated commercial products⁹⁶ has partially addressed these risk concerns at a recommended dose of no greater than 1 $\mu\text{g}/\text{kg}/\text{d}$ (0.018 $\mu\text{mol}/\text{kg}/\text{d}$), to a maximum of 50 $\mu\text{g}/\text{d}$ (0.9 $\mu\text{mol}/\text{d}$).²

The TE aluminum has no significant therapeutic benefits but is present as a contaminant in some PN additives and can accumulate to toxic levels when the GI tract is bypassed, as in the case of neonates and patients with renal impairment.¹⁰¹ Aluminum accumulation has been associated with neurotoxicity, metabolic bone disease, and Alzheimer disease in patients receiving long-term PN/HPN. PN products and other potential drug additives packaged in glass containers are more susceptible to aluminum contamination, notably, calcium gluconate, inorganic phosphates and cysteine hydrochloride, albumin, and sucralfate.¹⁰²

The US Food and Drug Administration (FDA) mandate in 1986 restricting the aluminum content of large-volume PN products to 25 $\mu\text{g}/\text{L}$ was modified in 2000 to require small-volume PN additives to be labeled with their maximum aluminum concentration and that PN patient levels should not exceed 5 $\mu\text{g}/\text{L}$ (0.2 $\mu\text{mol}/\text{kg}/\text{d}$).¹⁰³ The new British Pharmacopoeia monograph for parenteral solutions (volumes unspecified) is proposing a similar limit of 25 $\mu\text{g}/\text{L}$. Until all manufacturers have significantly reduced the aluminum content of their products, it is recommended that in centers using PN products packaged in glass, blood levels of pediatric patients receiving long-term PN should be checked monthly.

Although rare, there have been case reports of hypersensitivity reactions believed to be due to surfactant excipients in certain, but not all, fat-soluble vitamin products. Acute adverse reactions have also been reported from rapid administration of large doses of some vitamins.¹⁰⁴

Even though currently available commercial TEs and vitamin products are convenient PN additives, beware that they might not meet the nutrition requirements of some

pediatric populations (for example, Zn requirements for premature neonates).²

When only one micronutrient is deficient, it may not be appropriate to increase the total volume of admixture to achieve requirement, as this will increase the provision of all other micronutrient components in the admixture. The opposite is true when one micronutrient is in the toxic range. Micronutrient components might need to be added individually rather than as premixed products.

Q10. What Are the Recommendations for Providing Micronutrients to Neonatal and Pediatric Patients When Suitable Products Are Unavailable?

Answer 10: Resources to assist healthcare professionals identify and manage shortages are available on the ASPEN, American Society of Health-System Pharmacists (ASHP), and FDA websites and from local regulatory authorities.

Recommendations

1. Maintain regular access to national society and/or regulatory agency websites for updates on the products' supply/availability.
2. Evaluate the use of adult multivitamin/multi-TE products at reduced doses for pediatric PN regimens.
3. If adult multivitamins are used in neonates, products containing polysorbate 80 or 20 or propylene glycol should be avoided.
4. Administer individual micronutrient parenteral additives, especially the key vitamins—thiamin, folic acid, and pyridoxine—that are required daily.
5. Consider using oral/enteral micronutrient alternative products where clinically possible.
6. Increase monitoring and awareness of micronutrient deficiencies.
7. Document all adverse reactions related to shortages or unavailability of pediatric products.

Rationale

Nonavailability of appropriate pediatric micronutrient products or persistent shortages can lead to inadequate dosing and consequently nutrient deficiencies. If specific pediatric micronutrient products are unlicensed in a particular country or are unavailable because of prolonged shortages, regulatory agencies may approve the temporary importation of alternative products.

ASPEN and other national PN and EN societies continuously monitor shortages of PN components through regular communications with the FDA, other regulatory agencies, pharmaceutical manufacturers, professional healthcare organizations, and clinicians.

Since 2016, ASPEN's Clinical Practice Nutrition Product Shortage Subcommittee has developed product-shortage recommendations to help clinicians manage PN therapy during times of product shortages. These recommendations, which are continuously updated, can be accessed via the ASPEN website (www.nutritioncare.org). In essence, it is recommended to reserve parenteral micronutrients for PN-dependent patients and reserve pediatric products for children. If no pediatric micronutrient products are available, then adult products can be considered at a pro rata reduced daily dose based on the weight of the patient. However, it is important to be aware that some adult products may contain levels of certain excipients and of aluminum, which may be toxic to neonates; some adult multivitamins contain propylene glycol and polysorbate 80 and 20 as excipients, which could be toxic in infants born at <36 weeks' gestation or under 1500 g. Other organic excipients such as glycine are not believed to be toxic. However, their use will marginally increase the total glycine content of the PN regimen. Clinical judgment must therefore balance the potential risks of micronutrient deficiencies from prescribing micronutrient-free PN against the potential toxicity from incorporating these adult components into the PN regimen.

If neither pediatric nor adult multivitamin or multi-TE products are available, then individual parenteral micronutrient products should be considered at dosages appropriate for the patient's age and weight. In particular, thiamin, ascorbic acid, pyridoxine, and folic acid should be given daily.¹⁰⁴ Switching to oral or enteral multivitamin/multi-TE supplements should also be considered if appropriate, with certain provisos.

Whichever strategy is adopted, it is important to notify patients receiving long-term PN/HPN when and how their PN formulation has been modified by incorporating alternative PN components. It is also advisable to increase the frequency of monitoring of serum or other appropriate biochemical markers for micronutrient status and to increase awareness of signs and symptoms of deficiencies. All adverse events or medication hazards related to shortages or unavailability of micronutrients should be documented.

Acknowledgments

We would like to thank Baxter International Inc. for initiating the discussion regarding the need for the document. Baxter International Inc. was not involved in the preparation of the document or in the recommendations made, nor did they provide any financial contribution toward the development of this document.

Statement of Authorship

It is asserted that all listed authors contributed to conception/design of the project; contributed to acquisition, analysis, and interpretation of the data; and contributed sections from which P. A. Ball, G. Hardy, T. Wong, and S. J. Moltu drafted the article. All authors critically revised the article; and all authors

agreed to be fully accountable for ensuring the integrity and accuracy of the work. All authors read and approved the final article.

References

- Bronsky J, Campoy C, Braegger C, et al. ESPGHAN/ESPER/ESPR/CSPEN guidelines on pediatric parenteral nutrition: Vitamins. *Clin Nutr*. 2018;37(6):2366-2378.
- Domellöf M, Szitanyi P, Simchowicz V, et al. ESPGHAN/ESPER/ESPR/CSPEN guidelines on pediatric parenteral nutrition: iron and trace minerals. *Clin Nutr*. 2018;37(6):2354-2359.
- Allwood MC, Kearney MCJ. Compatibility and stability of additives in parenteral nutrition admixtures. *Nutrition*. 1998;14(9):697-706.
- Driscoll DF. Stability and compatibility assessment techniques for total parenteral nutrition admixtures: setting the bar according to pharmacopeial standards. *Curr Opin Clin Nutr Metab Care*. 2005;8(3):297.
- Muhlebach S, Driscoll D, Hardy G. Pharmaceutical Aspects of PN Support. In: Sobotka L, eds. *Basics in Clinical Nutrition*. 4th ed. Czech Republic: Galen; 2011:373-392.
- Safe practices for parenteral nutrition formulations. national advisory group on standards and practice guidelines for parenteral nutrition. *JPEN J Parenter Enteral Nutr*. 1998;22(2):49-66.
- Ball PA. Methods of assessing stability of parenteral nutrition regimens. *Curr Opin Clin Nutr Metab Care*. 2001;4(5):345-350.
- Payne James J, Grimble, GK, Silk, DBA, , *Artificial Nutrition and Support Clinical Practice*. 2nd ed. Cambridge, UK: Cambridge University Press; 2012.1-758.
- Pluhator-Murton MM, Fedorak RN, Audette RJ, Marriage BJ, Yatscoff RW, Gramlich LM. Trace element contamination of total parenteral nutrition. 2. effect of storage duration and temperature. *JPEN J Parenter Enteral Nutr*. 1999;23(4):228-232.
- Druyan ME, Compher C, Boullata JI, et al. Clinical guidelines for the use of parenteral and enteral nutrition in adult and pediatric patients. *JPEN J Parenter Enteral Nutr*. 2012;36(1):77-80.
- Koletzko B, Goulet O, Hunt J, Krohn K, Shamir R. ESPEN/ESPGHAN Guidelines on paediatric parenteral nutrition. *J Pediatr Gastroenterol Nutr*. 2005;41(Supplement 2):S1-S87.
- Puntis JWL, Hojsak I, Ksiazek J, et al. ESPGHAN/ESPER/ESPR/CSPEN guidelines on pediatric parenteral nutrition: Organizational aspects. *Clin Nutr*. 2018;37(6):2392-2400.
- NICE (National Institute for Healthcare Excellence). Neonatal parenteral nutrition NICE guideline [NG154]. Published 2020. Accessed March 2, 2020. <https://www.nice.org.uk/guidance/NG154>
- Blaauw R, Osland E, Sriram K, et al. Parenteral provision of micronutrients to adult patients: an expert consensus paper. *JPEN J Parenter Enteral Nutr*. 2019;43(Suppl 1):S5-S23.
- Hardy G, Kleck S, Chourdakis M, Majid H. Towards an international metrology for clinical nutrition. *JPEN J Parenter Enteral Nutr*. 2019;43(3):445 (poster published online).
- Fraga CG. Relevance, essentiality and toxicity of trace elements in human health. *Mol Aspects Med*. 2005;26(4-5):235-244.
- Shenkin A. Basics in clinical nutrition: Physiological function and deficiency states of trace elements. *Clin Nutr Metabol* . 2008;3(6):e255-e258.
- Wong T, Hardy G. Micronutrient requirements in the critically ill child. In: Goday P, Nehta N, eds. *Pediatric Critical Care Nutrition*. New York: McGraw Hill; 2015:59-70.
- Osland EJ, Ali A, Nguyen T, Davis M, Gillanders L. Australasian society for parenteral and enteral nutrition (AuSPEN) adult vitamin guidelines for parenteral nutrition. *Asia Pac J Clin Nutr*. 2016;25(3):636-650.
- Stehle P, Stoffel-Wagner B, Kuhn KS. Parenteral trace element provision: recent clinical research and practical conclusions. *Eur J Clin Nutr*. 2016;70(8):886-893.
- Rivera JA, Hotz C, González-Cossío T, Neufeld L, García-Guerra A. The effect of micronutrient deficiencies on child growth: a review of results from community-based supplementation trials. *J Nutr*. 2003;133(11):4010S-4020S.
- Ayers P, Adams S, Boullata J, et al. A.S.P.E.N. parenteral nutrition safety consensus recommendations. *JPEN J Parenter Enteral Nutr*. 2014;38(3):296-333.
- Osland EJ, Ali A, Isenring E, Ball PA, Davis M, Gillanders L. Australasian society for parenteral and enteral nutrition (auspen) guidelines for supplementation of trace elements during parenteral nutrition. *Asia Pac J Clin Nutr*. 2014;23(4):545-554.
- Esposito S, Leonardi A, Lanciotti L, Cofini M, Muzi G, Penta L. Vitamin D and growth hormone in children: a review of the current scientific knowledge. *J Translat Med*. 2019;17(1):87.
- Puntis JWL. Nutritional support in neonatology. In: Sobotka L, eds. *Basics in Clinical Nutrition*. 3rd ed. Czech Republic: Galen; 2004:425-439.
- Boullata JI. Trace elements in critically ill patients. *J Infus Nurs*. 2013;36(1):16-23.
- Mihatsch W, Shamir R, van Goudoever JB, et al. ESPGHAN/ESPER/ESPR/CSPEN guidelines on pediatric parenteral nutrition: Guideline development process for the updated guidelines. *Clin Nutr*. 2018;37(6):2306-2308.
- Shenkin A. Vitamin and essential trace element recommendations during intravenous nutrition: theory and practice. *Proc Nutr Soc*. 1986;45(3):383-390.
- van Goudoever JB, Carnielli V, Darmaun D, et al. ESPGHAN/ESPER/ESPR/CSPEN guidelines on pediatric parenteral nutrition: Amino acids. *Clin Nutr*. 2018;37(6):2315-2323.
- Dao DT, Anez-Bustillos L, Cho BS, Li Z, Puder M, Gura KM. Assessment of micronutrient status in critically ill children: challenges and opportunities. *Nutrients*. 2017;9(11):1185-1211.
- Razavi Khorasani N, Moazzami B, Zahedi Tajrishi F, et al. The association between low levels of vitamin d and clinical outcomes in critically-ill children: a systematic review and meta-analysis. *Fetal Pediatr Pathol*. 2019:1-15.
- Duncan A, Talwar D, McMillan DC, Stefanowicz F, O'Reilly DSJ. Quantitative data on the magnitude of the systemic inflammatory response and its effect on micronutrient status based on plasma measurements. *Am J Clin Nutr*. 2012;95(1):64-71.
- Bronsky J, Catchpole A, Embleton N, et al. Assessment and interpretation of vitamin and trace element status in sick children. a position paper from the espghan committee in nutrition. *J Pediatr Gastroenterol Nutr*. 2020;70(6):873-881
- Georgieff MK, Mammel MC, Mills MM, Gunter EW, Johnson DE, Thompson TR. Effect of postnatal steroid administration on serum vitamin A concentrations in newborn infants with respiratory compromise. *J Pediatr*. 1989;114(2):301-304.
- Shenai JP, Kennedy KA, Chytil F, Stahlman MT. Clinical trial of vitamin A supplementation in infants susceptible to bronchopulmonary dysplasia. *J Pediatr*. 1987;111(2):269-277.
- Pathan N, Marino L. Chapter 17: Nutrition for the infant or child in the cardiac intensive care unit. In: Goday P, Mehta N, eds. *Pediatric Critical Care Nutrition*. New York: McGraw Hill; 2015:217-225.
- Lehman J. Chapter 6. Drug -Nutrient Interactions. In: Goday P, Mehta N, eds. *Pediatric Critical Care Nutrition*. New York: McGraw Hill; 2015:87-95.
- Clarke SE, Evans S, MacDonald A, Davies P, Booth IW. Randomized comparison of a nutrient-dense formula with an

- energy-supplemented formula for infants with faltering growth. *J Hum Nutr Dietet.* 2007;20(4):329-339.
39. Lapillonne A, Barbanti C, Lopera I, Moltu SJ. Nutrition of the neonate with congenital heart disease: existing evidence and practical implications. *Wld Rev Nutr Dietet.* Forthcoming 2020.
 40. Kamel AY, Dave NJ, Zhao VM, Griffith DP, Connor Jr MJ, Ziegler TR. Micronutrient alterations during continuous renal replacement therapy in critically ill adults: a Retrospective Study. *Nutr Clin Pract.* 2018;33(3):439-446.
 41. Druml W, Cano N, Teplan V. Nutritional Support in Renal Disease. In: Sobotka L, eds. *Basics in Clinical Nutrition.* 4th ed. Czech Republic: Galen; 2011:479-481.
 42. Manzanares W, Hardy G. Thiamine supplementation in the critically ill. *Curr Opin Clin Nutr Metab Care.* 2011;14(6):610-617.
 43. Cruickshank AM, Telfer ABM, Shenkin A. Thiamine deficiency in the critically ill. *Intensive Care Med.* 1988;14(4):384-387.
 44. Cano NJM, Aparicio M, Brunori G, et al. ESPEN Guidelines on parenteral nutrition: adult renal failure. *Clin Nutr.* 2009;28(4):401-414.
 45. Bechard LJ, Rothpletz-Puglia P, Touger-Decker R, Duggan C, Mehta NM. Influence of obesity on clinical outcomes in hospitalized children: a systematic review. *JAMA Pediatr.* 2013;167(5):476-482.
 46. Hsia DS, Fallon SC, Brandt ML. Adolescent bariatric surgery. *Arch Pediatr Adolesc Med.* 2012;166(8):757-766.
 47. Okada C, Skillman H, Krebs N. Chapter 21: nutrition in the critically ill obese child. In: Praveen SG, Nilesh MM, eds. *Pediatric Critical Care Nutrition.* New York: McGraw Hill; 269-286.
 48. Berger MM, Eggmann P, Heyland DK, et al. Reduction of nosocomial pneumonia after major burns by trace element supplementation: aggregation of two randomised trials. *Crit Care.* 2006;10(6):R153.
 49. Wong T. Parenteral trace elements in children: clinical aspects and dosage recommendations. *Curr Opin Clin Nutr Metab Care.* 2012;15(6):649-656.
 50. Clark A, Imran J, Madni T, Wolf SE. Nutrition and metabolism in burn patients. *Burns Trauma.* 2017;5:11.
 51. Rodriguez NA, Jeschke MG, Williams FN, Kamolz L, Herndon DN. Nutrition in burns: Galveston contributions. *JPEN J Parenter Enteral Nutr.* 2011;35(6):704-714.
 52. Natarajan M, Sekhar DR. Nutrition in burns patients. *IOSR J Dent Med Sci.* 2015;3(7):38-54.
 53. Sriram K, Lonchyna VA. Micronutrient supplementation in adult nutrition therapy: practical considerations. *JPEN J Parenter Enteral Nutr.* 2009;33(5):548-562.
 54. de Berranger E, Colinet S, Michaud L, et al. Severe selenium deficiency secondary to chylous loss. *JPEN J Parenter Enteral Nutr.* 2006;30(2):173-174.
 55. Shearer MJ. Vitamin K in Parenteral Nutrition. *Gastroenterol.* 2009;137(5):S105-S118.
 56. Vanek VW, Borum P, Buchman A, et al. A.S.P.E.N. position paper. *Nutr Clin Pract.* 2012;27(4):440-491.
 57. Titus R, Kastenmeier A, Otterson MF. Consequences of gastrointestinal surgery on drug absorption. *Nutr Clin Pract.* 2013;28(4):429-436.
 58. Pappas S, Krzywdka E, McDowell N. Nutrition and pancreaticoduodenectomy. *Nutr Clin Pract.* 2010;25(3):234-243.
 59. Parrott J, Frank L, Rabena R, Craggs-Dino L, Isom KA, Greiman L. American society for metabolic and bariatric surgery integrated health nutritional guidelines for the surgical weight loss patient 2016 update: micronutrients. *Surg Obes Relat Dis.* 2017;13(5):727-741.
 60. Yarandi SS, Griffith DP, Sharma R, Mohan A, Zhao VM, Ziegler TR. Optic neuropathy, myelopathy, anemia, and neutropenia caused by acquired copper deficiency after gastric bypass surgery. *J Clin Gastroenterol.* 2014;48(10):862-865.
 61. D'Aniello R, Terquem EL, Poupon J, et al. Parenteral zinc intake in newborns with jejunostomy or ileostomy: results of a monocentric Cohort Study. *J Pediatr Gastroenterol Nutr.* 2020;70(4):521-526.
 62. Yang CJ, Duro D, Zurakowski D, Lee M, Jaksic T, Duggan C. High prevalence of multiple micronutrient deficiencies in children with intestinal failure: a Longitudinal Study. *J Pediatr.* 2011;159(1):39-44.
 63. Butler TJ, Szekeley LJ, Grow JL. A standardized nutrition approach for very low birth weight neonates improves outcomes, reduces cost and is not associated with increased rates of necrotizing enterocolitis, sepsis or mortality. *J Perinatol.* 2013;33(11):851-857.
 64. Nghiem-Rao T, Cassidy L, Polzin E, Calkins C, Arca M, Goday P. Risks and benefits of prophylactic cyclic parenteral nutrition in surgical neonates. *Nutr Clin Pract.* 2013;28(6):745-752
 65. Namjoshi SS, Muradian S, Bechtold H, et al. Nutrition deficiencies in children with intestinal failure receiving chronic parenteral nutrition. *JPEN J Parenter Enteral Nutr.* 2018;42(2):427-435.
 66. Núñez-Ramos R, Gallego S, Gayda-Pimlott D, Macdonald S, Koeglmeier J, Hill S. PTH-215 Trace elements and vitamin levels in long-term home parenteral nutrition paediatric patients. *Gut.* 2015;64(Suppl 1):A504-A504.
 67. Kozeniecki M, Ludke R, Kerner J, Patterson B. Micronutrients in liver disease: roles, risk factors for deficiency, and recommendations for supplementation. *Nutr Clin Pract.* 2020;35(1):50-62.
 68. Ball PA, Barnett MI. The dissolved oxygen content of parenteral nutrition admixtures prepared by different methods. *Pharm Pharmacol Comm.* 1996;2(7):349-351.
 69. Allwood MC, Brown PW, Ghedini C, Hardy G. The stability of ascorbic acid in TPN mixtures stored in a multilayered bag. *Clin Nutr.* 1992;11(5):284-288.
 70. Cardoso DR, Libardi SH, Skibsted LH. Riboflavin as a photosensitizer. Effects on human health and food quality. *Food Funct.* 2012;3(5):487-502.
 71. Suwannasom N, Kao I, Pruß A, Georgieva R, Bäumlner H. Riboflavin: the health benefits of a forgotten natural vitamin. *Int J Mol Sci.* 2020;21(3):950-972.
 72. Remucal CK, McNeill K. Photosensitized amino acid degradation in the presence of riboflavin and its derivatives. *Environ Sci Technol.* 2011;45(12):5230-5237.
 73. Ribeiro DO, Pinto DC, Lima L, et al. Chemical stability study of vitamins thiamine, riboflavin, pyridoxine and ascorbic acid in parenteral nutrition for neonatal use. *Nutr J.* 2011;10(1):47.
 74. Billion-Rey F, Guillaumont M, Frederich A, Aulagner G. Stability of fat-soluble vitamins A (retinol palmitate), E (tocopherol acetate), and K1 (phylloquinone) in total parenteral nutrition at home. *JPEN J Parenter Enteral Nutr.* 1993;17(1):56-60.
 75. Haas C, Genzel-Boroviczény O, Koletzko B. Losses of vitamin A and E in parenteral nutrition suitable for premature infants. *Eur J Clin Nutr.* 2002;56(9):906-912.
 76. Watrobska-Swietlikowska D, MacLoughlin R. The effect of UV-protected ethylene vinyl acetate (EVA) bags on the physicochemical stability of pediatric parenteral nutrition admixtures. *Daru.* 2019;27(1):255-264.
 77. European Medicines Agency. PRAC recommendations on signals. Adopted at the 8-11 July 2019 PRAC meeting. EMA/PRAC/347675/2019. Published 2020. Accessed January 21, 2020. www.ema.europa.eu/en/human-regulatory/post-authorisation/pharmacovigilance/signal-management/prac-recommendations-safety-signals
 78. Driscoll DF, Bhargava HN, Li L, Zaim RH, Babayan VK, Bistrian BR. Physicochemical stability of total nutrient admixtures. *Am J Health Syst Pharm.* 1995;52(6):623-634.

79. Grand A, Jalabert A, Mercier G, et al. Influence of vitamins, trace elements, and iron on lipid peroxidation reactions in all-in-one admixtures for neonatal parenteral nutrition. *JPEN J Parenter Enteral Nutr.* 2011;35(4):505-510.
80. Zemrani B, McCallum Z, Bines JE. Trace element provision in parenteral nutrition in children: one size does not fit all. *Nutrients.* 2018;10(11):1819.
81. Gerasimidis K, Bronsky J, Catchpole A, et al. ESPGHAN committee on nutrition, assessment and interpretation of vitamin and trace element status in sick children: a position paper from the European Society for Paediatric Gastroenterology Hepatology, and Nutrition Committee on Nutrition. *J Pediatr Gastroenterol Nutr.* 2020;70(6):873-881.
82. Albahrani AA, Greaves RF. Fat-soluble vitamins: clinical indications and current challenges for chromatographic measurement. *Clin Biochem Rev.* 2016;37(1):27-47.
83. NICE guideline [NG49]. Non-alcoholic fatty liver disease (NAFLD): assessment and management. Published 2016. Accessed February 16, 2020. www.nice.org.uk/guidance/ng49/
84. NICE guideline [CG32] Nutrition support for adults: oral nutrition support, enteral tube feeding and parenteral nutrition. Published 2006. Accessed March 27, 2020. www.nice.org.uk/guidance/cg32
85. Sauberlich HE. *Laboratory Tests for the Assessment of Nutritional Status.* 2nd ed. New York: Routledge; 1999.
86. Tsang RC, Uauy R, Koletzko B, Zlotkin SH. *Nutrition of the Preterm Infant: Scientific Basis and Practical Guidelines.* 2nd ed. Cincinnati, OH: Digital Educational Publishing; 2005.
87. Smith A, Feuling MB, Larson-Nath C, et al. Laboratory monitoring of children on home parenteral nutrition: a Prospective Study. *JPEN J Parenter Enteral Nutr.* 2018;42(1):148-155.
88. Boateng AA, Sriram K, Meguid MM, Crook M. Refeeding syndrome: treatment considerations based on collective analysis of literature case reports. *Nutrition.* 2010;26(2):156-167.
89. Dunn RL, Stettler N, Mascarenhas MR. Refeeding syndrome in hospitalized pediatric patients. *Nutr Clin Pract.* 2003;18(4):327-332.
90. Fuentebella J, Kerner JA. Refeeding Syndrome. *Pediatr Clin N Am.* 2009;56(5):1201-1210.
91. Koletzko BG, Goulet O. Nutritional support in infants and adolescents. In: Sobotka L, eds. *Basics in Clinical Nutrition.* Czech Republic: Galen; 2011:625-652.
92. Koekkoek WAC, Van Zanten ARH. Is refeeding syndrome relevant for critically ill patients? *Curr Opin Clin Nutr Metab Care.* 2018;21(2):130-137.
93. Heming N, Montravers P, Lasocki S. Iron deficiency in critically ill patients: highlighting the role of hepcidin. *Crit Care.* 2011;15(2):210
94. Blackmer AB, Bailey E. Management of copper deficiency in cholestatic infants. *Nutr Clin Pract.* 2013;28(1):75-86.
95. Corkins M. Copper metabolism and pediatric cholestasis. *Curr Opin Clin Nutr Metab Care.* 2011;14(6):642-646.
96. Domellöf M. Nutritional care of premature infants: microminerals. *World Rev Nutr Diet.* 2014;110:121-139.
97. Frem J, Sarson Y, Sternberg T, Cole C. Copper supplementation in parenteral nutrition of cholestatic infants. *J Pediatr Gastroenterol Nutr.* 2010;50(6):650-654.
98. Uchino A, Noguchi T, Nomiya K, et al. Manganese accumulation in the brain: MR imaging. *Neuroradiology.* 2007;49(9):715-720.
99. Hardy G. Manganese in parenteral nutrition: who, when, and why should we supplement? *Gastroenterol.* 2009;137(5):S29-S35.
100. Hardy IJ, Gillanders L, Hardy G. Is manganese an essential supplement for parenteral nutrition? *Curr Opin Clin Nutr Metab Care.* 2008;11(3):289-296.
101. Gura KM. Aluminium contamination in parenteral products. *Curr Opin Clin Nutr Metab Care.* 2014;17(6):551-557.
102. Hernández-Sánchez A, Tejada-González P, Arteta-Jiménez M. Aluminium in parenteral nutrition: a systematic review. *Eur J Clin Nutr.* 2013;67(3):230-238.
103. Food and Drug Administration. Aluminum in Large and Small Volume Parenterals Used in Total Parenteral Nutrition; Delay of Effective Date. 21 CFR 201. Published January 26, 2001. Accessed February 3, 2020. <https://www.federalregister.gov/documents/2001/01/26/01-2125/aluminum-in-large-and-small-volume-parenterals-used-in-total-parenteral-nutrition-delay-of-effective>
104. Christian VJ, Tallar M, Walia CLS, Sieracki R, Goday PS. Systematic review of hypersensitivity to parenteral nutrition. *JPEN J Parenter Enteral Nutr.* 2018;42(8):1222-1229.
105. Holcombe B, Mattox T, Plogsted S. Drug shortages: effect on parenteral nutrition therapy. *Nutr Clin Pract.* 2018;33(1):53-61.
106. NICE guideline [CG160] Nutrition support for adults: oral nutrition support, enteral tube feeding and parenteral nutrition. Published 2006. Accessed March 23, 2020. www.nice.org.uk/guidance/cg160
107. United Kingdom Department of Health. Spotting the Sick Child. Accessed May 23, 2020. <https://spottingthesickchild.com/about/>